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(71) Applicant *(for all designated States except US)*: **NEOSE TECHNOLOGIES, INC.** [US/US]; 102 Witmer Road, Horsham, PA 19044 (US).

(72) Inventors; and
(75) Inventors/Applicants *(for US only)*: **DEFREES, Shawn** [US/US]; 126 Filly Drive, North Wales, PA 19454 (US). **ZOPF, David, A.** [US/US]; 560 Beechtree Lane, Wayne, PA 19087 (US). **TAUDTE, Susann** [DE/DE]; 3374 East Buck Road, Pennsburg, PA 18073 (US). **WILLETT, W., Scott** [US/US]; 3820 Comley Circle, Doylestown, PA 18901 (US). **BAYER, Robert, J.** [US/US]; 6105 Dirac Street, San Diego, CA 92122 (US). **KALO, Matthew** [US/US]; 3649 Louisiana Street, # 110, San Diego, CA 92104 (US).

(74) Agents: **ESKER, Todd, W.** et al.; Morgan Lewis & Bockius LLP, 2 Palo Alto Square, 3000 El Camino Real, Suite 700, Palo Alto, CA 94306 (US).

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(54) Title: GLYCOPEGYLATED FACTOR VII AND FACTOR VIIA

(57) Abstract: The present invention provides conjugates between Factor VII or Factor Vila peptides and PEG moieties. The conjugates are linked via an intact glycosyl linking group that is interposed between and covalently attached to the peptide and the modifying group. The conjugates are formed from both glycosylated and unglycosylated peptides by the action of a glycosyltransferase. The glycosyltransferase ligates a modified sugar moiety onto either an amino acid or glycosyl residue on the peptide. Also provided are pharmaceutical formulations including the conjugates. Methods for preparing the conjugates are also within the scope of the invention.

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PATENT APPLICATION

GLYCOPEGYLATED FACTOR VII AND FACTOR VIIA

CROSS-REFERENCES TO OTHER APPLICATIONS

[0001] The present application is related to U.S. Provisional Patent Applications 60/746,868, filed May 9, 2006; 60/756,443, filed January 5, 2006; 60/733,649, filed November 4, 2005; 60/730,607, filed October 26, 2005; 60/725,894, filed October 11, 2005; 60/709, 983, filed August 19, 2005, which is incorporated by reference in its entirety for all purposes

SUMMARY OF THE INVENTION

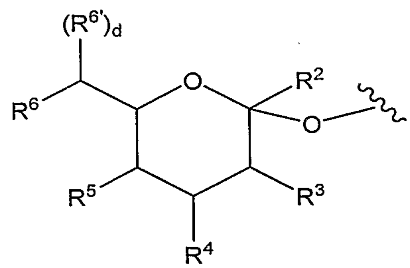
[0002] It has now been discovered that the controlled modification of Factor VII or Factor VIIa with one or more poly(ethylene glycol) moieties affords a novel Factor VII or Factor VIIa peptide conjugate with pharmacokinetic properties that are improved relative to the corresponding native (un-pegylated) Factor VII or Factor VIIa. Furthermore, cost effective methods for reliable and reproducible production of the Factor VII or Factor VIIa peptide conjugates of the invention have been discovered and developed.

[0003] In an exemplary embodiment, "glycopegylated" Factor VII or Factor VIIa molecules of the invention are produced by the enzyme mediated formation of a conjugate between a glycosylated or non-glycosylated Factor VII or Factor VIIa peptide and an enzymatically transferable saccharyl moiety that includes a modifying group, such as a polymeric modifying group such as poly(ethylene glycol), within its structure. The PEG moiety is attached to the saccharyl moiety directly (i.e., through a single group formed by the reaction of two reactive groups) or through a linker moiety, e.g., substituted or unsubstituted alkyl, substituted or unsubstituted heteroalkyl, etc.

[0004] Thus, in one aspect, the present invention provides a conjugate between a PEG moiety, *e.g.*, PEG and a peptide that has an *in vivo* activity similar or otherwise analogous to art-recognized Factor VII or Factor VIIa. In the conjugate of the invention, the PEG moiety is covalently attached to the peptide via an intact glycosyl linking group. Exemplary intact glycosyl linking groups include sialic acid moieties that are derivatized with PEG.

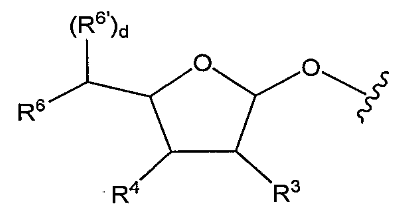
[0005] The polymeric modifying group can be attached at any position of a glycosyl moiety of Factor VII or Factor VIIa. Moreover, the polymeric modifying group can be bound to a glycosyl residue at any position in the amino acid sequence of a wild type or mutant Factor VII or Factor VIIa peptide.

[0006] In an exemplary embodiment, the invention provides an Factor VII or Factor VIIa peptide that is conjugated through a glycosyl linking group to a polymeric modifying group. Exemplary Factor VII or Factor VIIa peptide conjugates include a glycosyl linking group having a formula selected from:



I

; and



II

[0007] In Formulae I and II, R^2 is H, CH_2OR^7, $COOR^7$, COO^- or OR^7, in which R^7 represents H, substituted or unsubstituted alkyl or substituted or unsubstituted heteroalkyl. The symbols R^3, R^4, R^5, R^6 and $R^{6'}$ independently represent H, substituted or unsubstituted alkyl, OR^8, $NHC(O)R^9$. The index d is 0 or 1. R^8 and R^9 are independently selected from H, substituted or unsubstituted alkyl, substituted or unsubstituted heteroalkyl or sialic acid. At least one of R^3, R^4, R^5, R^6 or $R^{6'}$ includes the polymeric modifying group *e.g.*, PEG. In an exemplary embodiment, R^6 and $R^{6'}$, together with the carbon to which they are attached are components of the side chain of a sialyl moiety. In a further exemplary embodiment, this side chain is functionalized with the polymeric modifying group.

[0008] In an exemplary embodiment, the polymeric modifying group is bound to the glycosyl linking group, generally through a heteroatom on the glycosyl core (e.g., N, O), through a linker, L, as shown below:

$$(R^1)_w—L—\S$$

R^1 is the polymeric modifying group and L is selected from a bond and a linking group. The index w represents an integer selected from 1-6, preferably 1-3 and more preferably 1-2. Exemplary linking groups include substituted or unsubstituted alkyl, substituted or unsubstituted heteroalkyl moieties and sialic acid. An exemplary component of the linker is

an acyl moiety. Another exemplary linking group is an amino acid residue (e.g., cysteine, serine, lysine, and short oligopeptides, e.g., Lys-Lys, Lys-Lys-Lys, Cys-Lys, Ser-Lys, etc.)

[0009] When L is a bond, it is formed by reaction of a reactive functional group on a precursor of R^1 and a reactive functional group of complementary reactivity on a precursor of the glycosyl linking group. When L is a non-zero order linking group, L can be in place on the glycosyl moiety prior to reaction with the R^1 precursor. Alternatively, the precursors of R^1 and L can be incorporated into a preformed cassette that is subsequently attached to the glycosyl moiety. As set forth herein, the selection and preparation of precursors with appropriate reactive functional groups is within the ability of those skilled in the art. Moreover, coupling of the precursors proceeds by chemistry that is well understood in the art.

[0010] In an exemplary embodiment L is a linking group that is formed from an amino acid, or small peptide (e.g., 1-4 amino acid residues) providing a modified sugar in which the polymeric modifying moiety is attached through a substituted alkyl linker. Exemplary linkers include glycine, lysine, serine and cysteine. Amino acid analogs, as defined herein, are also of use as linker components. The amino acid may be modified with an additional component of a linker, e.g., alkyl, heteroalkyl, covalently attached through an acyl linkage, for example, an amide or urethane formed through an amine moiety of the amino acid residue.

[0011] In an exemplary embodiment, the glycosyl linking group has a structure according to Formula I and R^5 includes the polymeric modifying group. In another exemplary embodiment, R^5 includes both the polymeric modifying group and a linker, L, joining the polymeric modifying group to the glycosyl core. L can be a linear or branched structure. Similarly, the polymeric modifying group can be branched or linear.

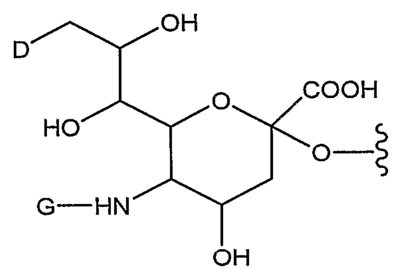
[0012] The polymeric modifying group comprises two or more repeating units that can be water-soluble or essentially insoluble in water. Exemplary water-soluble polymers of use in the compounds of the invention include PEG, e.g., m-PEG, PPG, e.g., m-PPG, polysialic acid, polyglutamate, polyaspartate, polylysine, polyethyeleneimine, biodegradable polymers (e.g., polylactide, polyglyceride), and functionalized PEG, e.g., terminal-functionalized PEG.

[0013] The glycosyl core of the glycosyl linking groups of use in the Factor VII or Factor VIIa peptide conjugates are selected from both natural and unnatural furanoses and pyranoses. The unnatural saccharides optionally include an alkylated or acylated hydroxyl and/or amine moiety, e.g., ethers, esters and amide substituents on the ring. Other unnatural saccharides include an H, hydroxyl, ether, ester or amide substituent at a position on the ring

at which such a substituent is not present in the natural saccharide. Alternatively, the carbohydrate is missing a substituent that would be found in the carbohydrate from which its name is derived, e.g., deoxy sugars. Still further exemplary unnatural sugars include both oxidized (e.g., -onic and –uronic acids) and reduced (sugar alcohols) carbohydrates. The sugar moiety can be a mono-, oligo- or poly-saccharide.

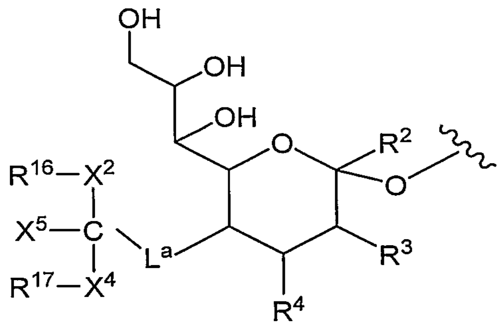
[0014] Exemplary natural sugars of use as components of glycosyl linking groups in the present invention include glucose, glucosamine, galactose, galactosamine, fucose, mannose, mannosamine, xylanose, ribose, N-acetyl glucose, N-acetyl glucosamine, N-acetyl galactose, N-acetyl galactosamine, and sialic acid.

[0015] In one embodiment, the present invention provides a Factor VII or Factor VIIa peptide conjugate comprising the moiety:



wherein D is a member selected from -OH and R^1-L-HN-; G is a member selected from H and R^1-L- and -C(O)(C_1-C_6)alkyl; R^1 is a moiety comprising a straight-chain or branched poly(ethylene glycol) residue; and L is a linker, e.g., a bond ("zero order"), substituted or unsubstituted alkyl and substituted or unsubstituted heteroalkyl. In exemplary embodiments, when D is OH, G is R^1-L-, and when G is –C(O)(C_1-C_6)alkyl, D is R^1-L-NH-.

[0016] In another aspect, the invention provides a Factor VII or VIIa peptide conjugate comprising a peptide which can be Factor VII or Factor VIIa. The conjugate also comprises a glycosyl linking group, wherein the glycosyl linking group is attached to an amino acid residue of said peptide, and wherein said glycosyl linking group comprises a sialyl linking group having a formula which is a member selected from:



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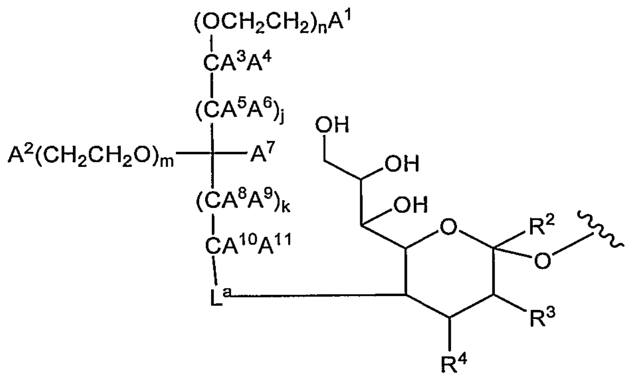
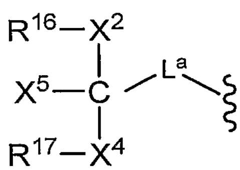
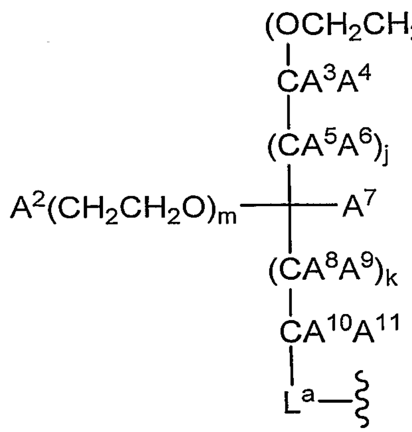


wherein



are modifying groups. R^2 is a member selected from H, CH_2OR^7, $COOR^7$, COO^- and OR^7. R^7 is a member selected from H, substituted or unsubstituted alkyl and substituted or unsubstituted heteroalkyl. R^3 and R^4 are members independently selected from H, substituted or unsubstituted alkyl, OR^8, and $NHC(O)R^9$. R^8 and R^9 are independently selected from H, substituted or unsubstituted alkyl, substituted or unsubstituted heteroalkyl and sialyl. L^a is a linker selected from a bond, substituted or unsubstituted alkyl and substituted or unsubstituted heteroalkyl. X^5, R^{16} and R^{17} are independently selected from non-reactive group and polymeric arms (e.g. PEG). X^2 and X^4 are independently selected linkage fragments joining polymeric moieties R^{16} and R^{17} to C. The index j is an integer selected from 1 to 15.

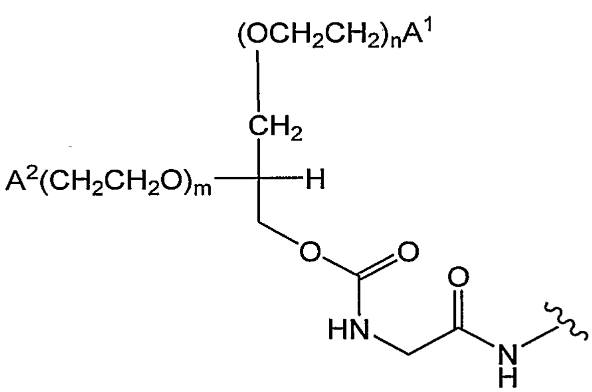
[0017] In another exemplary embodiment, the polymeric modifying group has a structure according to the following formula:



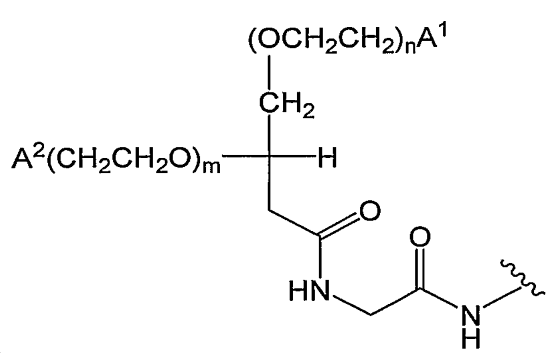
(IIIa)

in which the indices m and n are integers independently selected from 0 to 5000. A^1, A^2, A^3, A^4, A^5, A^6, A^7, A^8, A^9, A^{10} and A^{11} are members independently selected from H, substituted or unsubstituted alkyl, substituted or unsubstituted heteroalkyl, substituted or unsubstituted cycloalkyl, substituted or unsubstituted heterocycloalkyl, substituted or unsubstituted aryl, substituted or unsubstituted heteroaryl, $-NA^{12}A^{13}$, $-OA^{12}$ and $-SiA^{12}A^{13}$. A^{12} and A^{13} are members independently selected from substituted or unsubstituted alkyl, substituted or unsubstituted heteroalkyl, substituted or unsubstituted cycloalkyl, substituted or unsubstituted heterocycloalkyl, substituted or unsubstituted aryl, and substituted or unsubstituted heteroaryl.

[0018] In an exemplary embodiment, the polymeric modifying group has a structure according to the following formulae:



and



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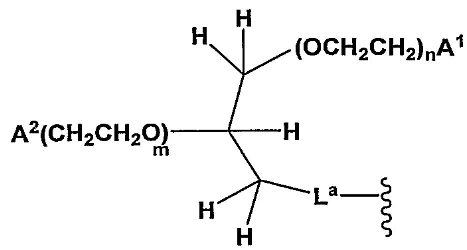
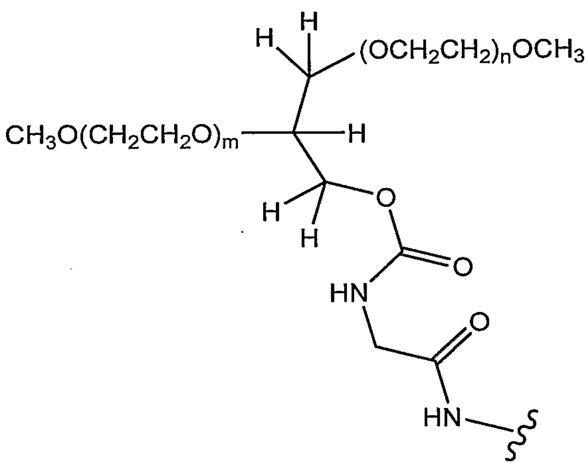
[0019] In another exemplary embodiment according to the formula above, the polymeric modifying group has a structure according to the following formula:

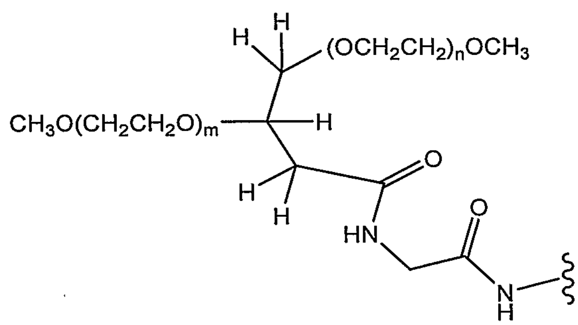


In an exemplary embodiment, A^1 and A^2 are each members selected from -OH and $-OCH_3$.

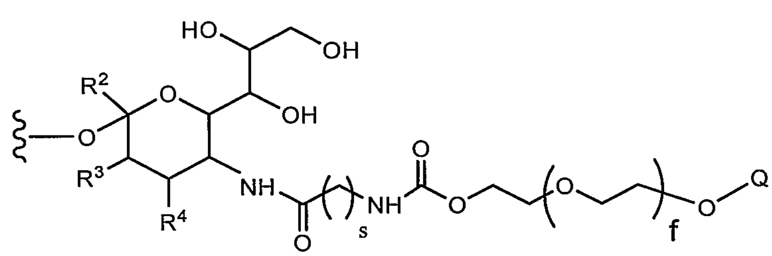
[0020] Exemplary polymeric modifying groups according to this embodiment include:



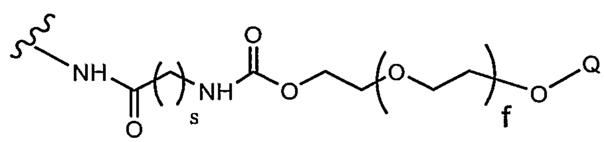
and



[0021] The invention provides a Factor VII or VIIa peptide conjugate comprising a peptide which is a member selected from Factor VII and Factor VIIa. The conjugate also comprises a glycosyl linking group, wherein the glycosyl linking group is attached to an amino acid residue of the peptide, and wherein the glycosyl linking group comprises a sialyl linking group having the formula:

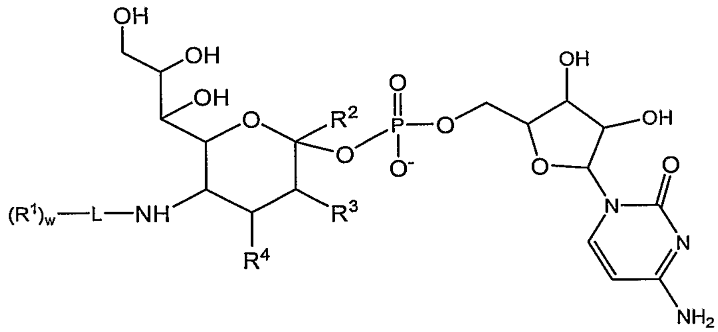


wherein



is a modifying group. The index s is an integer selected from 1 to 20. The index f is an integer selected from 1 to 2500. Q is a member selected from H and substituted or unsubstituted C_1-C_6 alkyl.

[0022] In an exemplary embodiment, the invention provides a modified sugar having the following formula:



[0023] The present invention provides methods of forming conjugates of Factor VII peptides, e.g., Factor VII and Factor VIIa. The methods include contacting a Factor VII/Factor VIIa peptide with a modified sugar donor that bears a modifying group covalently attached to a sugar. The modified sugar moiety is transferred from the donor onto an amino acid or glycosyl residue of the Factor VII/Factor VIIa peptide by the action of an enzyme. Representative enzymes include, but are not limited to, glycosyltransferases, e.g., sialyltransferases. The method includes contacting the Factor VII/Factor VIIa peptide with: a) a modified sugar donor; and b) an enzyme capable of transferring a modified sugar moiety from the modified sugar donor onto an amino acid or glycosyl residue of the peptide, under conditions appropriate to transfer a modified sugar moiety from the donor to an amino acid or glycosyl residue of the peptide, thereby synthesizing said Factor VII/Factor VIIa peptide conjugate.

[0024] In a preferred embodiment, prior to step a), the peptide is contacted with a sialidase, thereby removing at least a portion of the sialic acid on the peptide.

[0025] In another preferred embodiment, the Factor VII/Factor VIIa peptide is contacted with a sialidase, a glycosyltransferase and a modified sugar donor. In this embodiment, the peptide is in contact with the sialidase, glycosyltransferase and modified sugar donor essentially simultaneously, no matter the order of addition of the various components. The reaction is carried out under conditions appropriate for the sialidase to remove a sialic acid residue from the peptide; and the glycosyltransferase to transfer a modified sugar moiety from the modified sugar donor to an amino acid or glycosyl residue of the peptide.

[0026] In another preferred embodiment, the desialylation and conjugation are performed in the same vessel, and the desialylated peptide is preferably not purified prior to the

conjugation step. In another exemplary embodiment, the method further comprises a 'capping' step involving sialylation of the peptide conjugate. This step is performed in the same reaction vessel that contains the sialidase, sialyltransferase and modified sugar donor without prior purification.

[0027] In another preferred embodiment, the desialylation of the Factor VII/Factor VIIa peptide is performed, and the asialo peptide is purified. The purified asialo peptide is then subjected to conjugation reaction conditions. In another exemplary embodiment, the method further comprises a 'capping' step involving sialylation of the peptide conjugate. This step is performed in the same reaction vessel that contains the sialidase, sialyltransferase and modified sugar donor without prior purification.

[0028] In another exemplary embodiment, the capping step, sialylation of the peptide conjugate, is performed in the same reaction vessel that contains the sialidase, sialyltransferase and modified sugar donor without prior purification.

[0029] In an exemplary embodiment, the contacting is for a time less than 20 hours, preferably less than 16 hours, more preferably less than 12 hours, even more preferably less than 8 hours, and still more preferably less than 4 hours.

[0030] In a further aspect, the present invention provides a Factor VII/Factor VIIa peptide conjugate reaction mixture. The reaction mixture comprises: a) a sialidase; b) an enzyme which is a member selected from glycosyltransferase, exoglycosidase and endoglycosidase; c) a modified sugar; and d) a Factor VII/Factor VIIa peptide.

[0031] In another exemplary embodiment, the ratio of the sialidase to the Factor VII/Factor VIIa peptide is selected from 0.1 U/L:2 mg/mL to 10 U/L:1 mg/mL, preferably 0.5 U/L:2 mg/mL, more preferably 1.0 U/L:2 mg/mL, even more preferably 10 U/L:2 mg/mL, still more preferably 0.1 U/L:1 mg/mL, more preferably 0.5 U/L:1 mg/mL, even more preferably 1.0 U/L:1 mg/mL, and still more preferably 10 U/L:1 mg/mL.

[0032] In an exemplary embodiment, at least 10%, 20%, 30%, 40%, 50%, 60%, 70% or 80% of said Factor VII/Factor VIIa peptide conjugate includes at most two PEG moieties. The PEG moieties can be added in a one-pot process, or they can be added after the asialo Factor VII/Factor VIIa is purified.

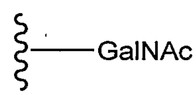
[0033] In another exemplary embodiment, at least 10%, 20%, 30%, 40%, 50%, 60%, 70% or 80% of the Factor VII/Factor VIIa peptide conjugate include at most one PEG

moiety. The PEG moiety can be added in a one-pot process, or it can be added after the asialo Factor VII/Factor VIIa is purified.

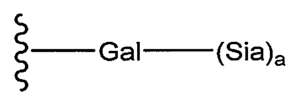
[0034] In a further exemplary embodiment, the method further comprises "capping", or adding sialic acid to the peptide conjugate. In another exemplary embodiment, sialidase is added, followed by a delay of 30 min, 1 hour, 1.5 hours, or 2 hours, followed by the addition of the glycosyltransferase, exoglycosidase, or endoglycosidase.

[0035] In another exemplary embodiment, sialidase is added, followed by a delay of 30 min, 1 hour, 1.5 hours, or 2 hours, followed by the addition of the glycosyltransfase, exoglycosidase, or endoglycosidase. Other objects and advantages of the invention will be apparent to those of skill in the art from the detailed description that follows.

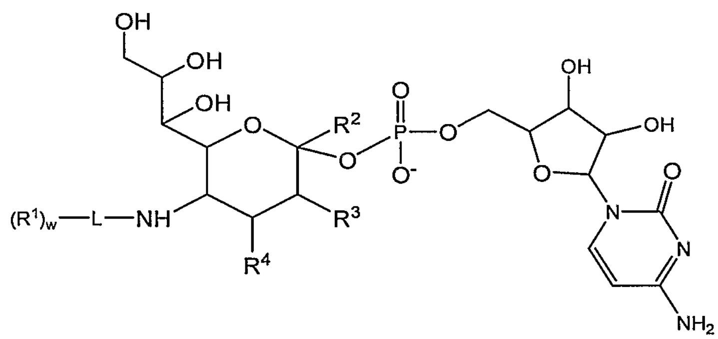
[0036] In another exemplary embodiment, the method includes: (a) contacting a Factor VII/Factor VIIa peptide comprising a glycosyl group selected from:



; and



with a modified sugar having the formula:



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and an appropriate transferase which transfers the glysocyl linking group onto a member selected from the GalNAc, Gal and the Sia of said glycosyl group, under conditions appropriate for said transfer. An exemplary modified sugar is CMP-sialic acid modified, through a linker moiety, with a polymer, e.g., a straight chain or branched poly(ethylene glycol) moiety.

[0037] The peptide can be acquired from essentially any source, however, in one embodiment, prior to being modified as discussed above, the Factor VII/Factor VIIa peptide is expressed in a suitable host. Mammalian (e.g., BHK, CHO), bacteria (e.g., E. coli) and insect cells (e.g., Sf-9) are exemplary expression systems providing Factor VII or Factor VIIa of use in the compositions and methods set forth herein.

[0038] In exemplary embodiments, a Factor VII/Factor VIIa peptide conjugate may be administered to patients for the treatment of a tissue injury such as ischemia, trauma, inflammation, or contact with toxic substances. In other exemplary embodiments, a Factor VII/Factor VIIa peptide conjugate may be administered to patients for the treatment of a patient having Hemophilia A, a patient with Hemophilia B, a patient having Hemophilia A, wherein the patient also has antibodies to Factor VIII, a patient having Hemophilia B, wherein the patient also has antibodies to Factor IX, and a patient having liver cirrhosis.

[0039] In another exemplary embodiment, a Factor VII/Factor VIIa peptide conjugate may be administered to patients for the treatment of bleeding in emergencies, elective surgery, cardiac surgery, spinal surgery, liver transplantation, partial hepatectomies, pelvic-acetabular fracture reconstruction, and allogeneic stem cell transplantation. In another exemplary embodiment, a Factor VII/Factor VIIa peptide conjugate may be administered to patients for the treatment of acute intracerebral haemorrhage, traumatic brain injury, variceal bleedings and upper gastrointestinal bleeding.

[0040] In another aspect, the invention provides a pharmaceutical formulation comprising a Factor VII/Factor VIIa peptide conjugate and a pharmaceutically acceptable carrier.

[0041] In the Factor VII/Factor VIIa peptide conjugate, essentially each of the amino acid residues to which the glycosyl linking group or modifying group is bound has the same structure. For example, if one peptide includes a Thr linked glycosyl residue, at least about 70%, 80%, 90%, 95%, 97%, 99%, 99.2%, 99.4%, 99.6%, or more preferably 99.8% of the peptides in the population will have the same glycosyl linking group covalently bound to the same Thr residue.

[0042] Other objects and advantages of the invention will be apparent to those of skill in the art from the detailed description that follows.

DESCRIPTION OF THE DRAWINGS

[0043] **FIG. 1** illustrates exemplary modified sialic acid nucleotides useful in the practice of the invention. **A.** Structure of exemplary branched (e.g., 30 KDa, 40 KDa) CMP-sialic acid-PEG sugar nucleotides. **B.** Structure of linear Factor VIIa-SA-PEG-10KDa.

[0044] **FIG. 2** is a synthetic scheme for producing an exemplary PEG-glycosyl linking group precursor (modified sugar) of use in preparing the conjugates of the invention.

[0045] **FIG. 3** is a table providing exemplary sialyltransferases of use in forming the glycoconjugates of the invention, e.g., to glycoPEGylate peptides with a modified sialic acid.

[0046] **FIG. 4**, comprising Figures 4A to 4E, sets forth exemplary schemes for remodeling glycan structures on Factor VII and Factor VIIa. Figure 4A is a diagram depicting the Factor VII and Factor VIIa peptides indicating the residues which bind to glycans contemplated for remodeling. Figure 4B is a diagram depicting the Factor VII and Factor VIIa peptides A (solid line) and B (dotted line) indicating the residues which bind to glycans contemplated for remodeling, and the formulas for the glycans. Figures 4C to 4E are diagrams of contemplated remodeling steps of the glycan of the peptide in Figure 4B based on the type of cell the peptide is expressed in and the desired remodeled glycan structure.

[0047] **FIG. 5**, comprising Figures 5A and 5B, is an exemplary nucleotide and corresponding amino acid sequence of Factor VIIa (SEQ ID NOS: 1 and 2, respectively).

[0048] **FIG. 6** is an image of an isoelectric focusing gel (pH 3-7) of asialo-Factor VIIa. Lane 1 is Factor VIIa; lanes 2-5 are asialo-Factor VIIa.

[0049] **FIG. 7** is a graph of a MALDI spectra of Factor VIIa.

[0050] **FIG. 8** is a graph of a MALDI spectra of Factor VIIa-SA-PEG-1KDa.

[0051] **FIG. 9** is a graph depicting a MALDI spectra of Factor VIIa-SA-PEG-10KDa.

[0052] **FIG. 10** is an image of an SDS-PAGE gel of PEGylated Factor VIIa. Lane 1 is asialo-Factor VIIa. Lane 2 is the product of the reaction of asialo-Factor VIIa and CMP-SA-PEG-1KDa with ST3Gal3 after 48 hr. Lane 3 is the product of the reaction of asialo-Factor VIIa and CMP-SA-PEG-1KDa with ST3Gal3 after 48 hr. Lane 4 is the product of the reaction of asialo-Factor VIIa and CMP-SA-PEG-10KDa with ST3Gal3 at 96 hr.

[0053] **FIG. 11 A-B** shows simultaneous desialylation, with less sialidase, and PEGylation. These figures highlight that capping in the presence of sialidase is efficient. Figure 11A shows the reaction course when the sialidase is at a level of 0.5 U/L. Lane 1 corresponds to native Factor VIIa while Lane 2 is asialo Factor VIIa. From Lane 3 to Lane 7, there is an increasing amount of PEGylated product as time progresses. In Lane 3, the major product is monoPEGylated (see spot at 64), while aliquots assayed at later times show the formation and increasing amounts of di (see spot just below 97), tri (see spot just above 97), and higher PEGylated products. Lanes 8 and 9 show the results of 'capping', or adding sialic

acid, to the reaction. When the reaction is capped, the extent of reaction is stopped, as can be seen from the similar PEGylated product distribution found in Lanes 5, 8 and 9. Figure 11 B shows the reaction course when the sialidase is at a level of 0.1 U/L.

[0054] **FIG. 12 A** and **B**. Fig 12 A shows the situation when the sialidase and the glycosyltransferase are added at the same time. Figure 12B shows the situation when the sialidase is added first, followed by glycosyltransferase after a 30 minute delay.

[0055] **FIG. 13** is a table of the peptides to which one or more glycosyl linking groups can be attached to order to provide the peptide conjugates of the invention.

[0056] **FIG. 14 A** and **B** displays chromatograms showing the results of HPLC experiments. **FIG. 14A** displays labeled chromatograms of Factor VIIa-SA-PEG-10KDa (top) and native Factor VIIa control (bottom) analyzed by the light chain method. The separation of LC (light chain), 1x10KDa-PEG-LC, 2x10KDa-PEG-LC, and 3x10KDa-PEG-LC from other products is shown. **FIG. 14B** displays labeled chromatograms of Factor VIIa-SA-PEG-10KDa (top) and native Factor VIIa control (bottom) analyzed by heavy chain method. The separation of HC (heavy chain), 1x10 KDa -PEG-HC, 2x10 KDa -PEG-HC, and 3x10 KDa -PEG-HC from other products is shown.

[0057] **FIG. 15 A** and **B** displays chromatograms showing the results of HPLC experiments. **FIG. 15A** displays labeled chromatograms of reduced native Factor VIIa control (top) and reduced Factor VIIa-SA-PEG-40KDa (bottom) analyzed by the light chain method. The separation of LC (light chain), 1x40KDa-PEG-LC, 2x40 KDa -PEG-LC, and 3x40KDa -PEG-LC from other products is shown. **FIG. 15B** displays labeled chromatograms of reduced native Factor VIIa control (top) and Factor VIIa-SA-PEG-40KDa (bottom) analyzed by the heavy chain method. The separation of HC (heavy chain), 1x40KDa -PEG-HC, 2x40KDa -PEG-HC, and 3x40KDa -PEG-HC from other products is shown.

DETAILED DESCRIPTION OF THE INVENTION AND THE PREFERRED EMBODIMENTS

Abbreviations

[0058] PEG, poly(ethyleneglycol); PPG, poly(propyleneglycol); Ara, arabinosyl; Fru, fructosyl; Fuc, fucosyl; Gal, galactosyl; GalNAc, N-acetylgalactosaminyl; Glc, glucosyl; GlcNAc, N-acetylglucosaminyl; Man, mannosyl; ManAc, mannosaminyl acetate; Xyl,

xylosyl; NeuAc, sialyl or N-acetylneuraminyl; Sia, sialyl or N-acetylneuraminyl; and derivatives and analogues thereof.

Definitions

[0059] Unless defined otherwise, all technical and scientific terms used herein generally have the same meaning as commonly understood by one of ordinary skill in the art to which this invention belongs. Generally, the nomenclature used herein and the laboratory procedures in cell culture, molecular genetics, organic chemistry and nucleic acid chemistry and hybridization are those well known and commonly employed in the art. Standard techniques are used for nucleic acid and peptide synthesis. The techniques and procedures are generally performed according to conventional methods in the art and various general references (*see generally*, Sambrook *et al.* MOLECULAR CLONING: A LABORATORY MANUAL, 2d ed. (1989) Cold Spring Harbor Laboratory Press, Cold Spring Harbor, N.Y., which is incorporated herein by reference), which are provided throughout this document. The nomenclature used herein and the laboratory procedures in analytical chemistry, and organic synthetic described below are those well known and commonly employed in the art. Standard techniques, or modifications thereof, are used for chemical syntheses and chemical analyses.

[0060] All oligosaccharides described herein are described with the name or abbreviation for the non-reducing saccharide (*i.e.*, Gal), followed by the configuration of the glycosidic bond (α or β), the ring bond (1 or 2), the ring position of the reducing saccharide involved in the bond (2, 3, 4, 6 or 8), and then the name or abbreviation of the reducing saccharide (*i.e.*, GlcNAc). Each saccharide is preferably a pyranose. For a review of standard glycobiology nomenclature, *see, Essentials of Glycobiology* Varki *et al.* eds. CSHL Press (1999).

[0061] Oligosaccharides are considered to have a reducing end and a non-reducing end, whether or not the saccharide at the reducing end is in fact a reducing sugar. In accordance with accepted nomenclature, oligosaccharides are depicted herein with the non-reducing end on the left and the reducing end on the right.

[0062] The term "sialic acid" or "sialyl" refers to any member of a family of nine-carbon carboxylated sugars. The most common member of the sialic acid family is N-acetyl-neuraminic acid (2-keto-5-acetamido-3,5-dideoxy-D-glycero-D-galactononulopyranos-1-onic acid (often abbreviated as Neu5Ac, NeuAc, or NANA). A second member of the family is N-glycolyl-neuraminic acid (Neu5Gc or NeuGc), in which the N-acetyl group of NeuAc is

hydroxylated. A third sialic acid family member is 2-keto-3-deoxy-nonulosonic acid (KDN) (Nadano *et al.* (1986) *J. Biol. Chem.* **261**: 11550-11557; Kanamori *et al., J. Biol. Chem.* **265**: 21811-21819 (1990)). Also included are 9-substituted sialic acids such as a 9-O-C_1-C_6 acyl-Neu5Ac like 9-O-lactyl-Neu5Ac or 9-O-acetyl-Neu5Ac, 9-deoxy-9-fluoro-Neu5Ac and 9-azido-9-deoxy-Neu5Ac. For review of the sialic acid family, *see, e.g.*, Varki, *Glycobiology* **2**: 25-40 (1992); *Sialic Acids: Chemistry, Metabolism and Function*, R. Schauer, Ed. (Springer-Verlag, New York (1992)). The synthesis and use of sialic acid compounds in a sialylation procedure is disclosed in international application WO 92/16640, published October 1, 1992.

[0063] "Peptide" refers to a polymer in which the monomers are amino acids and are joined together through amide bonds, alternatively referred to as a polypeptide. Additionally, unnatural amino acids, for example, β-alanine, phenylglycine and homoarginine are also included. Amino acids that are not gene-encoded may also be used in the present invention. Furthermore, amino acids that have been modified to include reactive groups, glycosylation sites, polymers, therapeutic moieties, biomolecules and the like may also be used in the invention. All of the amino acids used in the present invention may be either the D - or L - isomer. The L -isomer is generally preferred. In addition, other peptidomimetics are also useful in the present invention. As used herein, "peptide" refers to both glycosylated and unglycosylated peptides. Also included are peptides that are incompletely glycosylated by a system that expresses the peptide. For a general review, *see,* Spatola, A. F., in CHEMISTRY AND BIOCHEMISTRY OF AMINO ACIDS, PEPTIDES AND PROTEINS, B. Weinstein, eds., Marcel Dekker, New York, p. 267 (1983). A listing of some of the peptides of the invention is provided in **FIG. 13**.

[0064] The term "peptide conjugate," refers to species of the invention in which a peptide is conjugated with a modified sugar as set forth herein.

[0065] The term "amino acid" refers to naturally occurring and synthetic amino acids, as well as amino acid analogs and amino acid mimetics that function in a manner similar to the naturally occurring amino acids. Naturally occurring amino acids are those encoded by the genetic code, as well as those amino acids that are later modified, *e.g.*, hydroxyproline, γ-carboxyglutamate, and O-phosphoserine. Amino acid analogs refers to compounds that have the same basic chemical structure as a naturally occurring amino acid, *i.e.*, an α carbon that is bound to a hydrogen, a carboxyl group, an amino group, and an R group, *e.g.*, homoserine, norleucine, methionine sulfoxide, methionine methyl sulfonium. Such analogs have modified R groups (*e.g.*, norleucine) or modified peptide backbones, but retain the same basic chemical

structure as a naturally occurring amino acid. Amino acid mimetics refers to chemical compounds that have a structure that is different from the general chemical structure of an amino acid, but that function in a manner similar to a naturally occurring amino acid.

[0066] As used herein, the term "modified sugar," or "modified sugar residue", refers to a naturally- or non-naturally-occurring carbohydrate that is enzymatically added onto an amino acid or a glycosyl residue of a peptide in a process of the invention. The modified sugar is selected from enzyme substrates including, but not limited to sugar nucleotides (mono-, di-, and tri-phosphates), activated sugars (*e.g.*, glycosyl halides, glycosyl mesylates) and sugars that are neither activated nor nucleotides. The "modified sugar" is covalently functionalized with a "modifying group." Useful modifying groups include, but are not limited to, PEG moieties, therapeutic moieties, diagnostic moieties, biomolecules and the like. The modifying group is preferably not a naturally occurring, or an unmodified carbohydrate. The locus of functionalization with the modifying group is selected such that it does not prevent the "modified sugar" from being added enzymatically to a peptide.

[0067] The term "water-soluble" refers to moieties that have some detectable degree of solubility in water. Methods to detect and/or quantify water solubility are well known in the art. Exemplary water-soluble polymers include peptides, saccharides, poly(ethers), poly(amines), poly(carboxylic acids) and the like. Peptides can have mixed sequences of be composed of a single amino acid, *e.g.*, poly(lysine). An exemplary polysaccharide is poly(sialic acid). An exemplary poly(ether) is poly(ethylene glycol). Poly(ethylene imine) is an exemplary polyamine, and poly(acrylic) acid is a representative poly(carboxylic acid).

[0068] The polymer backbone of the water-soluble polymer can be poly(ethylene glycol) (i.e. PEG). However, it should be understood that other related polymers are also suitable for use in the practice of this invention and that the use of the term PEG or poly(ethylene glycol) is intended to be inclusive and not exclusive in this respect. The term PEG includes poly(ethylene glycol) in any of its forms, including alkoxy PEG, difunctional PEG, multiarmed PEG, forked PEG, branched PEG, pendent PEG (i.e. PEG or related polymers having one or more functional groups pendent to the polymer backbone), or PEG with degradable linkages therein.

[0069] The polymer backbone can be linear or branched. Branched polymer backbones are generally known in the art. Typically, a branched polymer has a central branch core moiety and a plurality of linear polymer chains linked to the central branch core. PEG is

commonly used in branched forms that can be prepared by addition of ethylene oxide to various polyols, such as glycerol, pentaerythritol and sorbitol. The central branch moiety can also be derived from several amino acids, such as lysine. The branched poly(ethylene glycol) can be represented in general form as $R(\text{-PEG-OH})_m$ in which R represents the core moiety, such as glycerol or pentaerythritol, and m represents the number of arms. Multi-armed PEG molecules, such as those described in U.S. Pat. No. 5,932,462, which is incorporated by reference herein in its entirety, can also be used as the polymer backbone.

[0070] Many other polymers are also suitable for the invention. Polymer backbones that are non-peptidic and water-soluble, within about 2 to about 300 loci for attachment, are particularly useful in the invention. Examples of suitable polymers include, but are not limited to, other poly(alkylene glycols), such as poly(propylene glycol) ("PPG"), copolymers of ethylene glycol and propylene glycol and the like, poly(oxyethylated polyol), poly(olefinic alcohol), poly(vinylpyrrolidone), poly(hydroxypropylmethacrylamide), poly(α-hydroxy acid), poly(vinyl alcohol), polyphosphazene, polyoxazoline, poly(N-acryloylmorpholine), such as described in U.S. Pat. No. 5,629,384, which is incorporated by reference herein in its entirety, and copolymers, terpolymers, and mixtures thereof. Although the molecular weight of each chain of the polymer backbone can vary, it is typically in the range of from about 100 Da to about 100,000 Da, often from about 6,000 Da to about 80,000 Da.

[0071] The "area under the curve" or "AUC", as used herein in the context of administering a peptide drug to a patient, is defined as total area under the curve that describes the concentration of drug in systemic circulation in the patient as a function of time from zero to infinity.

[0072] The term "half-life" or "t½", as used herein in the context of administering a peptide drug to a patient, is defined as the time required for plasma concentration of a drug in a patient to be reduced by one half. There may be more than one half-life associated with the peptide drug depending on multiple clearance mechanisms, redistribution, and other mechanisms well known in the art. Usually, alpha and beta half-lives are defined such that the alpha phase is associated with redistribution, and the beta phase is associated with clearance. However, with protein drugs that are, for the most part, confined to the bloodstream, there can be at least two clearance half-lives. For some glycosylated peptides, rapid beta phase clearance may be mediated via receptors on macrophages, or endothelial cells that recognize terminal galactose, N-acetylgalactosamine, N-acetylglucosamine, mannose, or fucose. Slower beta phase clearance may occur via renal glomerular filtration

for molecules with an effective radius < 2 nm (approximately 68 kD) and/or specific or non-specific uptake and metabolism in tissues. GlycoPEGylation may cap terminal sugars (*e.g.*, galactose or N-acetylgalactosamine) and thereby block rapid alpha phase clearance via receptors that recognize these sugars. It may also confer a larger effective radius and thereby decrease the volume of distribution and tissue uptake, thereby prolonging the late beta phase. Thus, the precise impact of glycoPEGylation on alpha phase and beta phase half-lives may vary depending upon the size, state of glycosylation, and other parameters, as is well known in the art. Further explanation of "half-life" is found in Pharmaceutical Biotechnology (1997, DFA Crommelin and RD Sindelar, eds., Harwood Publishers, Amsterdam, pp 101 – 120).

[0073] The term "glycoconjugation," as used herein, refers to the enzymatically mediated conjugation of a modified sugar species to an amino acid or glycosyl residue of a polypeptide, *e.g.*, a G-CSF peptide of the present invention. A subgenus of "glycoconjugation" is "glyco-PEGylation," in which the modifying group of the modified sugar is poly(ethylene glycol), and alkyl derivative (*e.g.*, m-PEG) or reactive derivative (*e.g.*, H_2N-PEG, HOOC-PEG) thereof.

[0074] The terms "large-scale" and "industrial-scale" are used interchangeably and refer to a reaction cycle that produces at least about 250 mg, preferably at least about 500 mg, and more preferably at least about 1 gram of glycoconjugate at the completion of a single reaction cycle.

[0075] The term, "glycosyl linking group," as used herein refers to a glycosyl residue to which a modifying group (e.g., PEG moiety, therapeutic moiety, biomolecule) is covalently attached; the glycosyl linking group joins the modifying group to the remainder of the conjugate. In the methods of the invention, the "glycosyl linking group" becomes covalently attached to a glycosylated or unglycosylated peptide, thereby linking the agent to an amino acid and/or glycosyl residue on the peptide. A "glycosyl linking group" is generally derived from a "modified sugar" by the enzymatic attachment of the "modified sugar" to an amino acid and/or glycosyl residue of the peptide. The glycosyl linking group can be a saccharide-derived structure that is degraded during formation of modifying group-modified sugar cassette (e.g., oxidation→Schiff base formation→reduction), or the glycosyl linking group may be intact. An "intact glycosyl linking group" refers to a linking group that is derived from a glycosyl moiety in which the saccharide monomer that links the modifying group and to the remainder of the conjugate is not degraded, e.g., oxidized, e.g., by sodium metaperiodate. "Intact glycosyl linking groups" of the invention may be derived from a

naturally occurring oligosaccharide by addition of glycosyl unit(s) or removal of one or more glycosyl unit from a parent saccharide structure.

[0076] The term, "non-glycosidic modifying group", as used herein, refers to modifying groups which do not include a naturally occurring sugar linked directly to the glycosyl linking group.

[0077] The term "targeting moiety," as used herein, refers to species that will selectively localize in a particular tissue or region of the body. The localization is mediated by specific recognition of molecular determinants, molecular size of the targeting agent or conjugate, ionic interactions, hydrophobic interactions and the like. Other mechanisms of targeting an agent to a particular tissue or region are known to those of skill in the art. Exemplary targeting moieties include antibodies, antibody fragments, transferrin, HS-glycoprotein, coagulation factors, serum proteins, β-glycoprotein, G-CSF, GM-CSF, M-CSF, EPO and the like.

[0078] As used herein, "therapeutic moiety" means any agent useful for therapy including, but not limited to, antibiotics, anti-inflammatory agents, anti-tumor drugs, cytotoxins, and radioactive agents. "Therapeutic moiety" includes prodrugs of bioactive agents, constructs in which more than one therapeutic moiety is bound to a carrier, e.g, multivalent agents. Therapeutic moiety also includes proteins and constructs that include proteins. Exemplary proteins include, but are not limited to, Granulocyte Colony Stimulating Factor (GCSF), Granulocyte Macrophage Colony Stimulating Factor (GMCSF), Interferon (e.g., Interferon-α, -β, -γ), Interleukin (e.g., Interleukin II), serum proteins (e.g., Factors VII, VIIa, VIII, IX, and X), Human Chorionic Gonadotropin (HCG), Follicle Stimulating Hormone (FSH) and Lutenizing Hormone (LH) and antibody fusion proteins (e.g. Tumor Necrosis Factor Receptor ((TNFR)/Fc domain fusion protein)).

[0079] As used herein, "pharmaceutically acceptable carrier" includes any material, which when combined with the conjugate retains the conjugates' activity and is non-reactive with the subject's immune systems. Examples include, but are not limited to, any of the standard pharmaceutical carriers such as a phosphate buffered saline solution, water, emulsions such as oil/water emulsion, and various types of wetting agents. Other carriers may also include sterile solutions, tablets including coated tablets and capsules. Typically such carriers contain excipients such as starch, milk, sugar, certain types of clay, gelatin, stearic acid or salts thereof, magnesium or calcium stearate, talc, vegetable fats or oils, gums,

glycols, or other known excipients. Such carriers may also include flavor and color additives or other ingredients. Compositions comprising such carriers are formulated by well known conventional methods.

[0080] As used herein, "administering," means oral administration, administration as a suppository, topical contact, intravenous, intraperitoneal, intramuscular, intralesional, intranasal or subcutaneous administration, or the implantation of a slow-release device *e.g.*, a mini-osmotic pump, to the subject. Administration is by any route including parenteral, and transmucosal (e.g., oral, nasal, vaginal, rectal, or transdermal). Parenteral administration includes, e.g., intravenous, intramuscular, intra-arteriole, intradermal, subcutaneous, intraperitoneal, intraventricular, and intracranial. Moreover, where injection is to treat a tumor, e.g., induce apoptosis, administration may be directly to the tumor and/or into tissues surrounding the tumor. Other modes of delivery include, but are not limited to, the use of liposomal formulations, intravenous infusion, transdermal patches, etc.

[0081] The term "ameliorating" or "ameliorate" refers to any indicia of success in the treatment of a pathology or condition, including any objective or subjective parameter such as abatement, remission or diminishing of symptoms or an improvement in a patient's physical or mental well-being. Amelioration of symptoms can be based on objective or subjective parameters; including the results of a physical examination and/or a psychiatric evaluation.

[0082] The term “therapy” refers to "treating" or "treatment" of a disease or condition including preventing the disease or condition from occurring in an animal that may be predisposed to the disease but does not yet experience or exhibit symptoms of the disease (prophylactic treatment), inhibiting the disease (slowing or arresting its development), providing relief from the symptoms or side-effects of the disease (including palliative treatment), and relieving the disease (causing regression of the disease).

[0083] The term “effective amount” or “an amount effective to” or a "therapeutically effective amount" or any grammatically equivalent term means the amount that, when administered to an animal for treating a disease, is sufficient to effect treatment for that disease.

[0084] The term “isolated” refers to a material that is substantially or essentially free from components, which are used to produce the material. For peptide conjugates of the invention, the term “isolated” refers to material that is substantially or essentially free from components which normally accompany the material in the mixture used to prepare the

peptide conjugate. "Isolated" and "pure" are used interchangeably. Typically, isolated peptide conjugates of the invention have a level of purity preferably expressed as a range. The lower end of the range of purity for the peptide conjugates is about 60%, about 70% or about 80% and the upper end of the range of purity is about 70%, about 80%, about 90% or more than about 90%.

[0085] When the peptide conjugates are more than about 90% pure, their purities are also preferably expressed as a range. The lower end of the range of purity is about 90%, about 92%, about 94%, about 96% or about 98%. The upper end of the range of purity is about 92%, about 94%, about 96%, about 98% or about 100% purity.

[0086] Purity is determined by any art-recognized method of analysis (*e.g.*, band intensity on a silver stained gel, polyacrylamide gel electrophoresis, HPLC, or a similar means).

[0087] "Essentially each member of the population," as used herein, describes a characteristic of a population of peptide conjugates of the invention in which a selected percentage of the modified sugars added to a peptide are added to multiple, identical acceptor sites on the peptide. "Essentially each member of the population" speaks to the "homogeneity" of the sites on the peptide conjugated to a modified sugar and refers to conjugates of the invention, which are at least about 80%, preferably at least about 90% and more preferably at least about 95% homogenous.

[0088] "Homogeneity," refers to the structural consistency across a population of acceptor moieties to which the modified sugars are conjugated. Thus, in a peptide conjugate of the invention in which each modified sugar moiety is conjugated to an acceptor site having the same structure as the acceptor site to which every other modified sugar is conjugated, the peptide conjugate is said to be about 100% homogeneous. Homogeneity is typically expressed as a range. The lower end of the range of homogeneity for the peptide conjugates is about 60%, about 70% or about 80% and the upper end of the range of purity is about 70%, about 80%, about 90% or more than about 90%.

[0089] When the peptide conjugates are more than or equal to about 90% homogeneous, their homogeneity is also preferably expressed as a range. The lower end of the range of homogeneity is about 90%, about 92%, about 94%, about 96% or about 98%. The upper end of the range of purity is about 92%, about 94%, about 96%, about 98% or about 100% homogeneity. The purity of the peptide conjugates is typically determined by one or more methods known to those of skill in the art, e.g., liquid chromatography-mass spectrometry

(LC-MS), matrix assisted laser desorption mass time of flight spectrometry (MALDITOF), capillary electrophoresis, and the like.

[0090] "Substantially uniform glycoform" or a "substantially uniform glycosylation pattern," when referring to a glycopeptide species, refers to the percentage of acceptor moieties that are glycosylated by the glycosyltransferase of interest (*e.g.*, fucosyltransferase). For example, in the case of a α1,2 fucosyltransferase, a substantially uniform fucosylation pattern exists if substantially all (as defined below) of the Galβ1,4-GlcNAc-R and sialylated analogues thereof are fucosylated in a peptide conjugate of the invention. In the fucosylated structures set forth herein, the Fuc-GlcNAc linkage is generally α1,6 or α1,3, with α1,6 generally preferred. It will be understood by one of skill in the art, that the starting material may contain glycosylated acceptor moieties (*e.g.*, fucosylated Galβ1,4-GlcNAc-R moieties). Thus, the calculated percent glycosylation will include acceptor moieties that are glycosylated by the methods of the invention, as well as those acceptor moieties already glycosylated in the starting material.

[0091] The term "substantially" in the above definitions of "substantially uniform" generally means at least about 40%, at least about 70%, at least about 80%, or more preferably at least about 90%, and still more preferably at least about 95% of the acceptor moieties for a particular glycosyltransferase are glycosylated.

[0092] Where substituent groups are specified by their conventional chemical formulae, written from left to right, they equally encompass the chemically identical substituents, which would result from writing the structure from right to left, e.g., $-CH_2O-$ is intended to also recite $-OCH_2-$.

[0093] The term "alkyl," by itself or as part of another substituent means, unless otherwise stated, a straight or branched chain, or cyclic hydrocarbon radical, or combination thereof, which may be fully saturated, mono- or polyunsaturated and can include di- and multivalent radicals, having the number of carbon atoms designated (*i.e.* C_1-C_{10} means one to ten carbons). Examples of saturated hydrocarbon radicals include, but are not limited to, groups such as methyl, ethyl, n-propyl, isopropyl, n-butyl, t-butyl, isobutyl, sec-butyl, cyclohexyl, (cyclohexyl)methyl, cyclopropylmethyl, homologs and isomers of, for example, n-pentyl, n-hexyl, n-heptyl, n-octyl, and the like. An unsaturated alkyl group is one having one or more double bonds or triple bonds. Examples of unsaturated alkyl groups include, but are not limited to, vinyl, 2-propenyl, crotyl, 2-isopentenyl, 2-(butadienyl), 2,4-pentadienyl, 3-

(1,4-pentadienyl), ethynyl, 1- and 3-propynyl, 3-butynyl, and the higher homologs and isomers. The term "alkyl," unless otherwise noted, is also meant to include those derivatives of alkyl defined in more detail below, such as "heteroalkyl." Alkyl groups that are limited to hydrocarbon groups are termed "homoalkyl".

[0094] The term "alkylene" by itself or as part of another substituent means a divalent radical derived from an alkane, as exemplified, but not limited, by $–CH_2CH_2CH_2CH_2$-, and further includes those groups described below as "heteroalkylene." Typically, an alkyl (or alkylene) group will have from 1 to 24 carbon atoms, with those groups having 10 or fewer carbon atoms being preferred in the present invention. A "lower alkyl" or "lower alkylene" is a shorter chain alkyl or alkylene group, generally having eight or fewer carbon atoms.

[0095] The terms "alkoxy," "alkylamino" and "alkylthio" (or thioalkoxy) are used in their conventional sense, and refer to those alkyl groups attached to the remainder of the molecule via an oxygen atom, an amino group, or a sulfur atom, respectively.

[0096] The term "heteroalkyl," by itself or in combination with another term, means, unless otherwise stated, a stable straight or branched chain, or cyclic hydrocarbon radical, or combinations thereof, consisting of the stated number of carbon atoms and at least one heteroatom selected from the group consisting of O, N, Si and S, and wherein the nitrogen and sulfur atoms may optionally be oxidized and the nitrogen heteroatom may optionally be quaternized. The heteroatom(s) O, N and S and Si may be placed at any interior position of the heteroalkyl group or at the position at which the alkyl group is attached to the remainder of the molecule. Examples include, but are not limited to, $-CH_2-CH_2-O-CH_3$, $-CH_2-CH_2-NH-CH_3$, $-CH_2-CH_2-N(CH_3)-CH_3$, $-CH_2-S-CH_2-CH_3$, $-CH_2-CH_2,-S(O)-CH_3$, $-CH_2-CH_2-S(O)_2-CH_3$, $-CH=CH-O-CH_3$, $-Si(CH_3)_3$, $-CH_2-CH=N-OCH_3$, and $–CH=CH-N(CH_3)-CH_3$. Up to two heteroatoms may be consecutive, such as, for example, $-CH_2-NH-OCH_3$ and $–CH_2-O-Si(CH_3)_3$. Similarly, the term "heteroalkylene" by itself or as part of another substituent means a divalent radical derived from heteroalkyl, as exemplified, but not limited by, $-CH_2-CH_2-S-CH_2-CH_2$- and $–CH_2-S-CH_2-CH_2-NH-CH_2$-. For heteroalkylene groups, heteroatoms can also occupy either or both of the chain termini (*e.g.*, alkyleneoxy, alkylenedioxy, alkyleneamino, alkylenediamino, and the like). Still further, for alkylene and heteroalkylene linking groups, no orientation of the linking group is implied by the direction in which the formula of the linking group is written. For example, the formula $–C(O)_2R'$- represents both $–C(O)_2R'$- and $–R'C(O)_2$-.

[0097] The terms "cycloalkyl" and "heterocycloalkyl", by themselves or in combination with other terms, represent, unless otherwise stated, cyclic versions of "alkyl" and "heteroalkyl", respectively. Additionally, for heterocycloalkyl, a heteroatom can occupy the position at which the heterocycle is attached to the remainder of the molecule. Examples of cycloalkyl include, but are not limited to, cyclopentyl, cyclohexyl, 1-cyclohexenyl, 3-cyclohexenyl, cycloheptyl, and the like. Examples of heterocycloalkyl include, but are not limited to, 1 –(1,2,5,6-tetrahydropyridyl), 1-piperidinyl, 2-piperidinyl, 3-piperidinyl, 4-morpholinyl, 3-morpholinyl, tetrahydrofuran-2-yl, tetrahydrofuran-3-yl, tetrahydrothien-2-yl, tetrahydrothien-3-yl, 1 –piperazinyl, 2-piperazinyl, and the like.

[0098] The terms "halo" or "halogen," by themselves or as part of another substituent, mean, unless otherwise stated, a fluorine, chlorine, bromine, or iodine atom. Additionally, terms such as "haloalkyl," are meant to include monohaloalkyl and polyhaloalkyl. For example, the term "halo(C_1-C_4)alkyl" is mean to include, but not be limited to, trifluoromethyl, 2,2,2-trifluoroethyl, 4-chlorobutyl, 3-bromopropyl, and the like.

[0099] The term "aryl" means, unless otherwise stated, a polyunsaturated, aromatic, substituent that can be a single ring or multiple rings (preferably from 1 to 3 rings), which are fused together or linked covalently. The term "heteroaryl" refers to aryl groups (or rings) that contain from one to four heteroatoms selected from N, O, and S, wherein the nitrogen and sulfur atoms are optionally oxidized, and the nitrogen atom(s) are optionally quaternized. A heteroaryl group can be attached to the remainder of the molecule through a heteroatom. Non-limiting examples of aryl and heteroaryl groups include phenyl, 1-naphthyl, 2-naphthyl, 4-biphenyl, 1-pyrrolyl, 2-pyrrolyl, 3-pyrrolyl, 3-pyrazolyl, 2-imidazolyl, 4-imidazolyl, pyrazinyl, 2-oxazolyl, 4-oxazolyl, 2-phenyl-4-oxazolyl, 5-oxazolyl, 3-isoxazolyl, 4-isoxazolyl, 5-isoxazolyl, 2-thiazolyl, 4-thiazolyl, 5-thiazolyl, 2-furyl, 3-furyl, 2-thienyl, 3-thienyl, 2-pyridyl, 3-pyridyl, 4-pyridyl, 2-pyrimidyl, 4-pyrimidyl, 5-benzothiazolyl, purinyl, 2-benzimidazolyl, 5-indolyl, 1-isoquinolyl, 5-isoquinolyl, 2-quinoxalinyl, 5-quinoxalinyl, 3-quinolyl, tetrazolyl, benzo[b]furanyl, benzo[b]thienyl, 2,3-dihydrobenzo[1,4]dioxin-6-yl, benzo[1,3]dioxol-5-yl and 6-quinolyl. Substituents for each of the above noted aryl and heteroaryl ring systems are selected from the group of acceptable substituents described below.

[0100] For brevity, the term "aryl" when used in combination with other terms (*e.g.*, aryloxy, arylthioxy, arylalkyl) includes both aryl and heteroaryl rings as defined above. Thus, the term "arylalkyl" is meant to include those radicals in which an aryl group is

attached to an alkyl group (*e.g.*, benzyl, phenethyl, pyridylmethyl and the like) including those alkyl groups in which a carbon atom (*e.g.*, a methylene group) has been replaced by, for example, an oxygen atom (*e.g.*, phenoxymethyl, 2-pyridyloxymethyl, 3-(1-naphthyloxy)propyl, and the like).

[0101] Each of the above terms (*e.g.*, "alkyl," "heteroalkyl," "aryl" and "heteroaryl") is meant to include both substituted and unsubstituted forms of the indicated radical. Preferred substituents for *each* type of radical are provided below.

[0102] Substituents for the alkyl and heteroalkyl radicals (including those groups often referred to as alkylene, alkenyl, heteroalkylene, heteroalkenyl, alkynyl, cycloalkyl, heterocycloalkyl, cycloalkenyl, and heterocycloalkenyl) are generically referred to as "alkyl group substituents," and they can be one or more of a variety of groups selected from, but not limited to: -OR', =O, =NR', =N-OR', -NR'R", -SR', -halogen, -SiR'R"R"', -OC(O)R', -C(O)R', $-CO_2R'$, -CONR'R", -OC(O)NR'R", -NR"C(O)R', -NR'-C(O)NR"R"', -NR"$C(O)_2R'$, -NR-C(NR'R"R"')=NR"", -NR-C(NR'R")=NR"', -S(O)R', $-S(O)_2R'$, $-S(O)_2NR'R"$, $-NRSO_2R'$, -CN and $-NO_2$ in a number ranging from zero to (2m'+1), where m' is the total number of carbon atoms in such radical. R', R", R"' and R"" each preferably independently refer to hydrogen, substituted or unsubstituted heteroalkyl, substituted or unsubstituted aryl, e.g., aryl substituted with 1-3 halogens, substituted or unsubstituted alkyl, alkoxy or thioalkoxy groups, or arylalkyl groups. When a compound of the invention includes more than one R group, for example, each of the R groups is independently selected as are each R', R", R"' and R"" groups when more than one of these groups is present. When R' and R" are attached to the same nitrogen atom, they can be combined with the nitrogen atom to form a 5-, 6-, or 7-membered ring. For example, -NR'R" is meant to include, but not be limited to, 1-pyrrolidinyl and 4-morpholinyl. From the above discussion of substituents, one of skill in the art will understand that the term "alkyl" is meant to include groups including carbon atoms bound to groups other than hydrogen groups, such as haloalkyl (*e.g.*, $-CF_3$ and $-CH_2CF_3$) and acyl (*e.g.*, $-C(O)CH_3$, $-C(O)CF_3$, $-C(O)CH_2OCH_3$, and the like).

[0103] Similar to the substituents described for the alkyl radical, substituents for the aryl and heteroaryl groups are generically referred to as "aryl group substituents." The substituents are selected from, for example: halogen, -OR', =O, =NR', =N-OR', -NR'R", -SR', -halogen, -SiR'R"R"', -OC(O)R', -C(O)R', $-CO_2R'$, -CONR'R", -OC(O)NR'R", -NR"C(O)R', -NR'-C(O)NR"R"', -NR"$C(O)_2R'$, -NR-C(NR'R"R"')=NR"", -NR-C(NR'R")=NR"', -S(O)R', $-S(O)_2R'$, $-S(O)_2NR'R"$, $-NRSO_2R'$, -CN and $-NO_2$, -R', -

N_3, -CH(Ph)$_2$, fluoro(C_1-C_4)alkoxy, and fluoro(C_1-C_4)alkyl, in a number ranging from zero to the total number of open valences on the aromatic ring system; and where R', R", R"' and R"" are preferably independently selected from hydrogen, substituted or unsubstituted alkyl, substituted or unsubstituted heteroalkyl, substituted or unsubstituted aryl and substituted or unsubstituted heteroaryl. When a compound of the invention includes more than one R group, for example, each of the R groups is independently selected as are each R', R", R"' and R"" groups when more than one of these groups is present. In the schemes that follow, the symbol X represents "R" as described above.

[0104] Two of the substituents on adjacent atoms of the aryl or heteroaryl ring may optionally be replaced with a substituent of the formula –T-C(O)-(CRR')$_u$-U-, wherein T and U are independently –NR-, -O-, -CRR'- or a single bond, and u is an integer of from 0 to 3. Alternatively, two of the substituents on adjacent atoms of the aryl or heteroaryl ring may optionally be replaced with a substituent of the formula –A-(CH$_2$)$_r$-B-, wherein A and B are independently –CRR'-, -O-, -NR-, -S-, -S(O)-, -S(O)$_2$-, -S(O)$_2$NR'- or a single bond, and r is an integer of from 1 to 4. One of the single bonds of the new ring so formed may optionally be replaced with a double bond. Alternatively, two of the substituents on adjacent atoms of the aryl or heteroaryl ring may optionally be replaced with a substituent of the formula –(CRR')$_z$-X-(CR"R"')$_d$-, where z and d are independently integers of from 0 to 3, and X is -O-, -NR'-, -S-, -S(O)-, -S(O)$_2$-, or –S(O)$_2$NR'-. The substituents R, R', R" and R"' are preferably independently selected from hydrogen or substituted or unsubstituted (C_1-C_6)alkyl.

[0105] As used herein, the term "heteroatom" is meant to include oxygen (O), nitrogen (N), sulfur (S) and silicon (Si).

[0106] As used herein, Factor VII peptide refers to both Factor VII and Factor VIIa peptides. The terms generally refer to variants and mutants of these peptides, including addition, deletion, substitution and fusion protein mutants. Where both Factor VII and Factor VIIa are used, the use is intended to be illustrative of two species of the genus "Factor VII peptide".

[0107] The invention is meant to include salts of the compounds of the invention which are prepared with relatively nontoxic acids or bases, depending on the particular substituents found on the compounds described herein. When compounds of the present invention contain relatively acidic functionalities, base addition salts can be obtained by contacting the neutral form of such compounds with a sufficient amount of the desired base, either neat or in

a suitable inert solvent. Examples of base addition salts include sodium, potassium, lithium, calcium, ammonium, organic amino, or magnesium salt, or a similar salt. When compounds of the present invention contain relatively basic functionalities, acid addition salts can be obtained by contacting the neutral form of such compounds with a sufficient amount of the desired acid, either neat or in a suitable inert solvent. Examples of acid addition salts include those derived from inorganic acids like hydrochloric, hydrobromic, nitric, carbonic, monohydrogencarbonic, phosphoric, monohydrogenphosphoric, dihydrogenphosphoric, sulfuric, monohydrogensulfuric, hydriodic, or phosphorous acids and the like, as well as the salts derived from relatively nontoxic organic acids like acetic, propionic, isobutyric, maleic, malonic, benzoic, succinic, suberic, fumaric, lactic, mandelic, phthalic, benzenesulfonic, p-tolylsulfonic, citric, tartaric, methanesulfonic, and the like. Also included are salts of amino acids such as arginate and the like, and salts of organic acids like glucuronic or galactunoric acids and the like (see, for example, Berge et al., "Pharmaceutical Salts", Journal of Pharmaceutical Science 66: 1-19 (1977)). Certain specific compounds of the present invention contain both basic and acidic functionalities that allow the compounds to be converted into either base or acid addition salts.

[0108] The neutral forms of the compounds are preferably regenerated by contacting the salt with a base or acid and isolating the parent compounds in the conventional manner. The parent form of the compound differs from the various salt forms in certain physical properties, such as solubility in polar solvents.

[0109] "Salt counterion", as used herein, refers to positively charged ions that associate with a compound of the invention when one of its moieties is negatively charged (e.g. COO-). Examples of salt counterions include H^+, H_3O^+, ammonium, potassium, calcium, lithium, magnesium and sodium.

[0110] As used herein, the term "CMP-SA-PEG" is a cytidine monophosphate molecule which is conjugated to a sialic acid which comprises a polyethylene glycol moiety. If a length of the polyethylene glycol chain is not specified, then any PEG chain length is possible (e.g. 1KDa, 2KDa, 5 KDa, 10KDa, 20KDa, 30KDa, 40KDa). An exemplary CMP-SA-PEG is compound 5 in Scheme 1.

I. ***Introduction***

[0111] The present invention encompasses a method for the remodeling and modification of Factor VII. The blood coagulation pathway is a complex reaction comprising many

events. An intermediate event in this pathway is Factor VII, a proenzyme that participates in the extrinsic pathway of blood coagulation by converting (upon its activation to Factor VIIa) Factor X to Xa in the presence of tissue factor and calcium ions. Factor Xa in turn then converts prothrombin to thrombin in the presence of Factor Va, calcium ions and phospholipid. The activation of Factor X to Factor Xa is an event shared by both the intrinsic and extrinsic blood coagulation pathways, and therefore, Factor VIIa can be used for the treatment of patients with deficiencies or inhibitors of Factor VIII. There is also evidence to suggest that Factor VIIa may participate in the intrinsic pathway as well therefore increasing the prominence and importance of the role of Factor VII/Factor VIIa in blood coagulation.

[0112] Factor VII is a single-chain glycoprotein which circulates in the blood as an inactive zymogen. Exemplary nucleotide and amino acid sequences of Factor VIIa are provided in **FIG. 5**. Activation of Factor VII to VIIa may be catalyzed by several different plasma proteases, such as Factor XIIa. Activation of Factor VII occurs when the Factor VII peptide backbone is cleaved at asparagine 152. The activated product, Factor VIIa, is a glycoprotein which comprises a heavy chain and a light chain held together by at least one disulfide bond. Further, modified Factor VII molecules that cannot be converted to Factor VIIa have been described, and are useful as anti-coagulation remedies, such as in the case of blood clots, thrombosis, and the like. Given the importance of Factor VII in the blood coagulation pathway, and its use as a treatment for both increased and decreased levels of coagulation, it follows that a molecule that has a longer biological half-life, increased potency, and in general, a therapeutic profile more similar to wild-type Factor VII as it is synthesized and secreted in the healthy human would be beneficial and useful as a treatment for blood coagulation disorders.

[0113] While Factor VII is an important and useful compound for therapeutic applications, present methods for the production of Factor VII from recombinant cells result in a product with a rather short biological half-life and a non-optimal glycosylation pattern that could potentially lead to immunogenicity, loss of function, an increased need for both larger and more frequent doses in order to achieve the same effect, and the like.

[0114] To improve the effectiveness of recombinant Factor VII/Factor VIIa used for therapeutic purposes, the present invention provides conjugates of glycosylated and unglycosylated Factor VII/Factor VIIa peptides with a modifying group. The modifying groups can be selected from polymeric modifying groups such as, e.g., PEG (m-PEG), PPG

(m-PPG), etc., therapeutic moieties, diagnostic moieties, targeting moieties and the like. Modification of the Factor VII/Factor VIIa peptides, e.g., with a water-soluble polymeric modifying group can improve the stability and retention time of the recombinant Factor VII/Factor VIIa in a patient's circulation, and/or reduce the antigenicity of recombinant Factor VII/Factor VIIa.

[0115] The peptide conjugates of the invention can be formed by the enzymatic attachment of a modified sugar to the glycosylated or unglycosylated peptide. A glycosylation site and/or a modified glycosyl group provides a locus for conjugating a modified sugar bearing a modifying group to the peptide, e.g., by glycoconjugation.

[0116] The methods of the invention also make it possible to assemble peptide conjugates and glycopeptide conjugates that have a substantially homogeneous derivatization pattern. The enzymes used in the invention are generally selective for a particular amino acid residue, combination of amino acid residues, particular glycosyl residues, or combination of glycosyl residues of the peptide. The methods are also practical for large-scale production of peptide conjugates. Thus, the methods of the invention provide a practical means for large-scale preparation of peptide conjugates having preselected uniform derivatization patterns. The methods are particularly well suited for modification of therapeutic peptides, including but not limited to, glycopeptides that are incompletely glycosylated during production in cell culture cells (*e.g.*, mammalian cells, insect cells, plant cells, fungal cells, yeast cells, or prokaryotic cells) or transgenic plants or animals.

[0117] The Factor VII/Factor VIIa peptide conjugates can be produced as pharmaceutical formulations comprising a peptide conjugate as well as a pharmaceutically acceptable carrier. The Factor VII/Factor VIIa peptide conjugates may be administered to a patient selected from the group consisting of a hemophiliac patient having a bleeding episode, a patient having Hemophilia A, a patient with Hemophilia B, a patient having Hemophilia A, wherein the patient also has antibodies to Factor VIII, a patient having Hemophilia B, wherein the patient also has antibodies to Factor IX, a patient having liver cirrhosis, a cirrhotic patient having an orthotopic liver transplant, a cirrhotic patient having upper gastrointestinal bleeding, a patient having a bone marrow transplant, a patient having a liver resection, a patient having a partial hepatectomy, a patient undergoing pelvic-acetabular fracture reconstruction, a patient bleeding from an acute intercerebral hemmorage, a patient undergoing allogeneic stem cell transplantation, a patient bleeding from traumatic brain injury, a patient bleeding in an

emergency, a patient having bleeding from trauma, a patient undergoing variceal bleeding, a patient bleeding from elective surgery, a patient bleeding from cardiac surgery, a patient bleeding from spinal surgery, a liver resection a liver resection a liver resection. In an exemplary embodiment, the patient is a human patient.

[0118] The present invention also provides conjugates of glycosylated and unglycosylated peptides with increased therapeutic half-life due to, for example, reduced clearance rate, or reduced rate of uptake by the immune or reticuloendothelial system (RES). Moreover, the methods of the invention provide a means for masking antigenic determinants on peptides, thus reducing or eliminating a host immune response against the peptide. Selective attachment of targeting agents can also be used to target a peptide to a particular tissue or cell surface receptor that is specific for the particular targeting agent.

[0119] Determining optimal conditions for the preparation of Factor VII/Factor VIIa conjugates with water-soluble polymers, e.g., involves the optimization of numerous parameters, which are dependent on the identity of the peptide and of the water-soluble polymer. For example, when the polymer is poly(ethylene glycol), e.g., a branched poly(ethylene glycol), a balance is preferably established between the amount of polymer utilized in the reaction and the viscosity of the reaction mixture attributable to the presence of the polymer: if the polymer is too highly concentrated, the reaction mixture becomes viscous, slowing the rate of mass transfer and reaction.

[0120] Furthermore, though it is intuitively apparent to add an excess of enzyme, the present inventors have recognized that, when the enzyme is present in too great of an excess, the excess enzyme becomes a contaminant whose removal requires extra purification steps and material and unnecessarily increases the cost of the final product.

[0121] Moreover, it is generally desired to produce a peptide with a controlled level of modification. In some instances, it is desireable to add one modified sugar preferentially. In other instances, it is desireable to add two modified sugars preferentially. Thus, the reaction conditions are preferably controlled to influence the degree of conjugation of the modifying groups to the peptide.

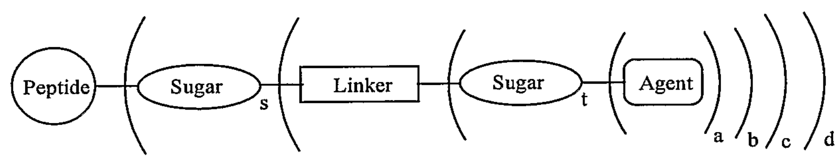
[0122] The present invention provides conditions under which the yield of a Factor VII/Factor VIIa peptide, having the desired level of conjugation, is maximized. The conditions in the exemplary embodiments of the inventions also recognize the expense of the various reagents and the materials and time necessary to purify the product: the reaction

conditions set forth herein are optimized to provide excellent yields of the desired product, while minimizing waste of costly reagents.

II. The Compositions of Matter/Peptide Conjugates

[0123] In a first aspect, the present invention provides a conjugate between a modified sugar and a Factor VII/Factor VIIa peptide. The present invention also provides a conjugate between a modifying group and a Factor VII/Factor VIIa peptide. A peptide conjugate can have one of several forms. In an exemplary embodiment, a peptide conjugate can comprise a Factor VII/Factor VIIa peptide and a modifying group linked to an amino acid of the peptide through a glycosyl linking group. In another exemplary embodiment, a peptide conjugate can comprise a Factor VII/Factor VIIa peptide and a modifying group linked to a glycosyl reside of the peptide through a glycosyl linking group. In another exemplary embodiment, the peptide conjugate can comprise a Factor VII/Factor VIIa peptide and a glycosyl linking group which is bound to both a glycopeptide carbohydrate and directly to an amino acid residue of the peptide backbone. In yet another exemplary embodiment, a peptide conjugate can comprise a Factor VII/Factor VIIa peptide and a modifying group linked directly to an amino acid residue of the peptide. In this embodiment, the peptide conjugate may not comprise a glycosyl group. In any of these embodiments, the Factor VII/Factor VIIa peptide may or not be glycosylated.

[0124] The conjugates of the invention will typically correspond to the general structure:



in which the symbols a, b, c, d and s represent a positive, non-zero integer; and t is either 0 or a positive integer. The "agent", or modifying group, can be a therapeutic agent, a bioactive agent, a detectable label, a polymeric modifying group such as a water-soluble polymer (*e.g.*, PEG, m-PEG, PPG, and m-PPG) or the like. The "agent", or modifying group, can be a peptide, *e.g.*, enzyme, antibody, antigen, etc. The linker can be any of a wide array of linking groups, *infra*. Alternatively, the linker may be a single bond or a "zero order linker."

II. A. Peptide

[0125] Factor VII is a single-chain polypeptide which is about 406 amino acids in length and has a molecular weight of approximately 50 KDa. Conversion of Factor VII to Factor VIIa occurs when the Factor VII peptide backbone is cleaved at asparagine 152. Factor VII

and/or Factor VIIa peptides contain two N-glycan sites: one is located at asparagine 145 and the other is located at asparagine 322. The N-glycan site at asparagine 145 is located on the light chain of FVIIa, while the N-glycan site at asparagine 322 is located on the heavy chain of FVIIa. Factor VII and/or Factor VIIa peptides contain two O-glycan sites.

[0126] Factor VII or Factor VIIa has been cloned and sequenced. In an exemplary embodiment, the Factor VIIa peptide has the sequence presented in SEQ ID NO:1:

[0127] The present invention should in no way be construed as limited to the Factor VII nucleic acid and amino acid sequences set forth herein. Use of Factor VII/Factor VIIa peptides of other sequences that are mutated to increase or decrease a property or modify a structural feature of the peptide are within the scope of the invention. For example, mutant Factor VII/Factor VIIa peptides of use in the invention include those that are provided with additional O-glycosylation sites or such sites at other positions. Moreover, mutant peptides that include one or more N-glycosylation site are of use in the invention. Variants of Factor VII are described in, for example, U.S. Patent Nos. 4,784,950 and 5,580,560, in which lysine-38, lysine-32, arginine--290, arginine-341, isoleucine-42, tyrosine-278, and tyrosine-332 is replaced by a variety of amino acids. Further, U.S. Patent Nos. 5,861,374, 6,039,944, 5,833,982, 5,788,965, 6,183,743, 5,997,864, and 5,817,788 describe Factor VII variants that are not cleaved to form Factor VIIa. The skilled artisan will recognize that the blood coagulation pathway and the role of Factor VII therein are well known, and therefore many variants, both naturally occurring and engineered, as described above, are included in the present invention. In an exemplary embodiment, a peptide having Factor VII/Factor VIIa activity has an amino acid sequence that is at least about 95% homologous to the amino acid sequences set forth herein. Perferably, the amino acid sequence is at least about 96%, 97%, 98% or 99% homologous to the amino acid sequences set forth herein.

[0128] In an exemplary embodiment, the amino acid residue to which the glycosyl linking group is attached is a member selected from serine, threonine and asparagine. In another exemplary embodiment, the peptide has a sequence of SEQ. ID. NO 1. In another exemplary embodiment, the amino acid residue is a member selected from Asn 145, Asn 322 and combinations thereof. In another exemplary embodiment, the peptide is a bioactive Factor VII/Factor VIIa peptide.

[0129] In yet another exemplary embodiment, the modified sugar and/or PEG moiety on the Factor VIIa peptide conjugate is located on the light chain. In yet another exemplary

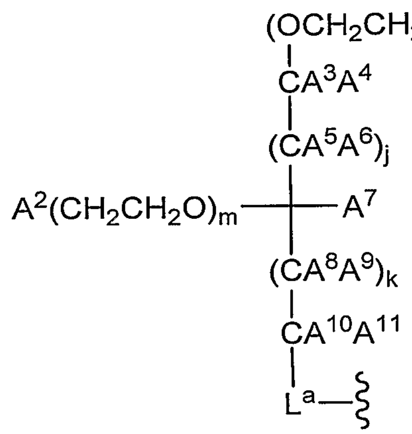
embodiment, the modified sugar and/or PEG moiety on the Factor VIIa peptide conjugate is predominantly on the heavy chain. In yet another exemplary embodiment, in a population of Factor VIIa peptide conjugates, the light chains predominantly contain a modified sugar and/or PEG moiety. In yet another exemplary embodiment, in a population of Factor VIIa peptide conjugates, the heavy chains predominantly contain a modified sugar and/or PEG moiety.

[0130] In another exemplary embodiment, the ratio of light chain: heavy chain functionalization in the population is about 33:66. In another exemplary embodiment, the ratio of light chain: heavy chain functionalization in the population is about 35:65. In another exemplary embodiment, the ratio of light chain: heavy chain functionalization in the population is about 40:60. In another exemplary embodiment, the ratio of light chain: heavy chain functionalization in the population is about 45:55. In another exemplary embodiment, the ratio is about 50:50. In another exemplary embodiment, the ratio is about 55:45. In another exemplary embodiment, the ratio is about 60:40. In another exemplary embodiment, the ratio is about 65:35. In another exemplary embodiment, the ratio is about 66:33. In another exemplary embodiment, the ratio is about 70:30. In another exemplary embodiment, the ratio is about 75:25. In another exemplary embodiment, the ratio is about 80:20. In another exemplary embodiment, the ratio is about 85:15. In another exemplary embodiment, the ratio is about 90:10. In another exemplary embodiment, the ratio of light chain: heavy chain functionalization in the population is greater than about 90:10.

[0131] Methods for the expression and to determine the activity of Factor VII/Factor VIIa are well known in the art, and are described in, for example, U.S. Patent No. 4,784,950. Briefly, expression of Factor VII, or variants thereof, can be accomplished in a variety of both prokaryotic and eukaryotic systems, including *E. coli*, CHO cells, BHK cells, insect cells using a baculovirus expression system, all of which are well known in the art.

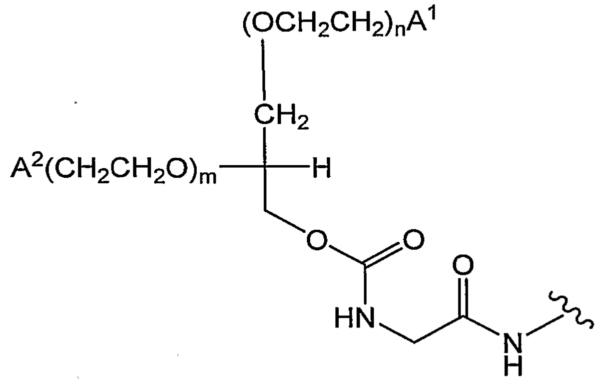
[0132] Assays for the activity of a Factor VII/Factor VIIa peptide conjugate prepared according to the methods of the present invention can be accomplished using methods well known in the art. As a non-limiting example, Quick et al. (Hemorragic Disease and Thrombosis, 2nd ed., Leat Febiger, Philadelphia, 1966), describes a one-stage clotting assay useful for determining the biological activity of a Factor VII molecule prepared according to the methods of the present invention.

[0133] The peptides used in the invention are not limited to Factor VII/Factor VIIa when the modifying group is:

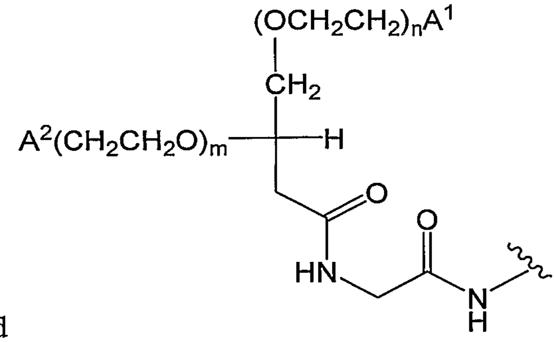


In these cases, the peptide in the peptide conjugate is a member selected from the peptides in **FIG. 13**. In these cases, the peptide in the peptide conjugate is a member selected from Factor VII, Factor VIIa, Factor VIII, Factor IX, Factor X, Factor XI, a peptide which is a member selected from erythropoietin, granulocyte colony stimulating factor (G-CSF), Granulocyte-Macrophage Colony Stimulating Factor (GM-CSF)interferon alpha, interferon beta, interferon gamma, α_1-antitrypsin (ATT, or α-1 protease inhibitor, glucocerebrosidase, Tissue-Type Plasminogen Activator (TPA), Interleukin-2 (IL-2), urokinase, human DNase, insulin, Hepatitis B surface protein (HbsAg), human growth hormone, TNF Receptor-IgG Fc region fusion protein (Enbrel™), anti-HER2 monoclonal antibody (Herceptin™), monoclonal antibody to Protein F of Respiratory Syncytial Virus (Synagis™), monoclonal antibody to TNF-α (Remicade™), monoclonal antibody to glycoprotein IIb/IIIa (Reopro™), monoclonal antibody to CD20 (Rituxan™), anti-thrombin III (AT III), human Chorionic Gonadotropin (hCG), alpha-galactosidase (Fabrazyme™), alpha-iduronidase (Aldurazyme™), follicle stimulating hormone, beta-glucosidase, anti-TNF-alpha monoclonal antibody (MLB 5075), glucagon-like peptide-1 (GLP-1), beta-glucosidase (MLB 5064), alpha-galactosidase A (MLB 5082) and fibroblast growth factor.

[0134] In an exemplary embodiment, the polymeric modifying group has a structure according to the following formulae:

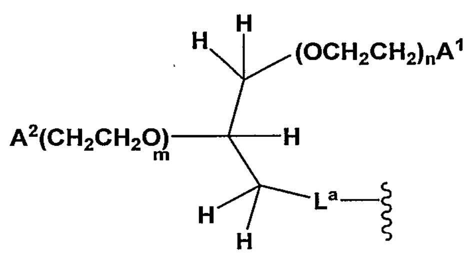


and



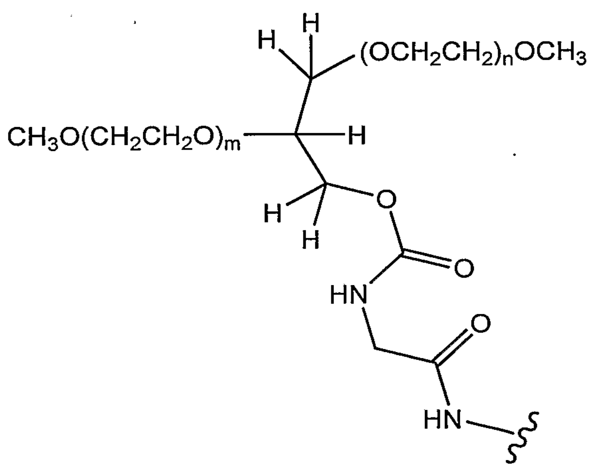
.

[0135] The peptides used in the invention are also not limited to Factor VII or Factor VIIa when the modifying group is:

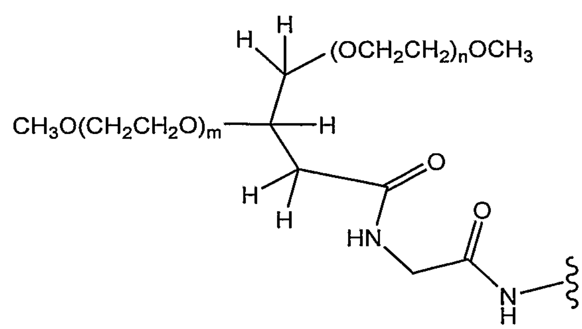


In an exemplary embodiment, A^1 and A^2 are each members selected from -OH and $-OCH_3$.

[0136] Exemplary polymeric modifying groups according to this embodiment include:



and



[0137] In an exemplary embodiment, in which the modifying group is a branched water-soluble polymer, such as those shown above, it is generally preferred that the concentration of sialidase is about 1.5 to about 2.5 U/L of reaction mixture. More preferably the amount of sialidase is about 2 U/L.

[0138] In another exemplary embodiment, about 5 to about 9 grams of peptide substrate is contacted with the amounts of sialidase set forth above.

[0139] The modified sugar is present in the reaction mixture in an amount from about 1 gram to about 6 grams, preferably from about 3 grams to about 4 grams. It is generally preferred to maintain the concentration of a modified sugar having a branched water-soluble polymer modifying moiety, e.g., the moiety shown above, at less than about 0.5 mM. In a preferred embodiment, the modifying group is a branched poly(ethylene glycol) having a molecular weight from about 20 KDa to about 60 KDa, more preferably, from about 30 KDa to about 50 KDa, and even more preferably about 40 KDa. An exemplary modifying group having a molecular weight of about 40 KDa is one that is from about 35 KDa to about 45 KDa.

[0140] Regarding the glycosyltransferase concentration, in a presently preferred embodiment, using the modifying group set forth above, the ratio of glycosyltransferase to peptide is about 40 μg/mL transferase to about 200 μM peptide.

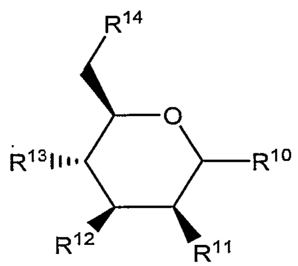
II. B. *Modified Sugar*

[0141] In an exemplary embodiment, the peptides of the invention are reacted with a modified sugar, thus forming a peptide conjugate. A modified sugar comprises a "sugar donor moiety" as well as a "sugar transfer moiety". The sugar donor moiety is any portion of the modified sugar that will be attached to the peptide, either through a glycosyl moiety or amino acid moiety, as a conjugate of the invention. The sugar donor moiety includes those atoms that are chemically altered during their conversion from the modified sugar to the glycosyl linking group of the peptide conjugate. The sugar transfer moiety is any portion of the modified sugar that will be not be attached to the peptide as a conjugate of the invention. For example, a modified sugar of the invention is the PEGylated sugar nucleotide, PEG-sialic acid CMP. For PEG-sialic acid CMP, the sugar donor moiety, or PEG-sialyl donor moiety, comprises PEG-sialic acid while the sugar transfer moiety, or sialyl transfer moiety, comprises CMP.

[0142] In modified sugars of use in the invention, the saccharyl moiety is preferably a saccharide, a deoxy-saccharide, an amino-saccharide, or an N-acyl saccharide. The term "saccharide" and its equivalents, "saccharyl," "sugar," and "glycosyl" refer to monomers, dimers, oligomers and polymers. The sugar moiety is also functionalized with a modifying group. The modifying group is conjugated to the saccharyl moiety, typically, through

conjugation with an amine, sulfhydryl or hydroxyl, e.g., primary hydroxyl, moiety on the sugar. In an exemplary embodiment, the modifying group is attached through an amine moiety on the sugar, e.g., through an amide, a urethane or a urea that is formed through the reaction of the amine with a reactive derivative of the modifying group.

[0143] Any saccharyl moiety can be utilized as the sugar donor moiety of the modified sugar. The saccharyl moiety can be a known sugar, such as mannose, galactose or glucose, or a species having the stereochemistry of a known sugar. The general formulae of these modified sugars are:



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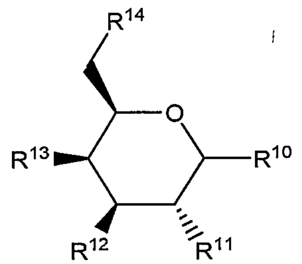
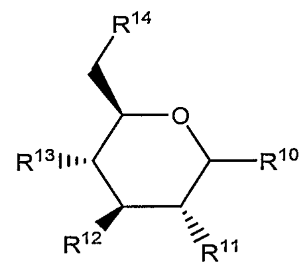


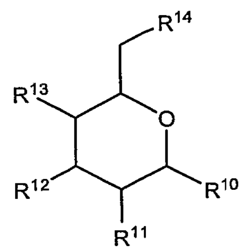
; and



Other saccharyl moieties that are useful in forming the compositions of the invention include, but are not limited to fucose and sialic acid, as well as amino sugars such as glucosamine, galactosamine, mannosamine, the 5-amine analogue of sialic acid and the like. The saccharyl moiety can be a structure found in nature or it can be modified to provide a site for conjugating the modifying group. For example, in one embodiment, the modified sugar provides a sialic acid derivative in which the 9-hydroxy moiety is replaced with an amine. The amine is readily derivatized with an activated analogue of a selected modifying group.

[0144] Examples of modified sugars of use in the invention are described in PCT Patent Application No. PCT/US05/002522, which is herein incorporated by reference.

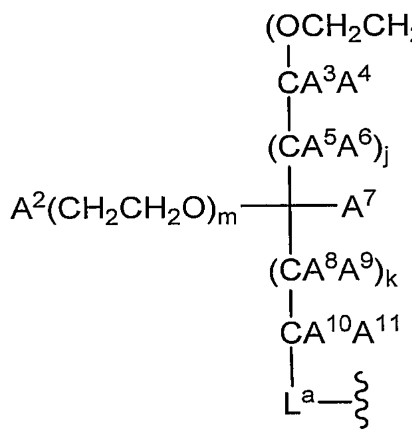
[0145] In a further exemplary embodiment, the invention utilizes modified sugars in which the 6-hydroxyl position is converted to the corresponding amine moiety, which bears a linker-modifying group cassette such as those set forth above. Exemplary glycosyl groups that can be used as the core of these modified sugars include Gal, GalNAc, Glc, GlcNAc, Fuc, Xyl, Man, and the like. A representative modified sugar according to this embodiment has the formula:



in which R^{11}-R^{14} are members independently selected from H, OH, $C(O)CH_3$, NH, and NH $C(O)CH_3$. R^{10} is a link to another glycosyl residue (-O-glycosyl) or to an amino acid of the Factor VII/Factor VIIa peptide (-NH-(Factor VII/Factor VIIa)). R^{14} is OR^1, NHR^1 or NH-L-R^1. R^1 and NH-L-R^1 are as described above.

II. C. *Glycosyl Linking Groups*

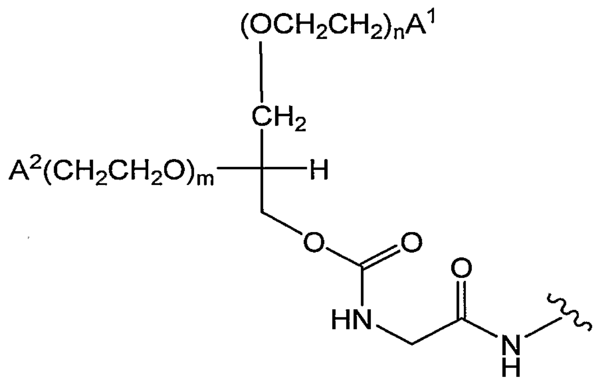
[0146] In an exemplary embodiment, the invention provides a peptide conjugate formed between a modified sugar of the invention and a Factor VII/Factor VIIa peptide. In another exemplary embodiment, when the modifying group on the modified sugar is



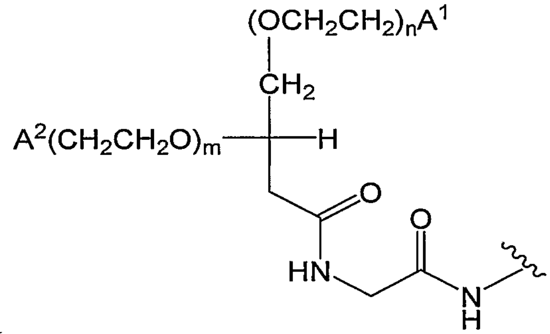
the peptide in the peptide conjugate is a member selected from the peptides in **FIG. 13**. In yet another exemplary embodiment, the peptide in the peptide conjugate is a member selected from Factor VII, Factor VIIa, Factor VIII, Factor IX, Factor X, Factor XI, erythropoietin, granulocyte colony stimulating factor (G-CSF), Granulocyte-Macrophage Colony Stimulating Factor (GM-CSF), interferon alpha, interferon beta, interferon gamma, α_1-antitrypsin (ATT, or α-1 protease inhibitor, glucocerebrosidase, Tissue-Type Plasminogen Activator (TPA), Interleukin-2 (IL-2), urokinase, human DNase, insulin, Hepatitis B surface protein (HbsAg), human growth hormone, TNF Receptor-IgG Fc region fusion protein (Enbrel™), anti-HER2 monoclonal antibody (Herceptin™), monoclonal antibody to Protein F of Respiratory Syncytial Virus (Synagis™), monoclonal antibody to TNF-α (Remicade™), monoclonal antibody to glycoprotein IIb/IIIa (Reopro™), monoclonal antibody to CD20 (Rituxan™), anti-thrombin III (AT III), human Chorionic Gonadotropin (hCG), alpha-

galactosidase (Fabrazyme™), alpha-iduronidase (Aldurazyme™), follicle stimulating hormone, beta-glucosidase, anti-TNF-alpha monoclonal antibody (MLB 5075), glucagon-like peptide-1 (GLP-1), beta-glucosidase (MLB 5064), alpha-galactosidase A (MLB 5082) and fibroblast growth factor;. In this embodiment, the sugar donor moiety (such as the saccharyl moiety and the modifying group) of the modified sugar becomes a "glycosyl linking group". The "glycosyl linking group" can alternatively refer to the glycosyl moiety which is interposed between the peptide and the modifying group.

[0147] In an exemplary embodiment, the polymeric modifying group has a structure according to the following formulae:

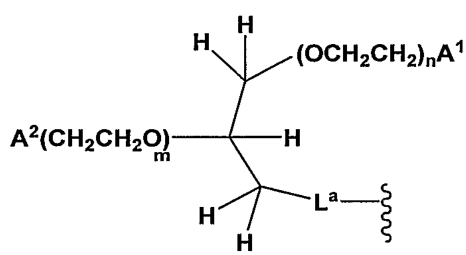


and



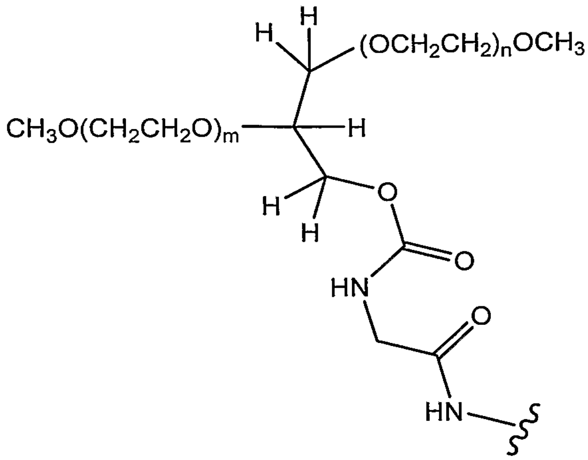
.

[0148] In an exemplary embodiment, modifying group on the modified sugar is:

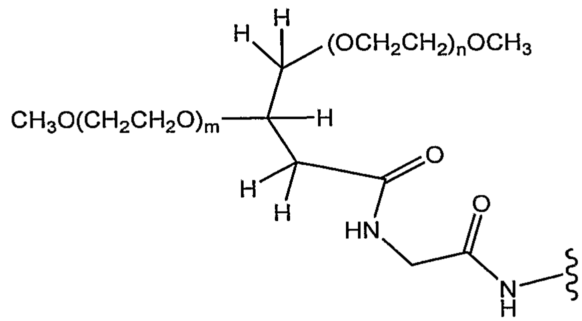


In an exemplary embodiment, A^1 and A^2 are each members selected from -OH and $-OCH_3$.

[0149] Exemplary polymeric modifying groups according to this embodiment include:

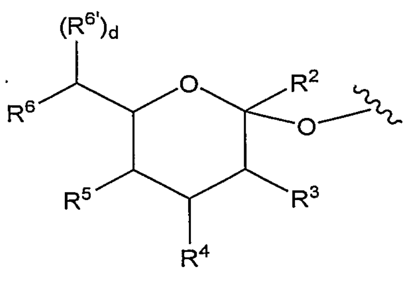


and



[0150] Due to the versatility of the methods available for adding and/or modifying glycosyl residues on a peptide, the glycosyl linking groups can have substantially any structure. In the discussion that follows, the invention is illustrated by reference to the use of selected derivatives of furanose and pyranose. Those of skill in the art will recognize that the focus of the discussion is for clarity of illustration and that the structures and compositions set forth are generally applicable across the genus of glycosyl linking groups and modified sugars. The glycosyl linking group can comprise virtually any mono- or oligo-saccharide. The glycosyl linking groups can be attached to an amino acid either through the side chain or through the peptide backbone. Alternatively the glycosyl linking groups can be attached to the peptide through a saccharyl moiety. This saccharyl moiety can be a portion of an O-linked or N-linked glycan structure on the peptide.

[0151] In an exemplary embodiment, the invention provides a peptide conjugate comprising an intact glycosyl linking group having a formula that is selected from:



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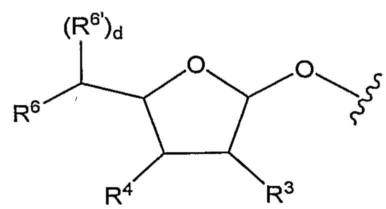
; and

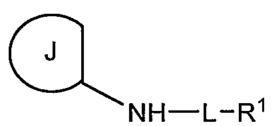


II

In Formulae I R^2 is H, CH_2OR^7, $COOR^7$ or OR^7, in which R^7 represents H, substituted or unsubstituted alkyl or substituted or unsubstituted heteroalkyl. When $COOR^7$ is a carboxylic acid or carboxylate, both forms are represented by the designation of the single structure

COO^- or COOH. In Formulae I and II, the symbols R^3, R^4, R^5, R^6 and $R^{6'}$ independently represent H, substituted or unsubstituted alkyl, OR^8, $NHC(O)R^9$. The index d is 0 or 1. R^8 and R^9 are independently selected from H, substituted or unsubstituted alkyl, substituted or unsubstituted heteroalkyl, sialic acid or polysialic acid. At least one of R^3, R^4, R^5, R^6 or $R^{6'}$ includes a modifying group. This modifying group can be a polymeric modifying moiety *e.g.*, PEG, linked through a bond or a linking group. In an exemplary embodiment, R^6 and $R^{6'}$, together with the carbon to which they are attached are components of the pyruvyl side chain of sialic acid. In a further exemplary embodiment, the pyruvyl side chain is functionalized with the polymeric modifying group. In another exemplary embodiment, R^6 and $R^{6'}$, together with the carbon to which they are attached are components of the side chain of sialic acid and the polymeric modifying group is a component of R^5.

[0152] In an exemplary embodiment, the invention utilizes a glycosyl linking group that has the formula:



in which J is a glycosyl moiety, L is a bond or a linker and R^1 is a modifying group, e.g., a polymeric modifying group. Exemplary bonds are those that are formed between an NH_2 moiety on the glycosyl moiety and a group of complementary reactivity on the modifying group. For example, when R^1 includes a carboxylic acid moiety, this moiety may be activated and coupled with the NH_2 moiety on the glycosyl residue affording a bond having the structure $NHC(O)R^1$. J is preferably a glycosyl moiety that is "intact", not having been degraded by exposure to conditions that cleave the pyranose or furanose structure, e.g. oxidative conditions, e.g., sodium periodate.

[0153] Exemplary linkers include alkyl and heteroalkyl moieties. The linkers include linking groups, for example acyl-based linking groups, e.g., -C(O)NH-, -OC(O)NH-, and the like. The linking groups are bonds formed between components of the species of the invention, e.g., between the glycosyl moiety and the linker (L), or between the linker and the modifying group (R^1). Other exemplary linking groups are ethers, thioethers and amines. For example, in one embodiment, the linker is an amino acid residue, such as a glycine residue. The carboxylic acid moiety of the glycine is converted to the corresponding amide by reaction with an amine on the glycosyl residue, and the amine of the glycine is converted

to the corresponding amide or urethane by reaction with an activated carboxylic acid or carbonate of the modifying group.

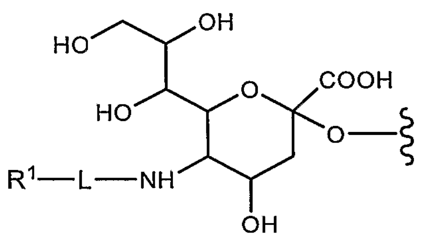
[0154] An exemplary species of NH-L-R^1 has the formula: $-NH\{C(O)(CH_2)_aNH\}_s\{C(O)(CH_2)_b(OCH_2CH_2)_cO(CH_2)_dNH\}_tR^1$, in which the indices s and t are independently 0 or 1. The indices a, b and d are independently integers from 0 to 20, and c is an integer from 1 to 2500. Other similar linkers are based on species in which an -NH moiety is replaced by another group, for example, -S, -O or $-CH_2$. As those of skill will appreciate one or more of the bracketed moieties corresponding to indices s and t can be replaced with a substituted or unsubstituted alkyl or heteroalkyl moiety.

[0155] More particularly, the invention utilizes compounds in which NH-L-R^1 is: $NHC(O)(CH_2)_aNHC(O)(CH_2)_b(OCH_2CH_2)_cO(CH_2)_dNHR^1$, $NHC(O)(CH_2)_b(OCH_2CH_2)_cO(CH_2)_dNHR^1$, $NHC(O)O(CH_2)_b(OCH_2CH_2)_cO(CH_2)_dNHR^1$, $NH(CH_2)_aNHC(O)(CH_2)_b(OCH_2CH_2)_cO(CH_2)_dNHR^1$, $NHC(O)(CH_2)_aNHR^1$, $NH(CH_2)_aNHR^1$, and NHR^1. In these formulae, the indices a, b and d are independently selected from the integers from 0 to 20, preferably from 1 to 5. The index c is an integer from 1 to about 2500.

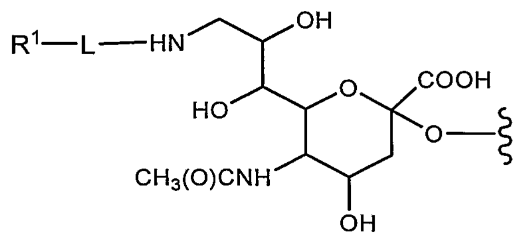
[0156] In an exemplary embodiment, c is selected such that the PEG moiety is approximately 1 kD, 5 kD, 10, kD, 15 kD, 20 kD, 25 kD, 30 kD, 35 kD, 40 kD or 45 kD.

[0157] For the purposes of convenience, the glycosyl linking groups in the remainder of this section will be based on a sialyl moiety. However, one of skill in the art will recognize that another glycosyl moiety, such as mannosyl, galactosyl, glucosyl, or fucosyl, could be used in place of the sialyl moiety.

[0158] In an exemplary embodiment, the glycosyl linking group is an intact glycosyl linking group, in which the glycosyl moiety or moieties forming the linking group are not degraded by chemical (e.g., sodium metaperiodate) or enzymatic (e.g., oxidase) processes. Selected conjugates of the invention include a modifying group that is attached to the amine moiety of an amino-saccharide, e.g., mannosamine, glucosamine, galactosamine, sialic acid etc. Exemplary modifying group-intact glycosyl linking group cassettes according to this motif are based on a sialic acid structure, such as those having the formulae:

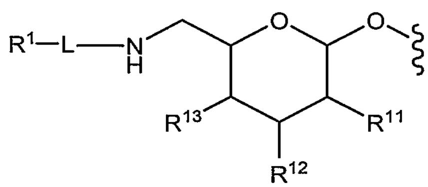


; and



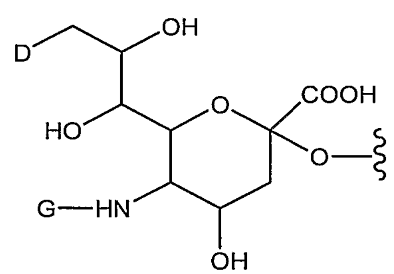
[0159] In the formulae above, R^1 and L are as described above. Further detail about the structure of exemplary R^1 groups is provided below.

[0160] In still a further exemplary embodiment, the conjugate is formed between a peptide and a modified sugar in which the modifying group is attached through a linker at the 6-carbon position of the modified sugar. Thus, illustrative glycosyl linking groups according to this embodiment have the formula:



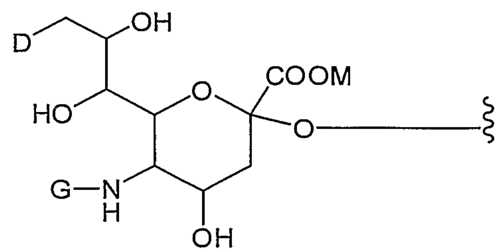
in which the radicals are as discussed above. Glycosyl linking groups include, without limitation, glucose, glucosamine, N-acetyl-glucosamine, galactose, galactosamine, N-acetyl-galactosamine, mannose, mannosamine, N-acetyl-mannosamine, and the like.

[0161] In one embodiment, the present invention provides a peptide conjugate comprising the following glycosyl linking group:



wherein D is a member selected from -OH and R^1-L-HN-; G is a member selected from H and R^1-L- and -C(O)(C_1-C_6)alkyl; R^1 is a moiety comprising a straight-chain or branched poly(ethylene glycol) residue; and L is a linker, e.g., a bond ("zero order"), substituted or unsubstituted alkyl and substituted or unsubstituted heteroalkyl. In exemplary embodiments, when D is OH, G is R^1-L-, and when G is –C(O)(C_1-C_6)alkyl, D is R^1-L-NH-.

[0162] In one embodiment, the present invention provides a peptide conjugate comprising the following glycosyl linking group:

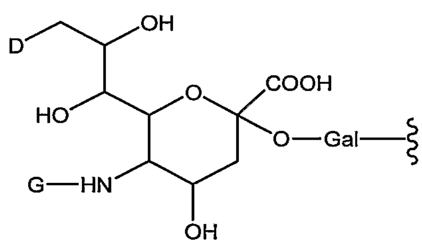


;

D is a member selected from -OH and R^1-L-HN-;G is a member selected from R^1-L- and -C(O)(C_1-C_6)alkyl-R^1; R^1 is a moiety comprising a member selected from a straight-chain poly(ethylene glycol) residue and branched poly(ethylene glycol) residue; and
M is a member selected from H, a salt counterion and a single negative charge; L is a linker which is a member selected from a bond, substituted or unsubstituted alkyl and substituted or unsubstituted heteroalkyl. In an exemplary embodiment, when D is OH, G is R^1-L-. In another exemplary embodiment, when G is –C(O)(C_1-C_6)alkyl, D is R^1-L-NH-.

[0163] In any the compounds of the invention, a COOH group can alternatively be COOM, wherein M is a member selected from H, a negative charge, and a salt counterion.

[0164] The invention provides a peptide conjugate that includes a glycosyl linking group having the formula:



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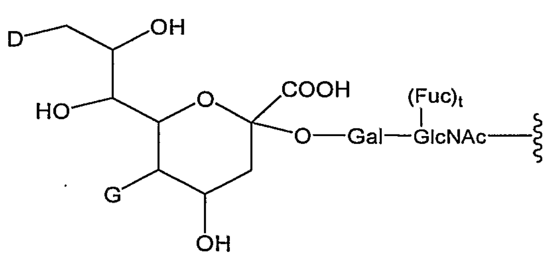
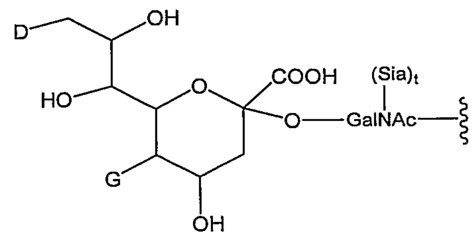
[0165] In other embodiments, the glycosyl linking group has the formula:



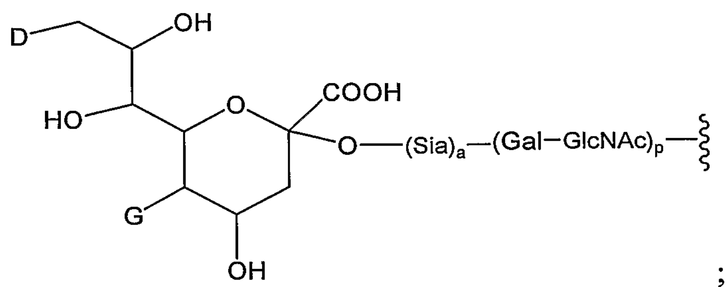
in which the index t is 0 or 1.

[0166] In a still further exemplary embodiment, the glycosyl linking group has the formula:



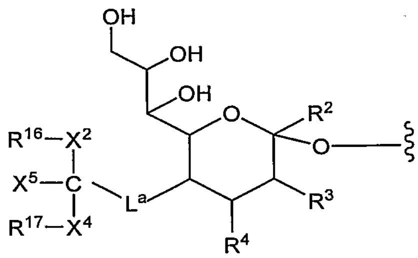
in which the index t is 0 or 1.

[0167] In yet another embodiment, the glycosyl linking group has the formula:



in which the index p represents and integer from 1 to 10; and a is either 0 or 1.

[0168] In another exemplary embodiment, the peptide conjugate comprises a glycosyl moiety selected from the formulae:



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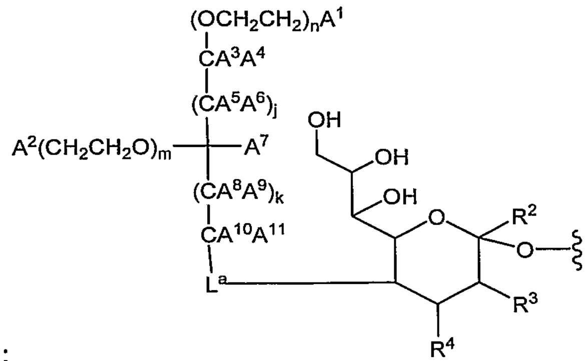

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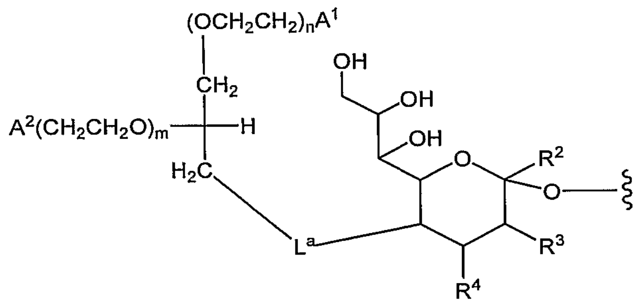

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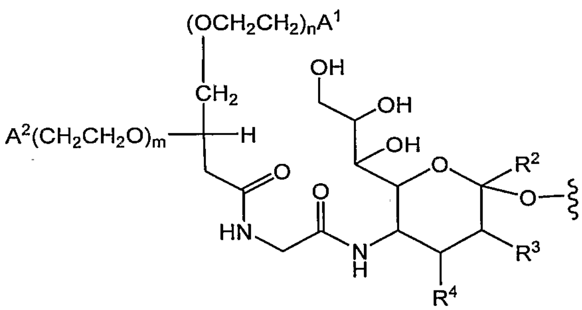

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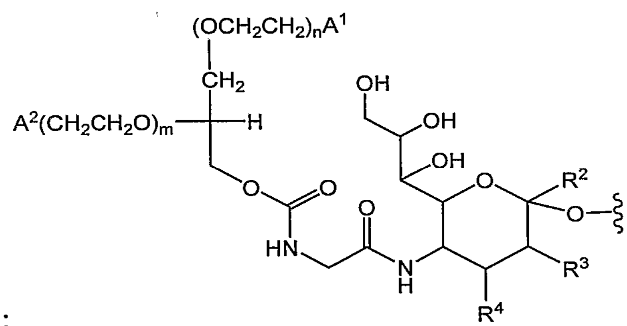

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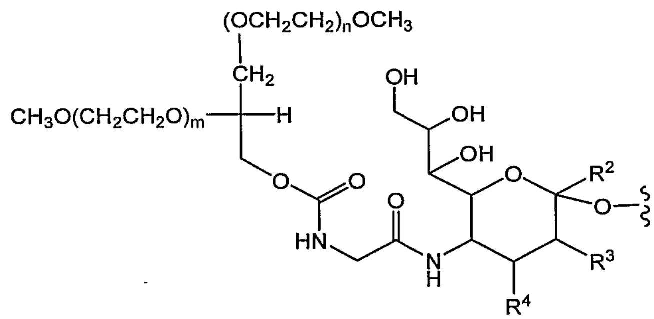

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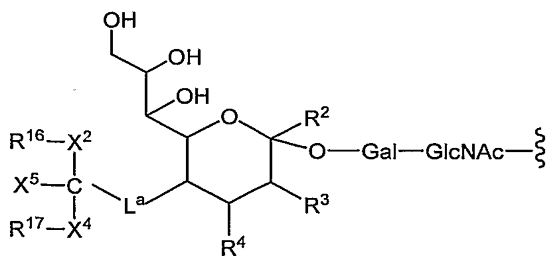

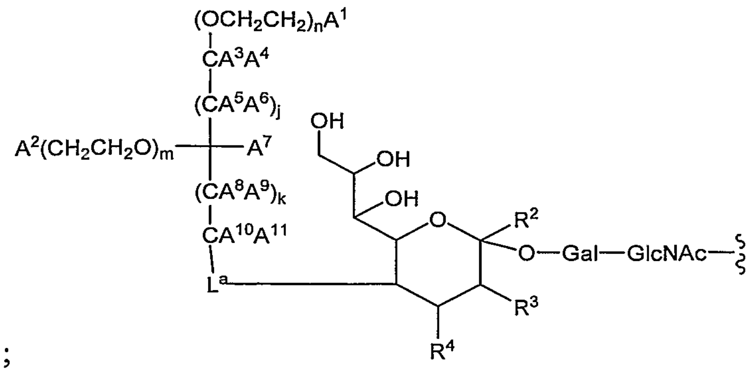

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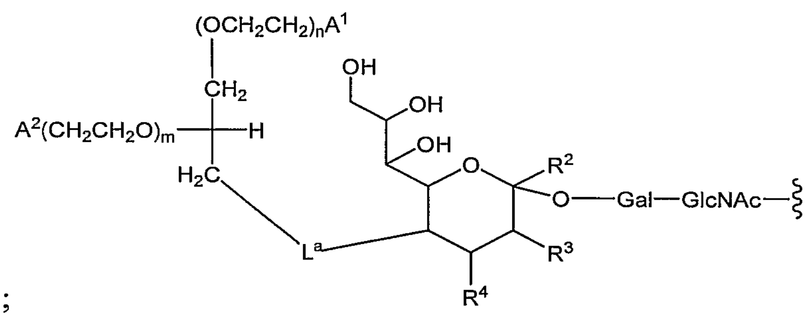

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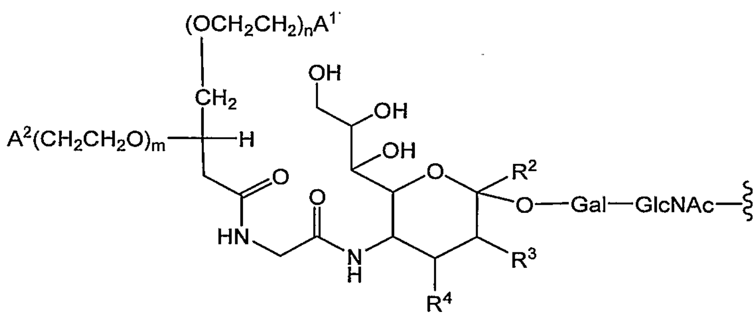

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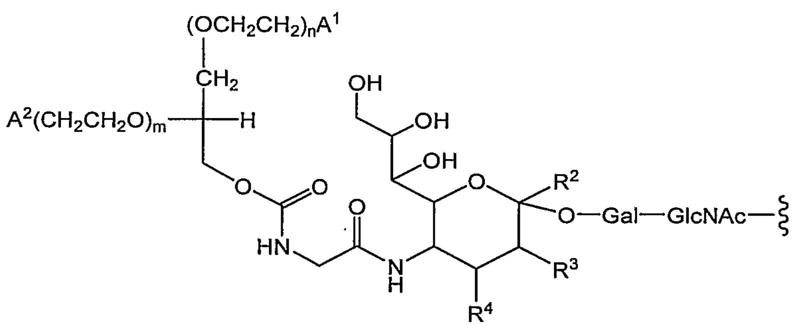

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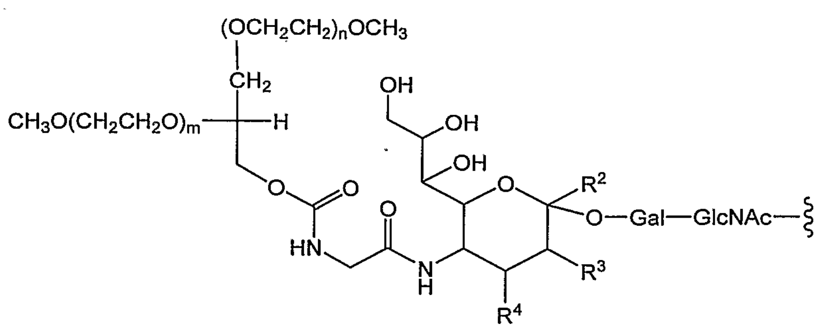

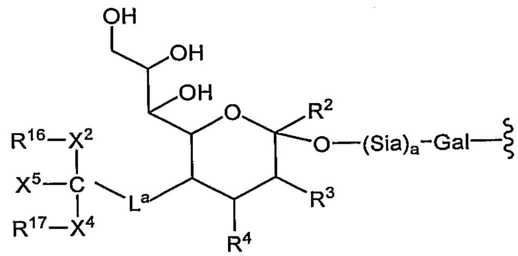
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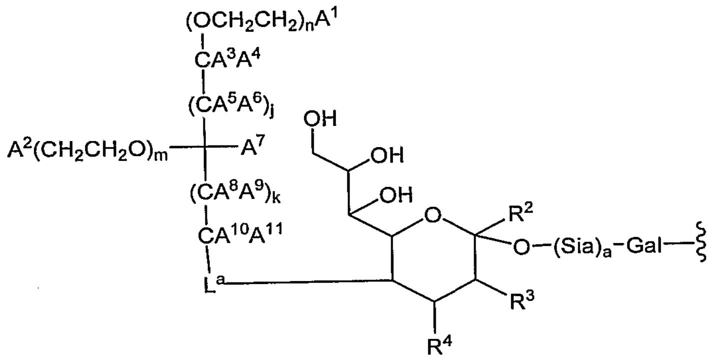
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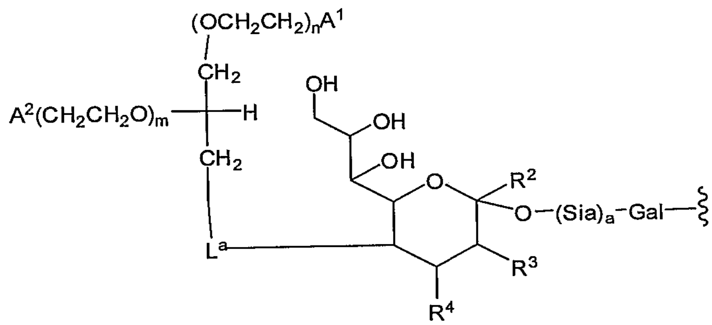
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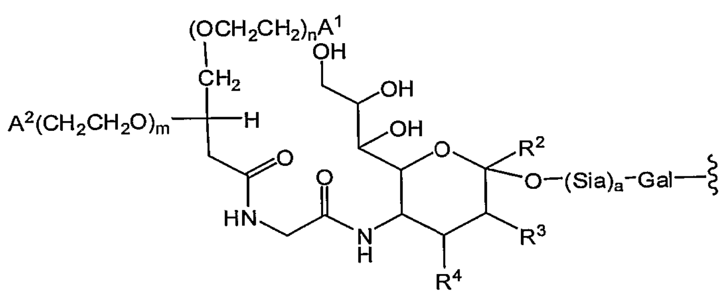
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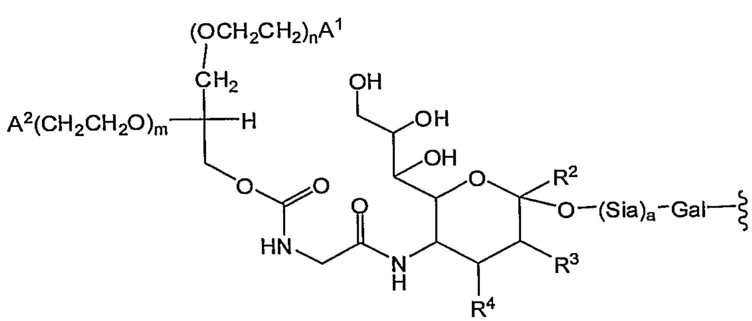
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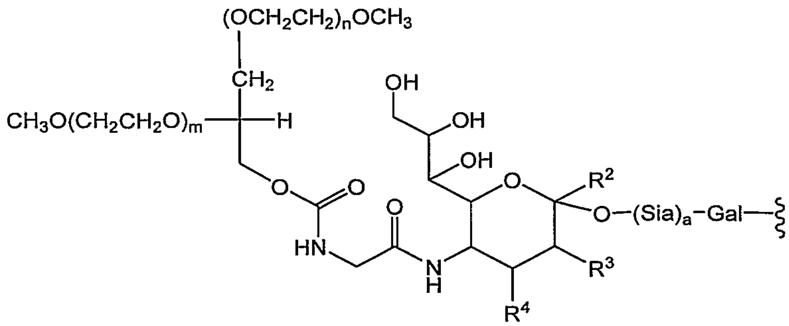

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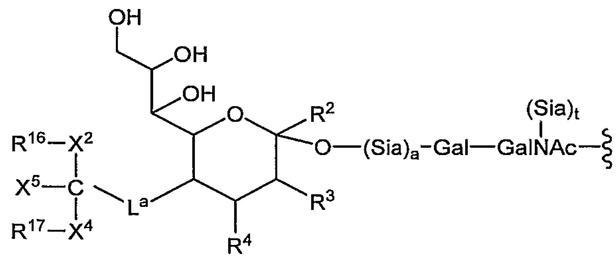

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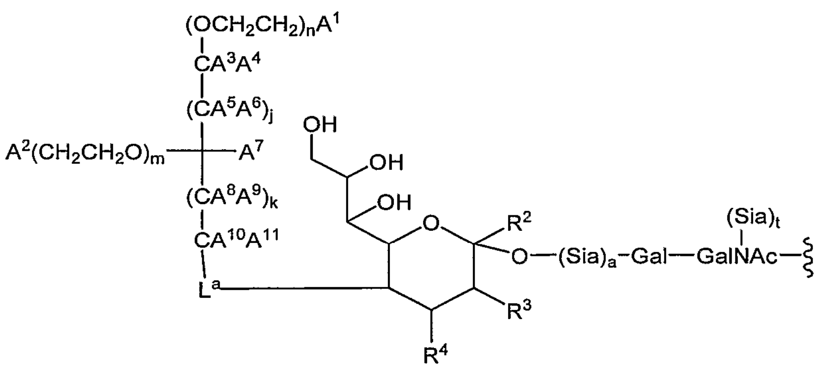

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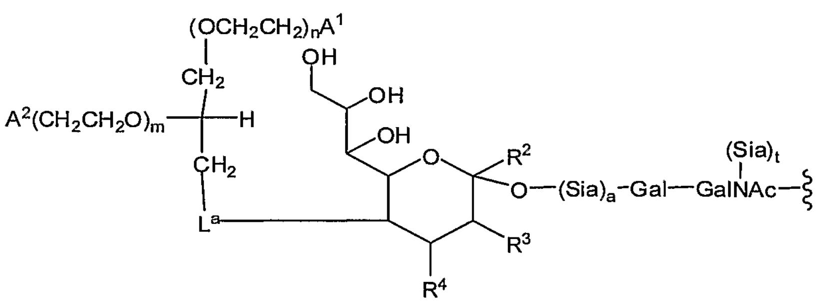

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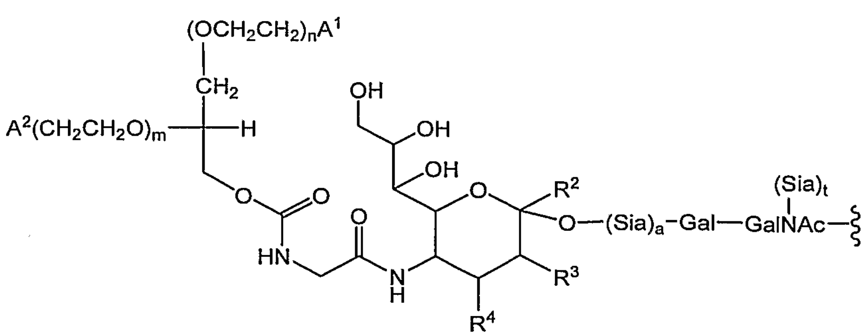

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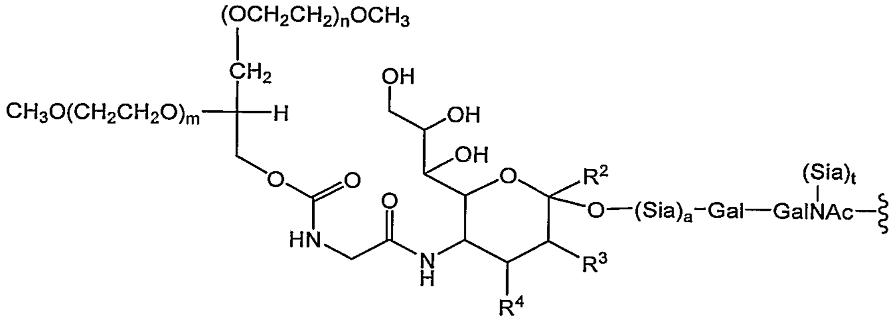

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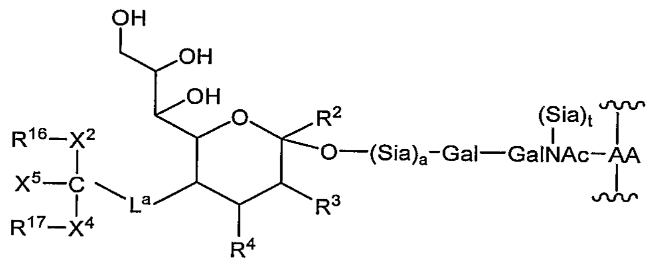

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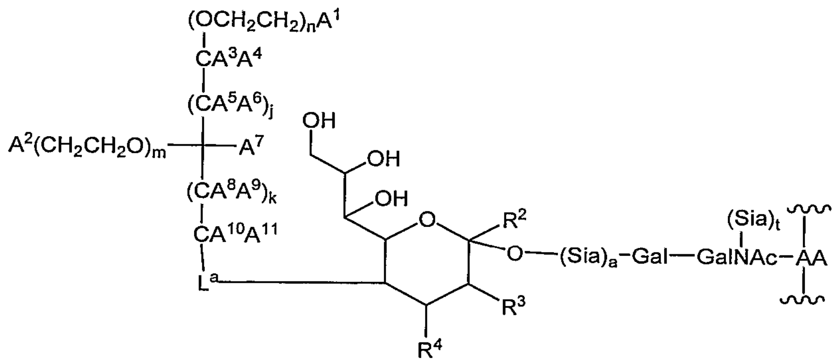

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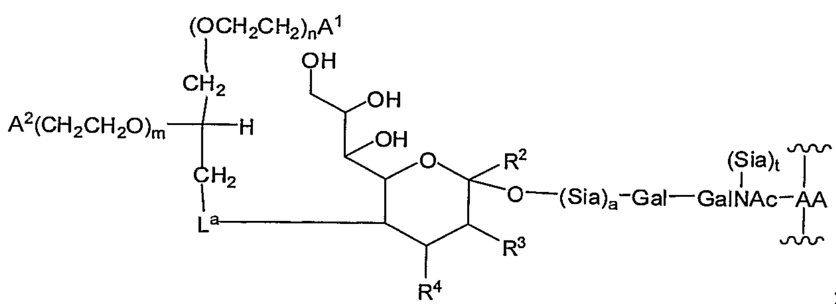

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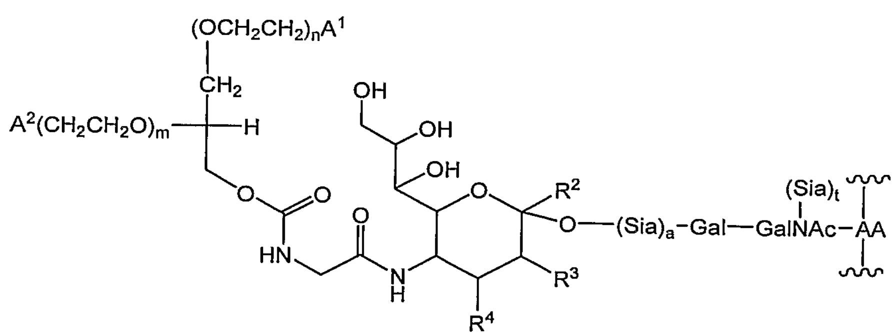

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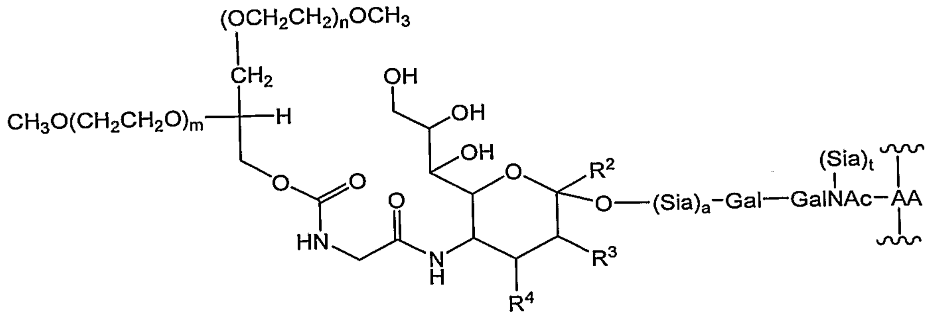

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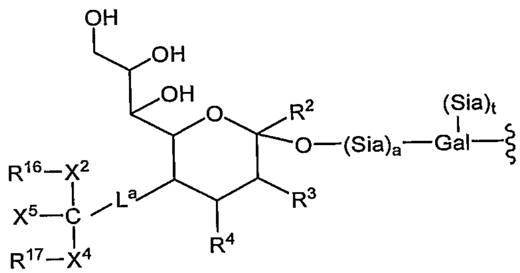

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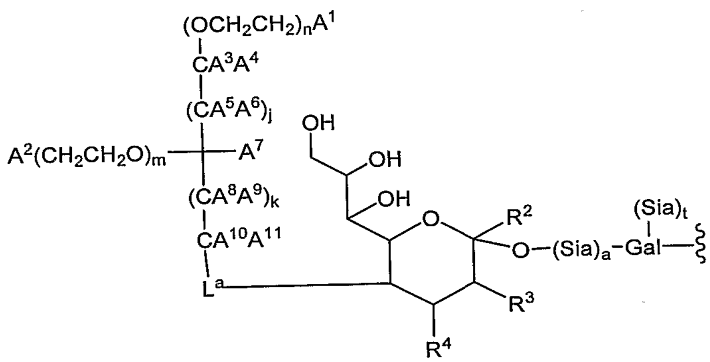

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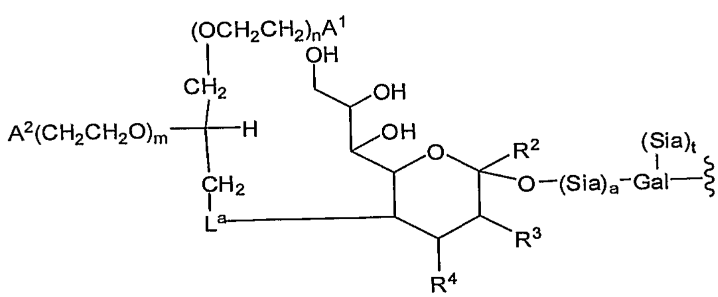

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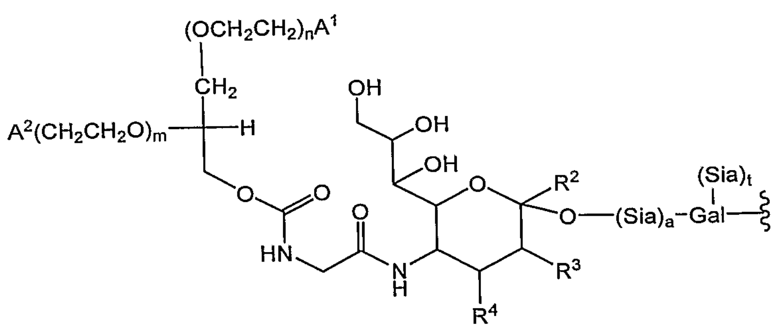

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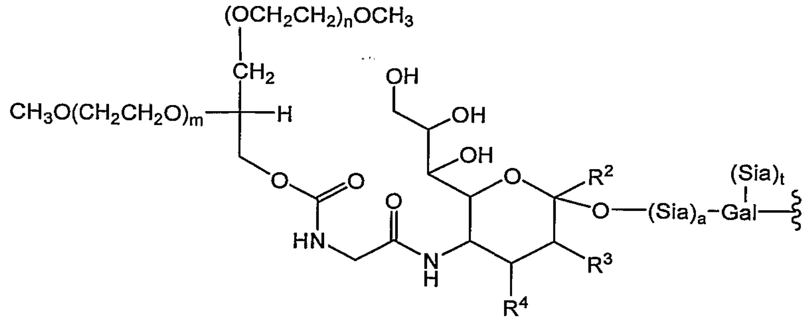

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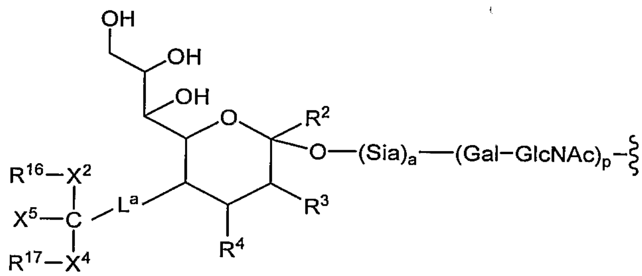

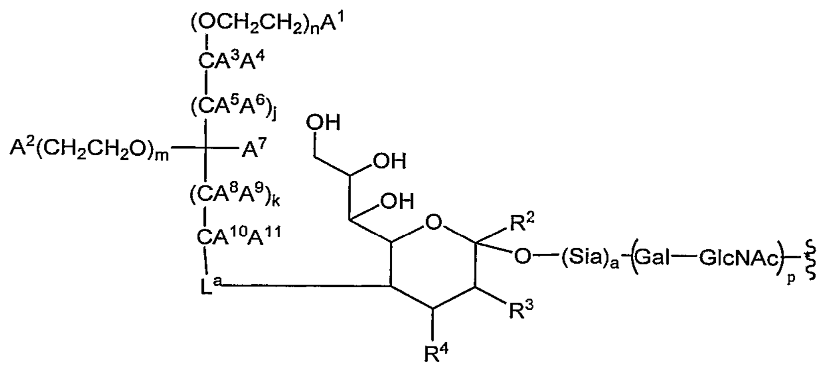

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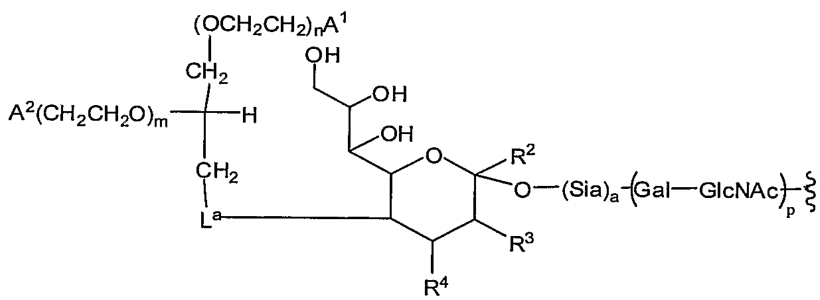

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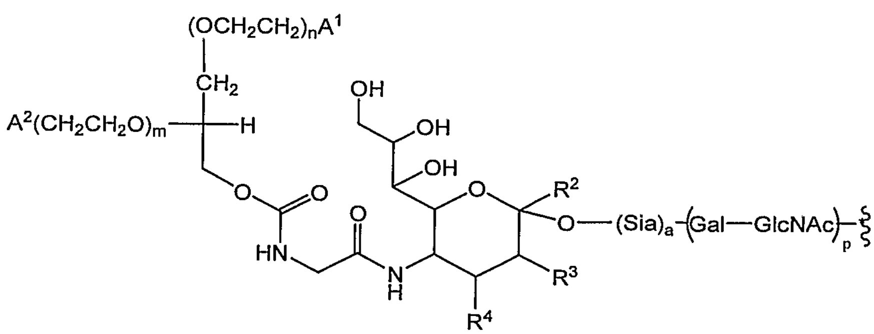

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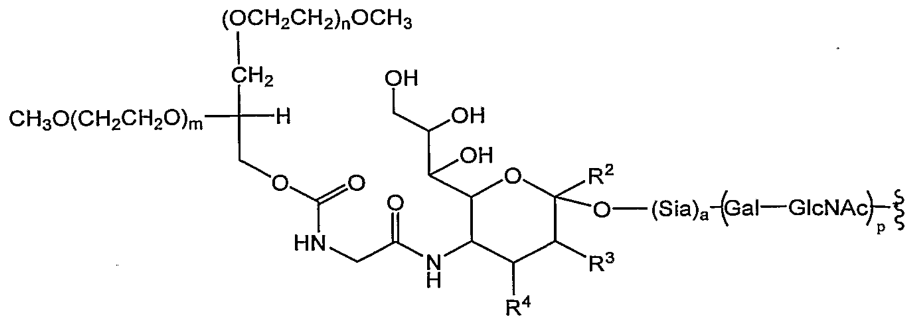

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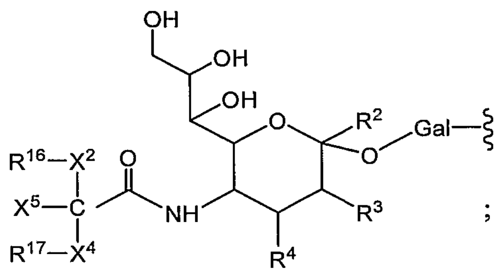

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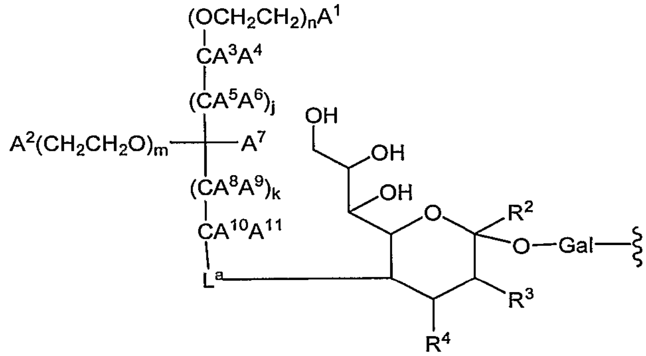

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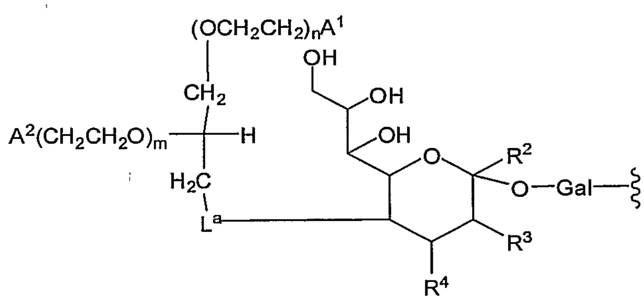

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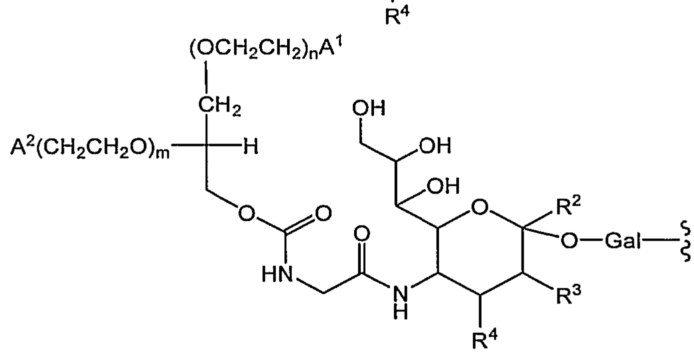

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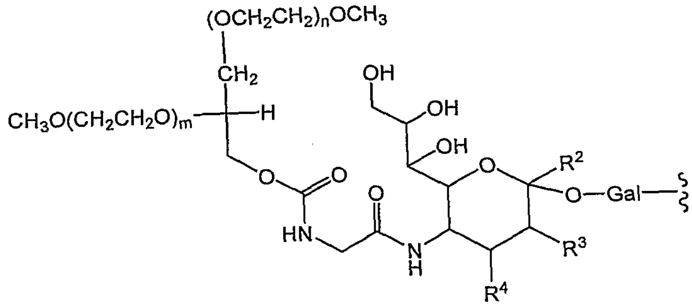

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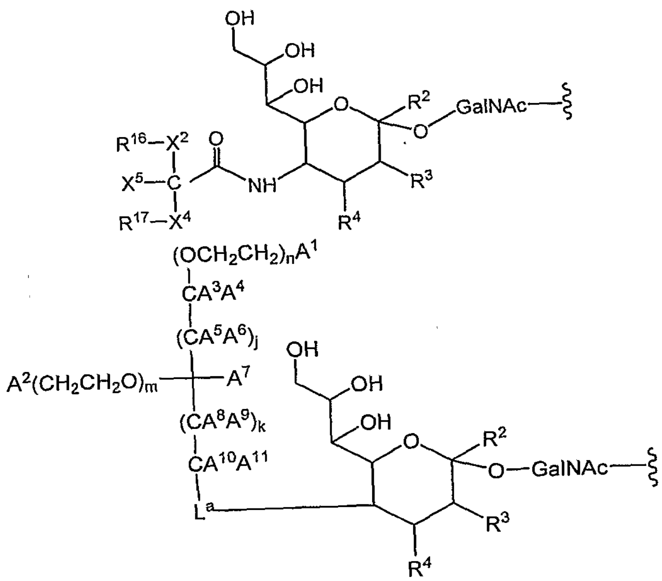

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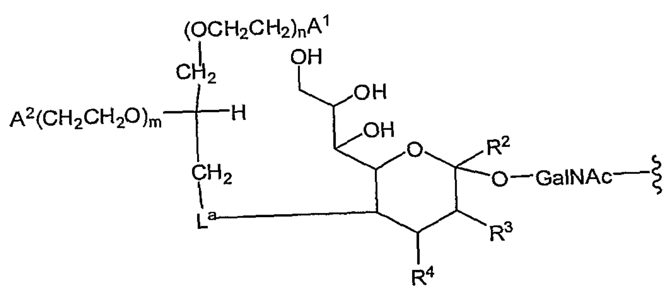

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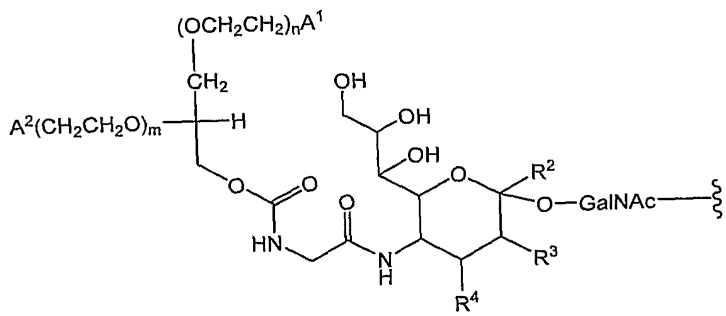

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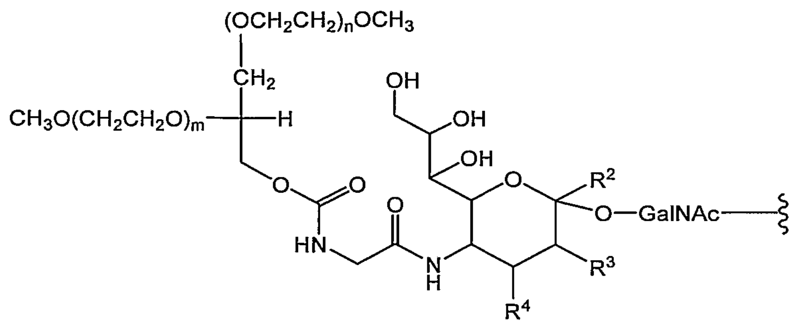

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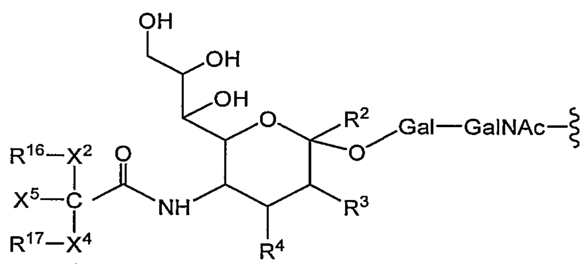

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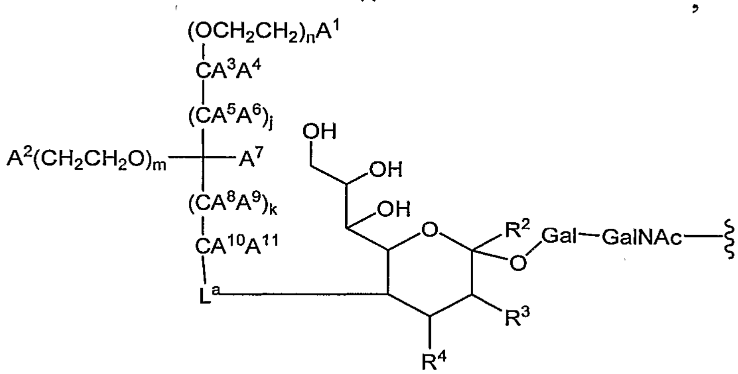

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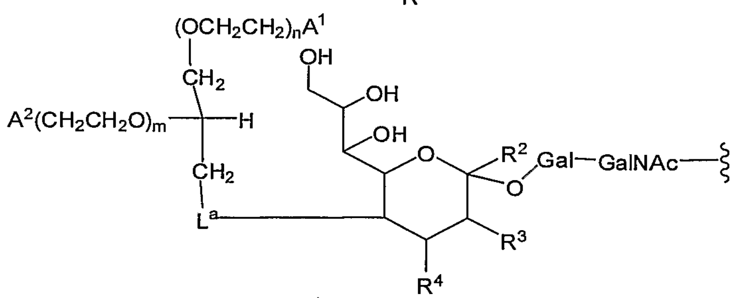

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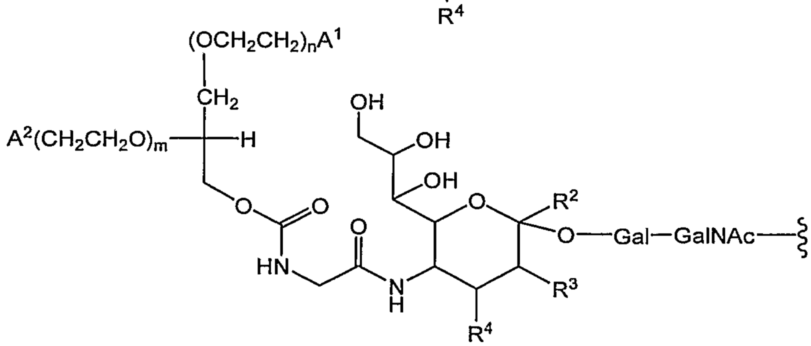

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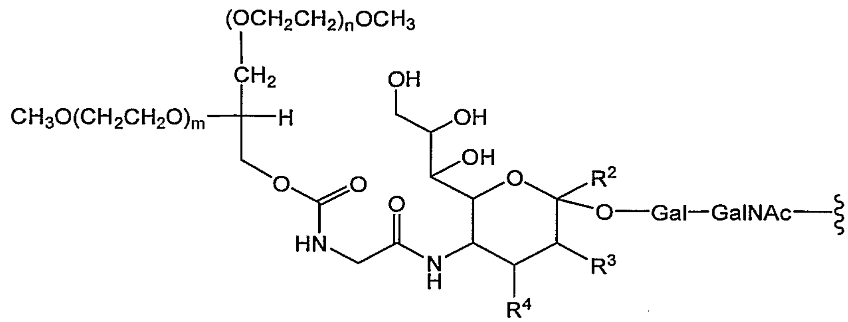

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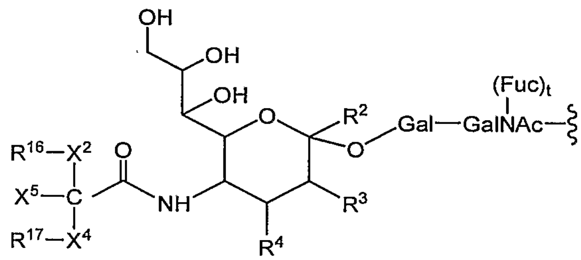

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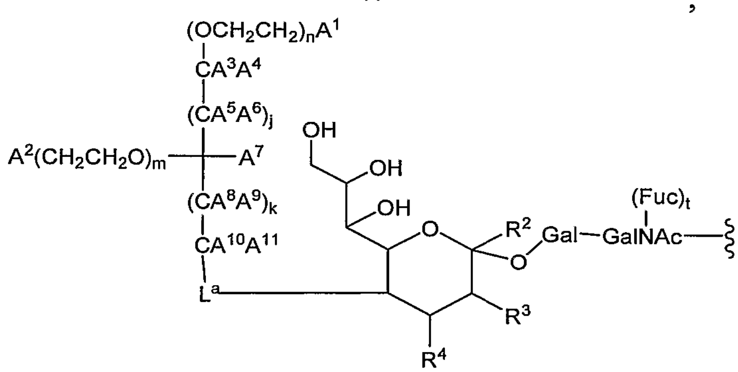

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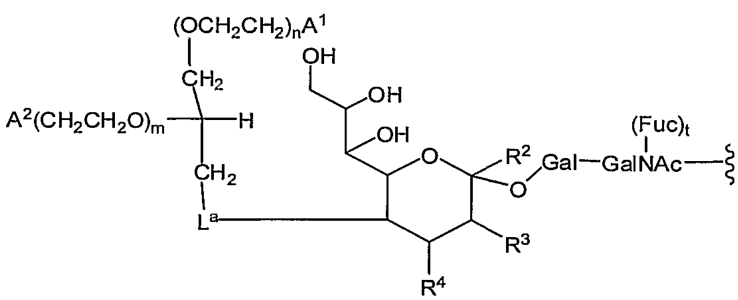

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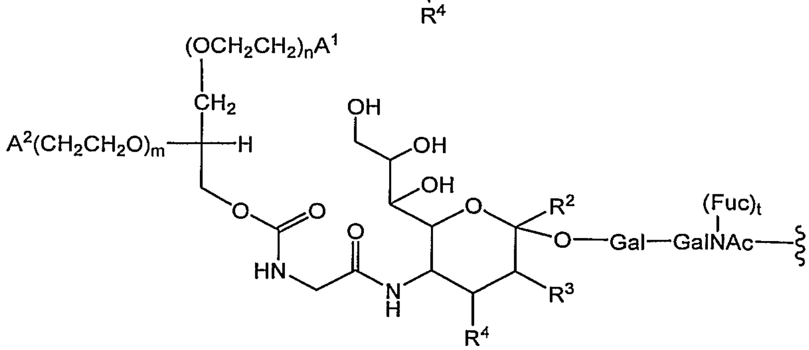

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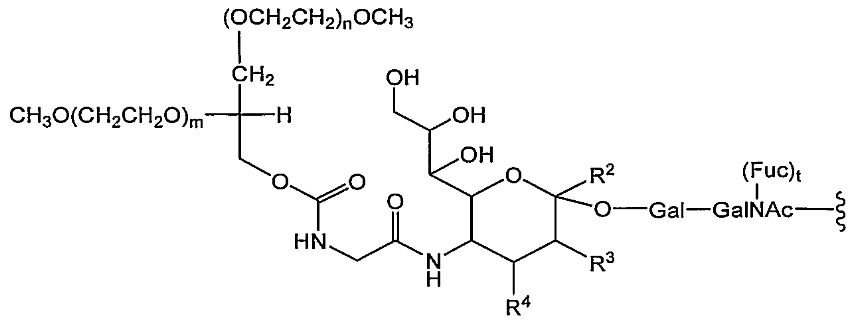

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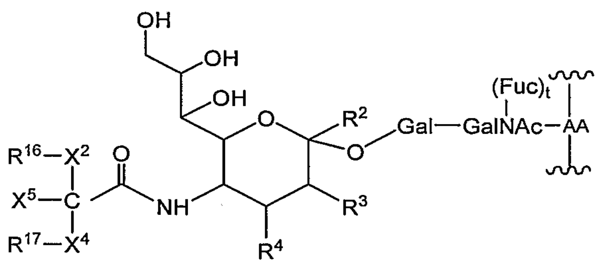

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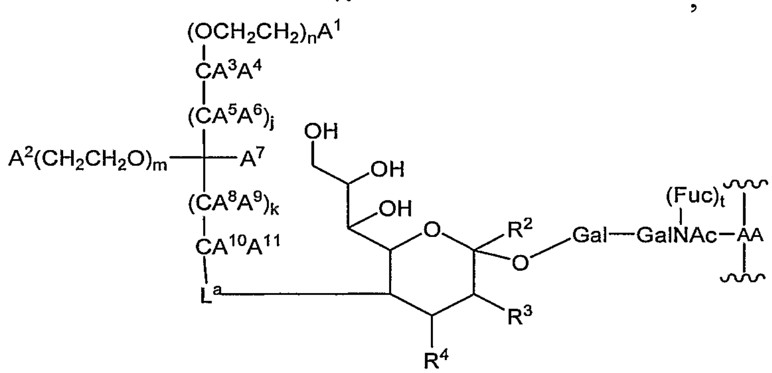

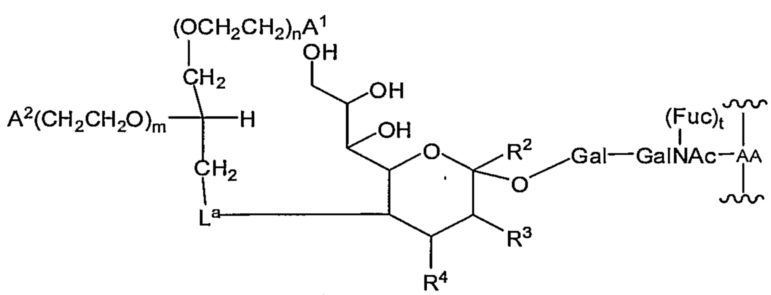

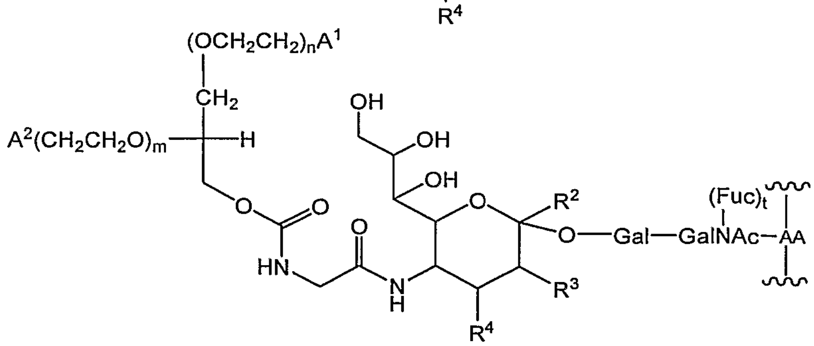

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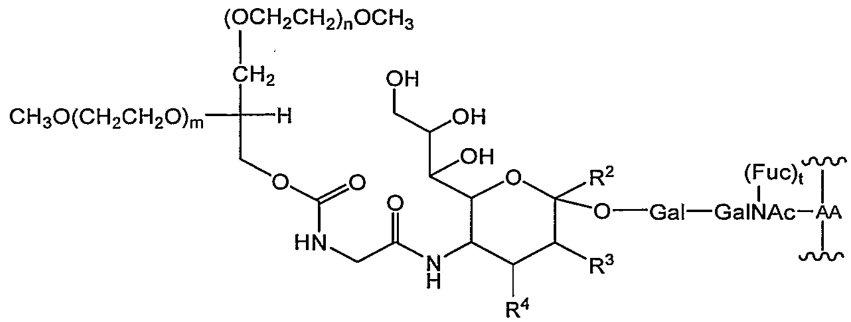

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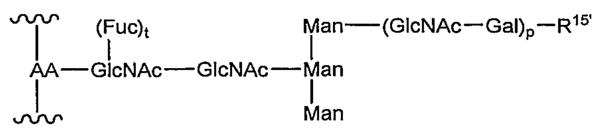
in which the index a and the linker L^a are as discussed above. The index p is an integer from 1 to 10. The indices t and a are independently selected from 0 or 1. Each of these groups can be included as components of the mono-, bi-, tri- and tetra-antennary saccharide structures set forth above. AA is an amino acid residue of the peptide.

[0169] In an exemplary embodiment, the PEG moiety has a molecular weight of about 20 KDa. In another exemplary embodiment, the PEG moiety has a molecular weight of about 5 KDa. In another exemplary embodiment, the PEG moiety has a molecular weight of about 10 KDa. In another exemplary embodiment, the PEG moiety has a molecular weight of about 40 KDa.

[0170] In an exemplary embodiment, the glycosyl linking group is a branched SA-PEG-10 KDa moiety based on a cysteine residue, and one or two of these glycosyl linking groups are covalently attached to the peptide. In another exemplary embodiment, the glycosyl linking group is a branched SA-PEG-10 KDa moiety based on a lysine residue, and one or two of these glycosyl linking groups are covalently attached to the peptide. In an exemplary embodiment, the glycosyl linking group is a branched SA-PEG-10 KDa moiety based on a cysteine residue, and one or two of these glycosyl linking groups are covalently attached to the peptide. In an exemplary embodiment, the glycosyl linking group is a branched SA-PEG-10 KDa moiety based on a lysine residue, and one or two of these glycosyl linking groups are covalently attached to the peptide. In an exemplary embodiment, the glycosyl linking group is a branched SA-PEG-5KDa moiety based on a cysteine residue, and one, two or three of these glycosyl linking groups are covalently attached to the peptide. In an exemplary embodiment, the glycosyl linking group is a branched SA-PEG-5KDa moiety based on a lysine residue, and one, two or three of these glycosyl linking groups are covalently attached to the peptide. In an exemplary embodiment, the glycosyl linking group is a branched SA-PEG-40KDa moiety based on a cysteine residue, and one or two of these glycosyl linking groups are covalently attached to the peptide. In an exemplary embodiment, the glycosyl

linking group is a branched SA-PEG-40KDa moiety based on a lysine residue, and one or two of these glycosyl linking groups are covalently attached to the peptide.

[0171] In an exemplary embodiment, a glycoPEGylated peptide conjugate of the invention selected from the formulae set forth below:



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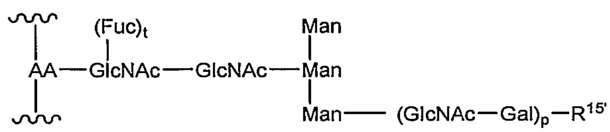

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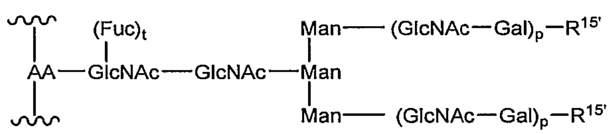

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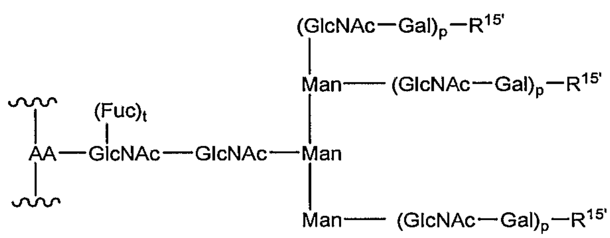

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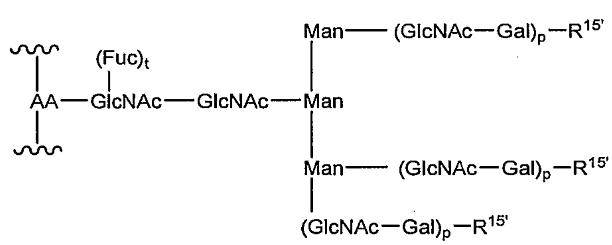

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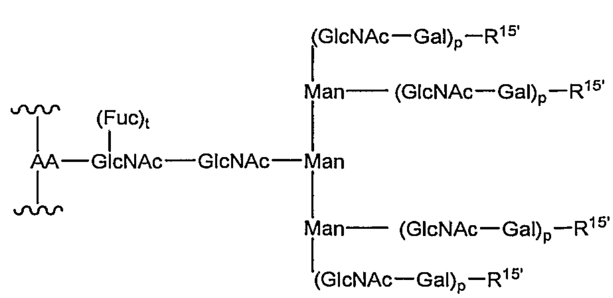
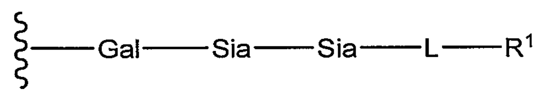


[0172] In the formulae above, the index t is an integer from 0 to 1 and the index p is an integer from 1 to 10. The symbol $R^{15'}$ represents H, OH (e.g., Gal-<u>OH</u>), a sialyl moiety, a sialyl linking group (i.e., sialyl linking group-polymeric modifying group (Sia-L-R^1), or a

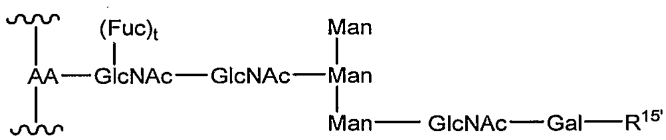
sialyl moiety to which is bound a polymer modified sialyl moiety (e.g., Sia-Sia-L-R^1) ("Sia-Sia^{p}")). Exemplary polymer modified saccharyl moieties have a structure according to Formulae I and II. An exemplary peptide conjugate of the invention will include at least one glycan having a $R^{15'}$ that includes a structure according to Formulae I or II. The oxygen, with the open valence, of Formulae I and II is preferably attached through a glycosidic linkage to a carbon of a Gal or GalNAc moiety. In a further exemplary embodiment, the oxygen is attached to the carbon at position 3 of a galactose residue. In an exemplary embodiment, the modified sialic acid is linked α2,3-to the galactose residue. In another exemplary embodiment, the sialic acid is linked α2,6-to the galactose residue.

[0173] In an exemplary embodiment, the sialyl linking group is a sialyl moiety to which is bound a polymer modified sialyl moiety (e.g., Sia-Sia-L-R^1) ("Sia-Sia^{p}"). Here, the glycosyl linking group is linked to a galactosyl moiety through a sialyl moiety:



An exemplary species according to this motif is prepared by conjugating Sia-L-R^1 to a terminal sialic acid of a glycan using an enzyme that forms Sia-Sia bonds, e.g., CST-II, ST8Sia-II, ST8Sia-III and ST8Sia-IV.

[0174] In another exemplary embodiment, the glycans on the peptide conjugates have a formula that is selected from the group:



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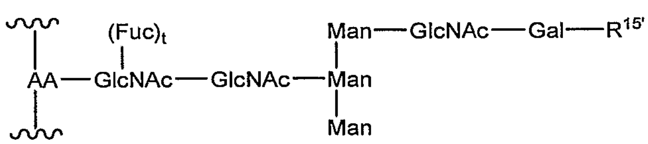

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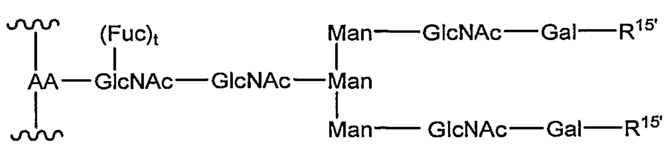


;

and combinations thereof.

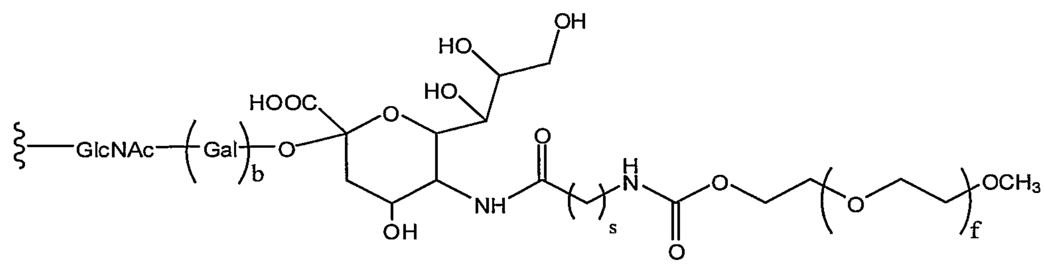
[0175] In each of the formulae above, $R^{15'}$ is as discussed above. Moreover, an exemplary peptide conjugate of the invention will include at least one glycan with an R^{15} moiety having a structure according to Formulae I or II.

[0176] In another exemplary embodiment, the glycosyl linking group comprises at least one glycosyl linking group having the formula:

$$\text{—(GlcNAc—Gal)}_p\text{—}R^{15} \quad ; \text{ and } \quad \text{—(GlcNAc—Gal)}_p\text{—Sia—}R^{15}$$

wherein R^{15} is said sialyl linking group; and the index p is an integer selected from 1 to 10.

[0177] In an exemplary embodiment, the glycosyl linking moiety has the formula:

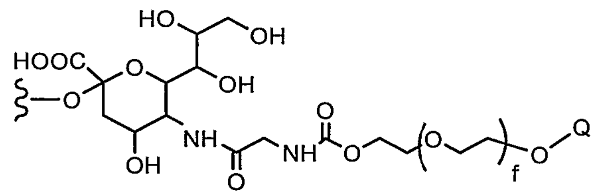


in which b is an integer from 0 to 1. The index s represents an integer from 1 to 10; and the index f represents an integer from 1 to 2500.

[0178] In an exemplary embodiment, the polymeric modifying group is PEG. In another exemplary embodiment, the PEG moiety has a molecular weight of about 20 KDa. In another exemplary embodiment, the PEG moiety has a molecular weight of about 5 KDa. In another exemplary embodiment, the PEG moiety has a molecular weight of about 10 KDa. In another exemplary embodiment, the PEG moiety has a molecular weight of about 40 kDa. In another exemplary embodiment the glycosyl linking group is attached to Asn145, Asn322, Ser52, Ser60 or combinations thereof.

[0179] In an exemplary embodiment, the glycosyl linking group is a linear SA-PEG-10 KDa moiety, and one or two of these glycosyl linking groups are covalently attached to the peptide. In another exemplary embodiment, the glycosyl linking group is a linear SA-PEG-20 KDa moiety, and one or two of these glycosyl linking groups are covalently attached to the peptide. In an exemplary embodiment, the glycosyl linking group is a linear SA-PEG-5 KDa moiety, and one, two or three of these glycosyl linking groups are covalently attached to the peptide. In an exemplary embodiment, the glycosyl linking group is a linear SA-PEG-40 KDa moiety, and one or two of these glycosyl linking groups are covalently attached to the peptide.

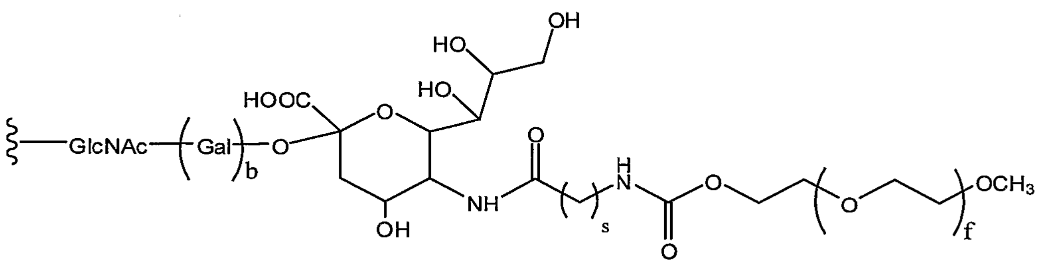
[0180] In another exemplary embodiment, the glycosyl linking group is a sialyl linking group having the formula:



In another exemplary embodiment, Q is a member selected from H and CH_3. In another exemplary embodiment, wherein said glycosyl linking group has the formula:

$$\wr\!-\!-(\text{GlcNAc}-\text{Gal})_p-R^{15} \quad ; \text{ and } \quad \wr\!-\!-(\text{GlcNAc}-\text{Gal})_p-\text{Sia}-R^{15}$$

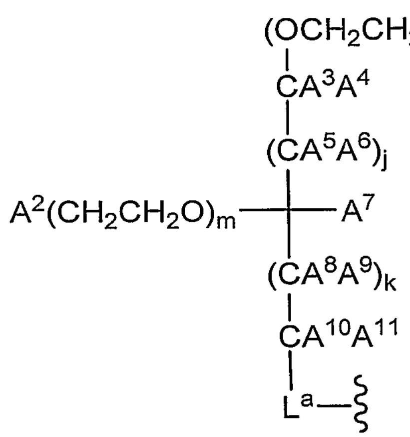
wherein R^{15} is said sialyl linking group; and the index p is an integer selected from 1 to 10. In an exemplary embodiment, the glycosyl linking group comprises the formula:



wherein the index b is an integer selected from 0 and 1. In an exemplary embodiment, the index s is 1; and the index f is an integer selceted from about 200 to about 300. In another exemplary embodiment, the glycosyl linking group is a member selected from SA-PEG-10 KDa and SA-PEG-20 KDa, and wherein the number of said glycosyl linking groups which are covalently attached to the Factor VII/Factor VIIa peptide is an integer selected from 1 to 2. In another exemplary embodiment, the glycosyl linking group is member selected from SA-PEG-5 KDa and SA-PEG-40 KDa, and wherein the number of said glycosyl linking groups which are covalently attached to the Factor VII/Factor VIIa peptide is an integer selected from 1 to 3.

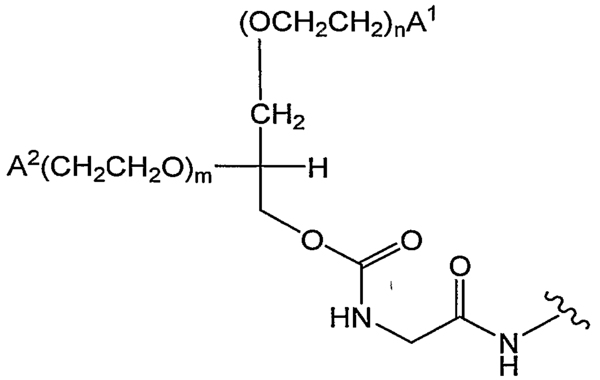
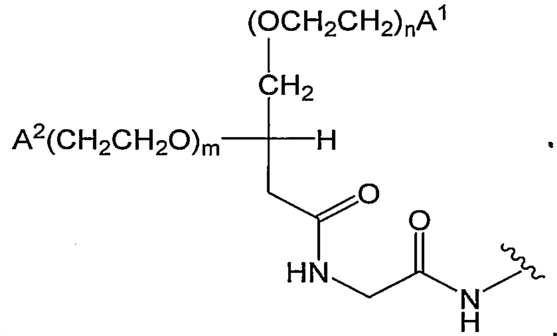
II. D. *Modifying Groups*

[0181] The peptide conjugates of the invention comprise a modifying group. This group can be covalently attached to a Factor VII/Factor VIIa peptide through an amino acid or a glycosyl linking group. In another exemplary embodiment, when the modifying group is

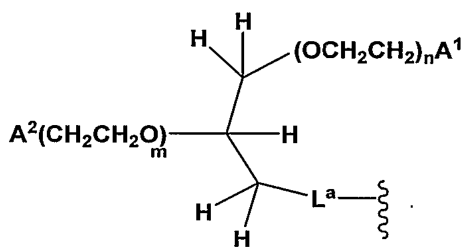


the peptide in the peptide conjugate is a member selected from the peptides in **FIG. 13**. In another exemplary embodiment, the peptide in the peptide conjugate is a member selected from Factor VII, Factor VIIa, Factor VIII, Factor IX, Factor X, Factor XI, erythropoietin, granulocyte colony stimulating factor (G-CSF), Granulocyte-Macrophage Colony Stimulating Factor (GM-CSF)interferon alpha, interferon beta, interferon gamma, α_1-antitrypsin (ATT, or α-1 protease inhibitor, glucocerebrosidase, Tissue-Type Plasminogen Activator (TPA), Interleukin-2 (IL-2), urokinase, human DNase, insulin, Hepatitis B surface protein (HbsAg), human growth hormone, TNF Receptor-IgG Fc region fusion protein (Enbrel™), anti-HER2 monoclonal antibody (Herceptin™), monoclonal antibody to Protein F of Respiratory Syncytial Virus (Synagis™), monoclonal antibody to TNF-α (Remicade™), monoclonal antibody to glycoprotein IIb/IIIa (Reopro™), monoclonal antibody to CD20 (Rituxan™), anti-thrombin III (AT III), human Chorionic Gonadotropin (hCG), alpha-galactosidase (Fabrazyme™), alpha-iduronidase (Aldurazyme™), follicle stimulating hormone, beta-glucosidase, anti-TNF-alpha monoclonal antibody (MLB 5075), glucagon-like peptide-1 (GLP-1), beta-glucosidase (MLB 5064), alpha-galactosidase A (MLB 5082) and fibroblast growth factor. "Modifying groups" can encompass a variety of structures including targeting moieties, therapeutic moieties, biomolecules. Additionally, "modifying groups" include polymeric modifying groups, which are polymers which can alter a property of the peptide such as its bioavailability or its half-life in the body.

[0182] In an exemplary embodiment, the polymeric modifying group has a structure according to the following formulae:

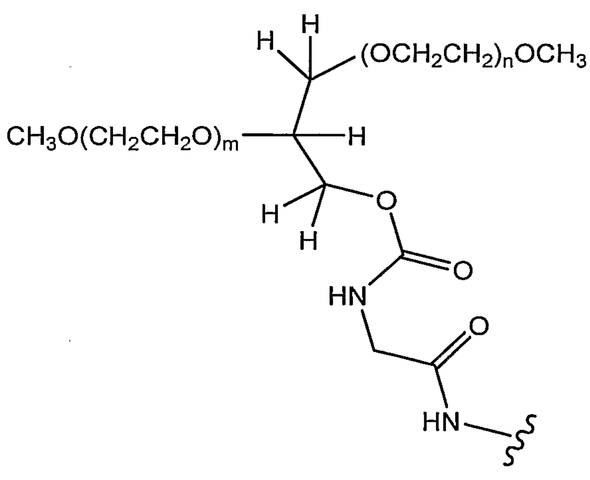
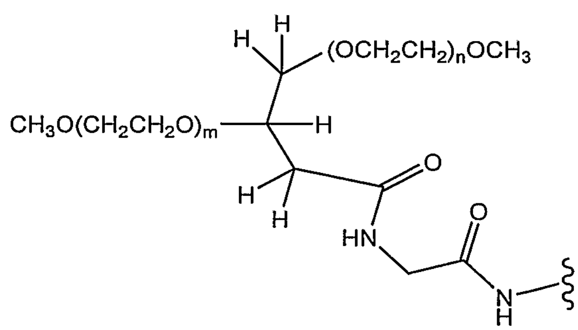
 and 

[0183] In another exemplary embodiment according to the formula above, the polymeric modifying group has a structure according to the following formula:



In an exemplary embodiment, A^1 and A^2 are each members selected from -OH and $-OCH_3$.

[0184] Exemplary polymeric modifying groups according to this embodiment include:

 and 

[0185] For the purposes of convenience, the modifying groups in the remainder of this section will be largely based on polymeric modifying groups such as water soluble and water insoluble polymers. However, one of skill in the art will recognize that other modifying

groups, such as targeting moieties, therapeutic moieties and biomolecules, could be used in place of the polymeric modifying groups.

II. D. i. ***Linkers of the Modifying Groups***

[0186] The linkers of the modifying group serve to attach the modifying group (ie polymeric modifying groups, targeting moieties, therapeutic moieties and biomolecules) to the peptide. In an exemplary embodiment, the polymeric modifying group is bound to a glycosyl linking group, generally through a heteroatom, e.g, nitrogen, on the core through a linker, L, as shown below:

$$(R^1)_w\text{—}L\text{—}\wr \, .$$

R^1 is the polymeric moiety and L is selected from a bond and a linking group. The index w represents an integer selected from 1-6, preferably 1-3 and more preferably 1-2. Exemplary linking groups include substituted or unsubstituted alkyl, substituted or unsubstituted heteroalkyl moieties and sialic acid. An exemplary component of the linker is an acyl moiety.

[0187] An exemplary compound according to the invention has a structure according to Formulae I or II above, in which at least one of R^2, R^3, R^4, R^5, R^6 or $R^{6'}$ has the formula:

$$\wr\text{—}NH\text{—}L\text{—}R^1 \, .$$

[0188] In another example according to this embodiment at least one of R^2, R^3, R^4, R^5, R^6 or $R^{6'}$ has the formula:

$$\wr\text{—}NHC(O)(CH_2)_s\text{—}NHC(O)\text{—}R^1$$

in which s is an integer from 0 to 20 and R^1 is a linear polymeric modifying moiety.

[0189] In an exemplary embodiment, the polymeric modifying group -linker construct is a branched structure that includes two or more polymeric chains attached to central moiety. In this embodiment, the construct has the formula:

$$(R^1)_{w'}\text{—}L\text{—}\wr$$

in which R^1 and L are as discussed above and w' is an integer from 2 to 6, preferably from 2 to 4 and more preferably from 2 to 3.

[0190] When L is a bond it is formed between a reactive functional group on a precursor of R^1 and a reactive functional group of complementary reactivity on the saccharyl core. When

L is a non-zero order linker, a precursor of L can be in place on the glycosyl moiety prior to reaction with the R^1 precursor. Alternatively, the precursors of R^1 and L can be incorporated into a preformed cassette that is subsequently attached to the glycosyl moiety. As set forth herein, the selection and preparation of precursors with appropriate reactive functional groups is within the ability of those skilled in the art. Moreover, coupling the precursors proceeds by chemistry that is well understood in the art.

[0191] In an exemplary embodiment, L is a linking group that is formed from an amino acid, or small peptide (e.g., 1-4 amino acid residues) providing a modified sugar in which the polymeric modifying group is attached through a substituted alkyl linker. Exemplary linkers include glycine, lysine, serine and cysteine. The PEG moiety can be attached to the amine moiety of the linker through an amide or urethane bond. The PEG is linked to the sulfur or oxygen atoms of cysteine and serine through thioether or ether bonds, respectively.

[0192] In an exemplary embodiment, R^5 includes the polymeric modifying group. In another exemplary embodiment, R^5 includes both the polymeric modifying group and a linker, L, joining the modifying group to the remainder of the molecule. As discussed above, L can be a linear or branched structure. Similarly, the polymeric modifying group can be branched or linear.

II. D. ii. *__Water-Soluble Polymers__*

[0193] Many water-soluble polymers are known to those of skill in the art and are useful in practicing the present invention. The term water-soluble polymer encompasses species such as saccharides (e.g., dextran, amylose, hyalouronic acid, poly(sialic acid), heparans, heparins, etc.); poly (amino acids), e.g., poly(aspartic acid) and poly(glutamic acid); nucleic acids; synthetic polymers (e.g., poly(acrylic acid), poly(ethers), e.g., poly(ethylene glycol); peptides, proteins, and the like. The present invention may be practiced with any water-soluble polymer with the sole limitation that the polymer must include a point at which the remainder of the conjugate can be attached.

[0194] Methods for activation of polymers can also be found in WO 94/17039, U.S. Pat. No. 5,324,844, WO 94/18247, WO 94/04193, U.S. Pat. No. 5,219,564, U.S. Pat. No. 5,122,614, WO 90/13540, U.S. Pat. No. 5,281,698, and more WO 93/15189, and for conjugation between activated polymers and peptides, *e.g.* Coagulation Factor VIII (WO 94/15625), hemoglobin (WO 94/09027), oxygen carrying molecule (U.S. Pat. No.

4,412,989), ribonuclease and superoxide dismutase (Veronese *at al., App. Biochem. Biotech.* **11**: 141-45 (1985)).

[0195] Exemplary water-soluble polymers are those in which a substantial proportion of the polymer molecules in a sample of the polymer are of approximately the same molecular weight; such polymers are "homodisperse."

[0196] The present invention is further illustrated by reference to a poly(ethylene glycol) conjugate. Several reviews and monographs on the functionalization and conjugation of PEG are available. *See,* for example, Harris, M*acronol. Chem. Phys.* **C25**: 325-373 (1985); Scouten, *Methods in Enzymology* **135**: 30-65 (1987); Wong *et al., Enzyme Microb. Technol.* **14**: 866-874 (1992); Delgado *et al., Critical Reviews in Therapeutic Drug Carrier Systems* **9**: 249-304 (1992); Zalipsky, *Bioconjugate Chem.* **6**: 150-165 (1995); and Bhadra, *et al., Pharmazie*, **57**:5-29 (2002). Routes for preparing reactive PEG molecules and forming conjugates using the reactive molecules are known in the art. For example, U.S. Patent No. 5,672,662 discloses a water soluble and isolatable conjugate of an active ester of a polymer acid selected from linear or branched poly(alkylene oxides), poly(oxyethylated polyols), poly(olefinic alcohols), and poly(acrylomorpholine).

[0197] U.S. Patent No. 6,376,604 sets forth a method for preparing a water-soluble 1-benzotriazolylcarbonate ester of a water-soluble and non-peptidic polymer by reacting a terminal hydroxyl of the polymer with di(1-benzotriazoyl)carbonate in an organic solvent. The active ester is used to form conjugates with a biologically active agent such as a protein or peptide.

[0198] WO 99/45964 describes a conjugate comprising a biologically active agent and an activated water soluble polymer comprising a polymer backbone having at least one terminus linked to the polymer backbone through a stable linkage, wherein at least one terminus comprises a branching moiety having proximal reactive groups linked to the branching moiety, in which the biologically active agent is linked to at least one of the proximal reactive groups. Other branched poly(ethylene glycols) are described in WO 96/21469, U.S. Patent No. 5,932,462 describes a conjugate formed with a branched PEG molecule that includes a branched terminus that includes reactive functional groups. The free reactive groups are available to react with a biologically active species, such as a protein or peptide, forming conjugates between the poly(ethylene glycol) and the biologically active species. U.S. Patent

No. 5,446,090 describes a bifunctional PEG linker and its use in forming conjugates having a peptide at each of the PEG linker termini.

[0199] Conjugates that include degradable PEG linkages are described in WO 99/34833; and WO 99/14259, as well as in U.S. Patent No. 6,348,558. Such degradable linkages are applicable in the present invention.

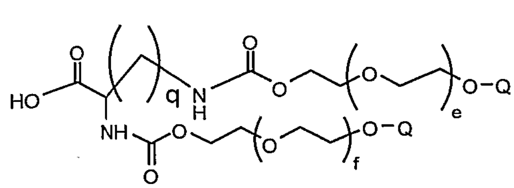
[0200] The art-recognized methods of polymer activation set forth above are of use in the context of the present invention in the formation of the branched polymers set forth herein and also for the conjugation of these branched polymers to other species, e.g., sugars, sugar nucleotides and the like.

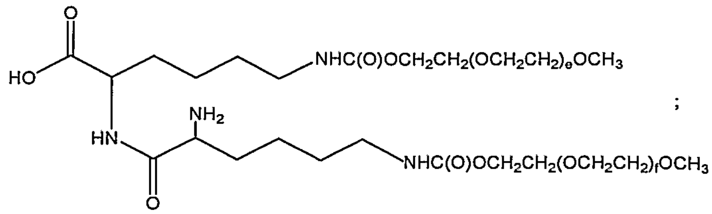
[0201] An exemplary water-soluble polymer is poly(ethylene glycol), e.g., methoxy-poly(ethylene glycol). The poly(ethylene glycol) used in the present invention is not restricted to any particular form or molecular weight range. For unbranched poly(ethylene glycol) molecules the molecular weight is preferably between 500 and 100,000. A molecular weight of 2000-60,000 is preferably used and preferably of from about 5,000 to about 40,000.

II. D. iii. ***Branched Water Soluble Polymers***

[0202] In another embodiment the poly(ethylene glycol) is a branched PEG having more than one PEG moiety attached. Examples of branched PEGs are described in U.S. Pat. No. 5,932,462; U.S. Pat. No. 5,342,940; U.S. Pat. No. 5,643,575; U.S. Pat. No. 5,919,455; U.S. Pat. No. 6,113,906; U.S. Pat. No. 5,183,660; WO 02/09766; Kodera Y., *Bioconjugate Chemistry* **5**: 283-288 (1994); and Yamasaki et al., *Agric. Biol. Chem.*, **52**: 2125-2127, 1998. In a preferred embodiment the molecular weight of each poly(ethylene glycol) of the branched PEG is less than or equal to 40,000 daltons.

[0203] Representative polymeric modifying moieties include structures that are based on side chain-containing amino acids, e.g., serine, cysteine, lysine, and small peptides, e.g., lys-lys. Exemplary structures include:

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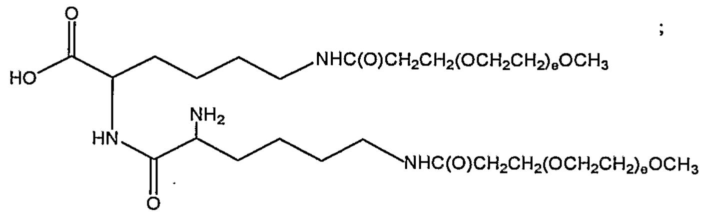
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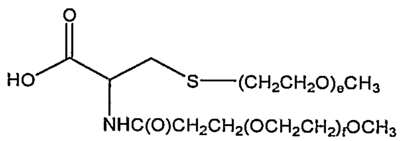
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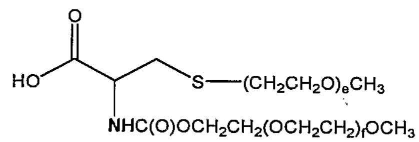

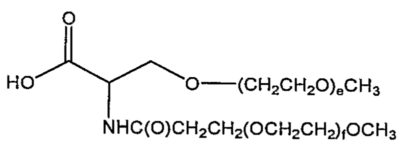
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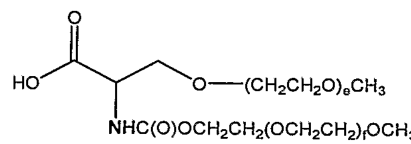

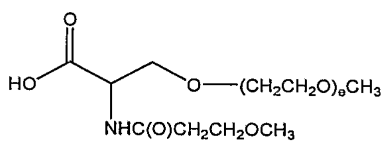
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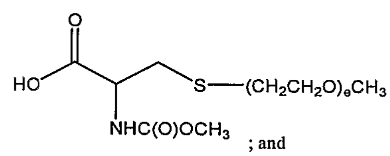
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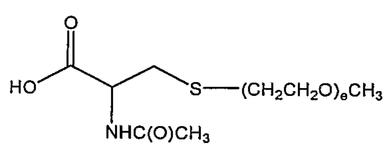
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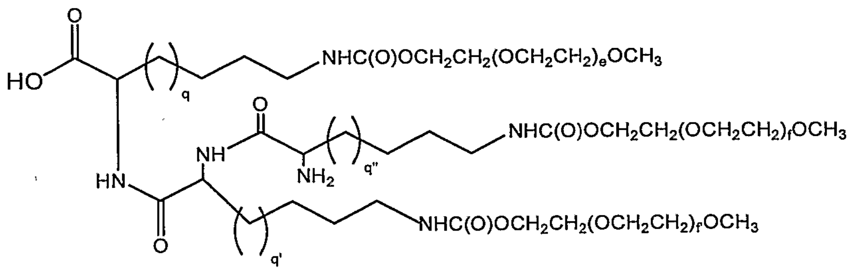
Those of skill will appreciate that the free amine in the di-lysine structures can also be pegylated through an amide or urethane bond with a PEG moiety.

[0204] In yet another embodiment, the polymeric modifying moiety is a branched PEG moiety that is based upon a tri-lysine peptide. The tri-lysine can be mono-, di-, tri-, or tetra-PEG-ylated. Exemplary species according to this embodiment have the formulae:



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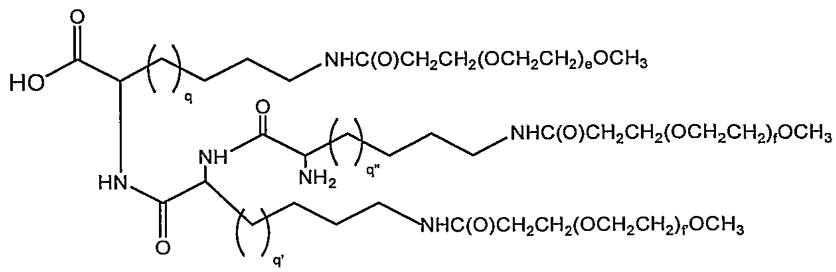


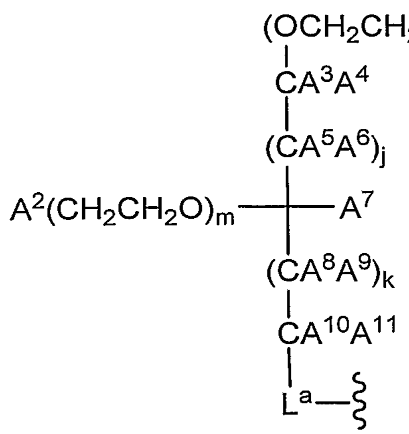
in which the indices e, f and f' are independently selected integers from 1 to 2500; and the indices q, q' and q" are independently selected integers from 1 to 20.

[0205] As will be apparent to those of skill, the branched polymers of use in the invention include variations on the themes set forth above. For example the di-lysine-PEG conjugate shown above can include three polymeric subunits, the third bonded to the α-amine shown as unmodified in the structure above. Similarly, the use of a tri-lysine functionalized with three or four polymeric subunits labeled with the polymeric modifying moiety in a desired manner is within the scope of the invention.

[0206] As discussed herein, the PEG of use in the conjugates of the invention can be linear or branched. An exemplary precursor of use to form the branched PEG containing peptide conjugates according to this embodiment of the invention has the formula:

$$\begin{array}{c} R^{16}-X^{2} \\ | \\ X^{5}-C-X^{3'} \\ | \\ R^{17}-X^{4} \end{array} \qquad \text{(III)}.$$

Another exemplary precursor of use to form the branched PEG containing peptide conjugates according to this embodiment of the invention has the formula:



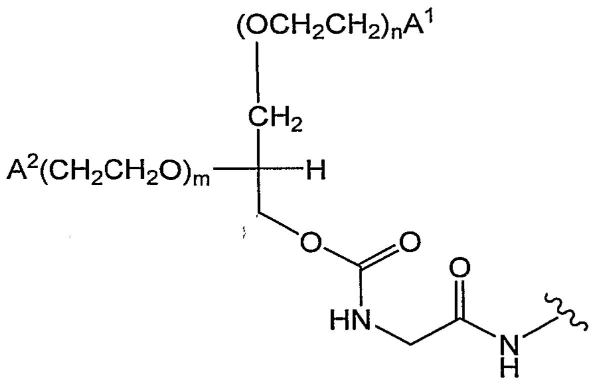
(IIIa).

[0207] The branched polymer species according to this formula are essentially pure water-soluble polymers. $X^{3'}$ is a moiety that includes an ionizable (e.g., OH, COOH, H_2PO_4, HSO_3, HPO_3, and salts thereof, etc.) or other reactive functional group, e.g., *infra*. C is carbon. X^5, R^{16} and R^{17} are independently selected from non-reactive groups (e.g., H, unsubstituted alkyl, unsubstituted heteroalkyl) and polymeric arms (e.g., PEG). X^2 and X^4 are linkage fragments that are preferably essentially non-reactive under physiological conditions, which may be the same or different. An exemplary linker includes neither aromatic nor ester moieties. Alternatively, these linkages can include one or more moiety that is designed to degrade under physiologically relevant conditions, e.g., esters, disulfides, etc. X^2 and X^4 join polymeric arms R^{16} and R^{17} to C. When $X^{3'}$ is reacted with a reactive functional group of complementary reactivity on a linker, sugar or linker-sugar cassette, $X^{3'}$ is converted to a component of linkage fragment X^3.

[0208] Exemplary linkage fragments for X^2, X^3 and X^4 are independently selected and include S, SC(O)NH, HNC(O)S, SC(O)O, O, NH, NHC(O), (O)CNH and NHC(O)O, and OC(O)NH, CH_2S, CH_2O , CH_2CH_2O, CH_2CH_2S, $(CH_2)_oO$, $(CH_2)_oS$ or $(CH_2)_oY'$-PEG wherein, Y' is S, NH, NHC(O), C(O)NH, NHC(O)O, OC(O)NH, or O and o is an integer from 1 to 50. In an exemplary embodiment, the linkage fragments X^2 and X^4 are different linkage fragments.

[0209] In an exemplary embodiment, the precursor (Formula III), or an activated derivative thereof, is reacted with, and thereby bound to a sugar, an activated sugar or a sugar nucleotide through a reaction between $X^{3'}$ and a group of complementary reactivity on the sugar moiety, e.g., an amine. Alternatively, $X^{3'}$ reacts with a reactive functional group on a precursor to linker, L. One or more of R^2, R^3, R^4, R^5, R^6 or $R^{6'}$ of Formulae I and II can include the branched polymeric modifying moiety, or this moiety bound through L.

[0210] In an exemplary embodiment, the polymeric modifying group has a structure according to the following formulae:



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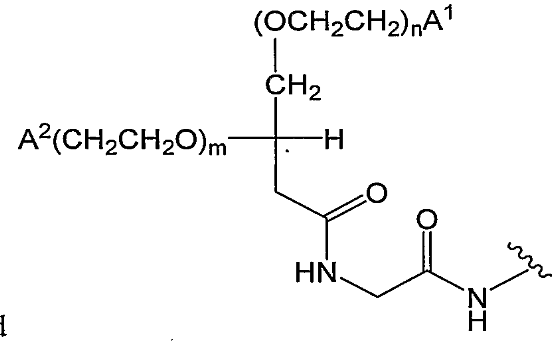

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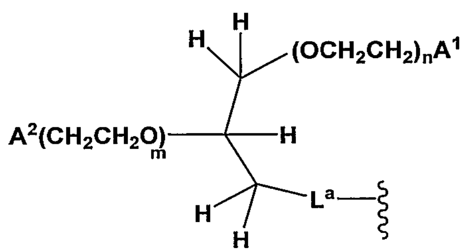
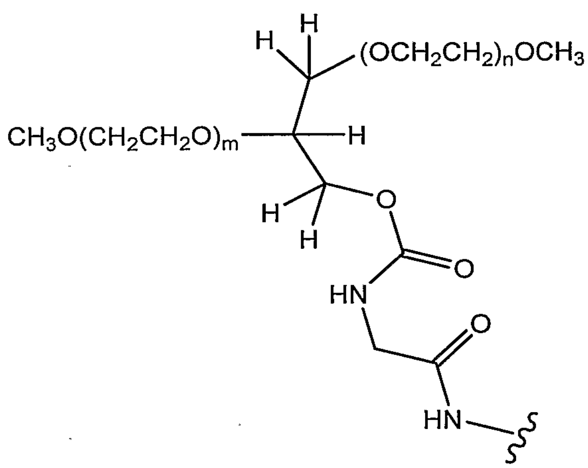
[0211] In another exemplary embodiment according to the formula above, the branched polymer has a structure according to the following formula:



In an exemplary embodiment, A^1 and A^2 are each selected from -OH and $-OCH_3$.

[0212] Exemplary polymeric modifying groups according to this embodiment include:



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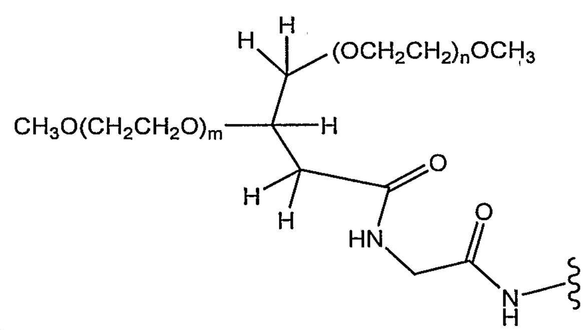
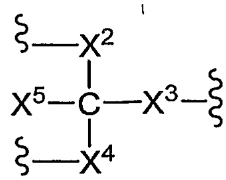
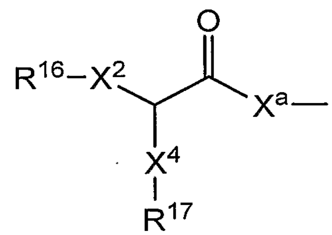


[0213] In an exemplary embodiment, the moiety:



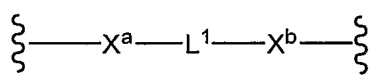
is the linker arm, L. In this embodiment, an exemplary linker is derived from a natural or unnatural amino acid, amino acid analogue or amino acid mimetic, or a small peptide formed from one or more such species. For example, certain branched polymers found in the compounds of the invention have the formula:



(IV)

[0214] X^a is a linkage fragment that is formed by the reaction of a reactive functional group, e.g., $X^{3'}$, on a precursor of the branched polymeric modifying moiety and a reactive functional group on the sugar moiety, or a precursor to a linker. For example, when $X^{3'}$ is a carboxylic acid, it can be activated and bound directly to an amine group pendent from an amino-saccharide (e.g., Sia, $GalNH_2$, $GlcNH_2$, $ManNH_2$, etc.), forming a X^a that is an amide. Additional exemplary reactive functional groups and activated precursors are described hereinbelow. The index c represents an integer from 1 to 10. The other symbols have the same identity as those discussed above.

[0215] In another exemplary embodiment, X^a is a linking moiety formed with another linker:

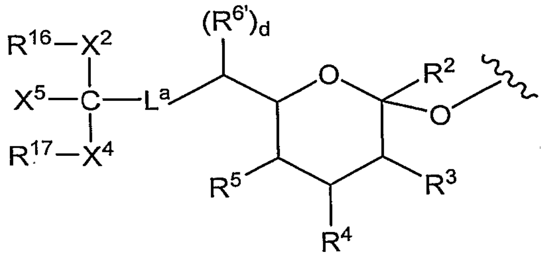


in which X^b is a second linkage fragment and is independently selected from those groups set forth for X^a, and, similar to L, L^1 is a bond, substituted or unsubstituted alkyl or substituted or unsubstituted heteroalkyl.

[0216] Exemplary species for X^a and X^b include S, SC(O)NH, HNC(O)S, SC(O)O, O, NH, NHC(O), C(O)NH and NHC(O)O, and OC(O)NH.

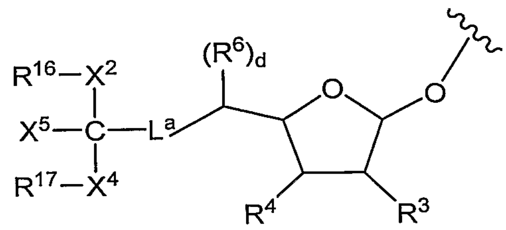
[0217] In another exemplary embodiment, X^4 is a peptide bond to R^{17}, which is an amino acid, di-peptide (e.g.,, Lys-Lys) or tri-peptide (e.g., Lys-Lys-Lys) in which the alpha-amine moiety(ies) and/or side chain heteroatom(s) are modified with a polymeric modifying moiety.

[0218] In a further exemplary embodiment, the peptide conjugates of the invention include a moiety, e.g., an R^{15} moiety that has a formula that is selected from:

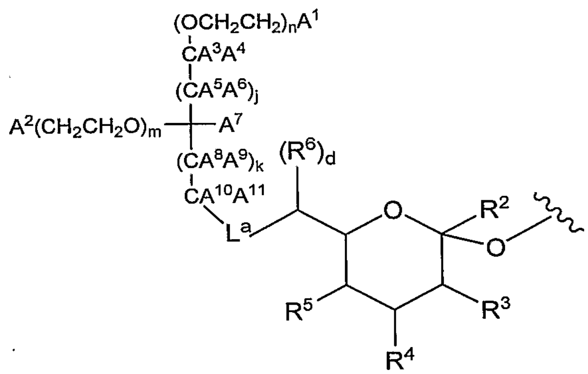


V

; and

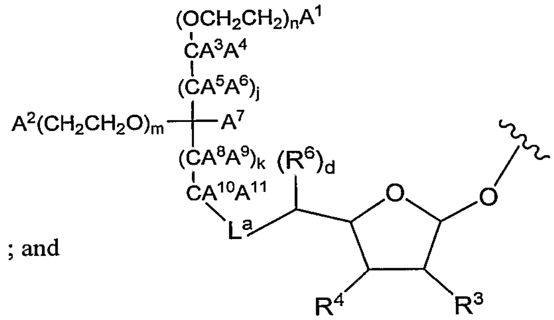


VI

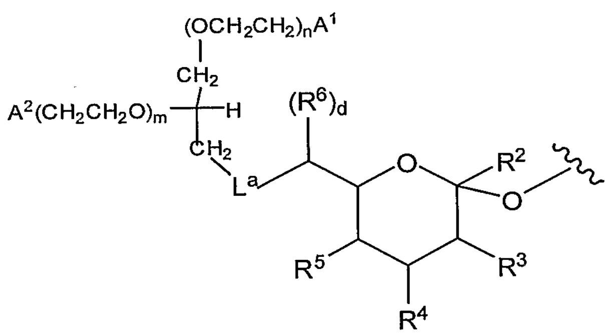


Va

; and

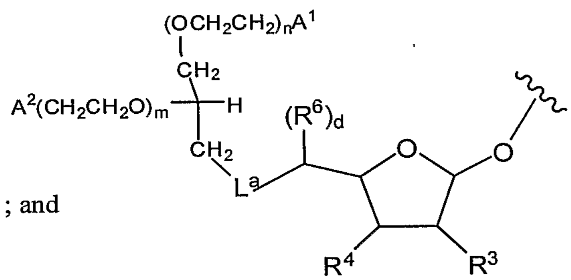


VIa



Vb

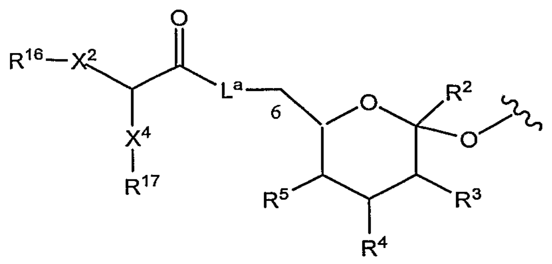
; and



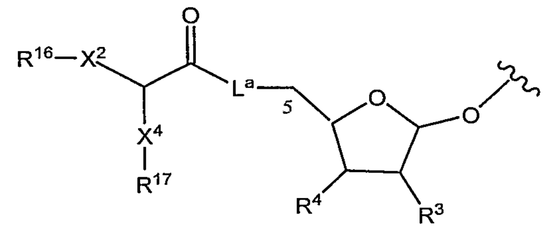
VIb

in which the identity of the radicals represented by the various symbols is the same as that discussed hereinabove. L^a is a bond or a linker as discussed above for L and L^1, e.g., substituted or unsubstituted alkyl or substituted or unsubstituted heteroalkyl moiety. In an exemplary embodiment, L^a is a moiety of the side chain of sialic acid that is functionalized with the polymeric modifying moiety as shown. Exemplary L^a moieties include substituted or unsubstituted alkyl chains that include one or more OH or NH_2.

[0219] In yet another exemplary embodiment, the invention provides peptide conjugates having a moiety, e.g., an R^{15} moiety with formula:



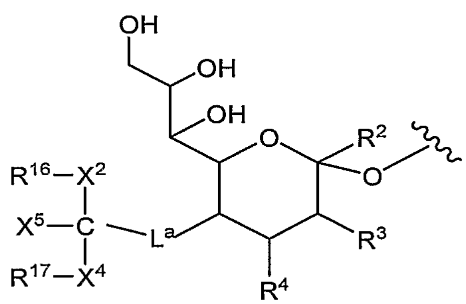
VI

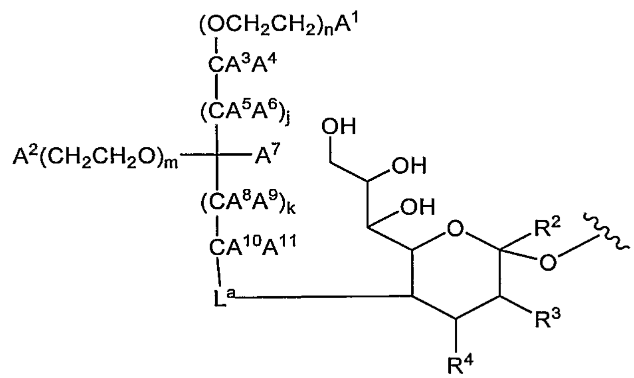


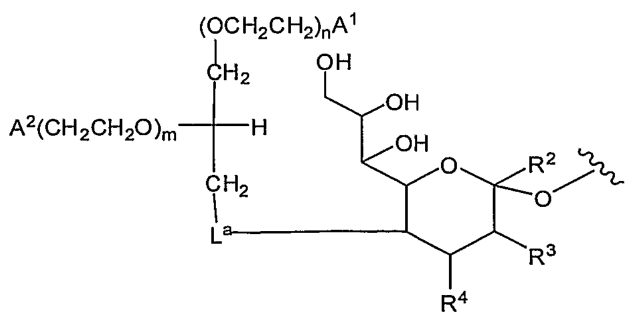
VII

The identity of the radicals represented by the various symbols is the same as that discussed hereinabove. As those of skill will appreciate, the linker arm in Formulae VI and VII is equally applicable to other modified sugars set forth herein. In exemplary embodiment, the species of Formulae VI and VII are the R^{15} moieties attached to the glycan structures set forth herein.

[0220] In yet another exemplary embodiment, the Factor VII/Factor VIIa peptide conjugate includes a R^{15} moiety with a formula which is a member selected from:





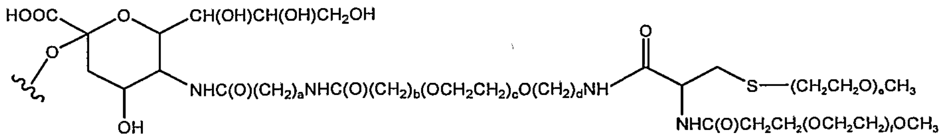


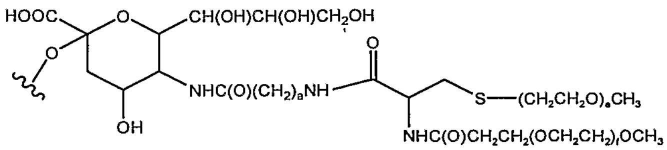
in which the identities of the radicals are as discussed above. An exemplary species for L^a is $-(CH_2)_jC(O)NH(CH_2)_hC(O)NH-$, in which the indices h and j are independently selected integers from 0 to 10. A further exemplary species is $-C(O)NH-$. The indices m and n are integers independently selected from 0 to 5000. A^1, A^2, A^3, A^4, A^5, A^6, A^7, A^8, A^9, A^{10} and A^{11} are members independently selected from H, substituted or unsubstituted alkyl, substituted or unsubstituted heteroalkyl, substituted or unsubstituted cycloalkyl, substituted or unsubstituted heterocycloalkyl, substituted or unsubstituted aryl, substituted or unsubstituted heteroaryl, $-NA^{12}A^{13}$, $-OA^{12}$ and $-SiA^{12}A^{13}$. A^{12} and A^{13} are members independently selected from substituted or unsubstituted alkyl, substituted or unsubstituted heteroalkyl, substituted or unsubstituted cycloalkyl, substituted or unsubstituted heterocycloalkyl, substituted or unsubstituted aryl, and substituted or unsubstituted heteroaryl.

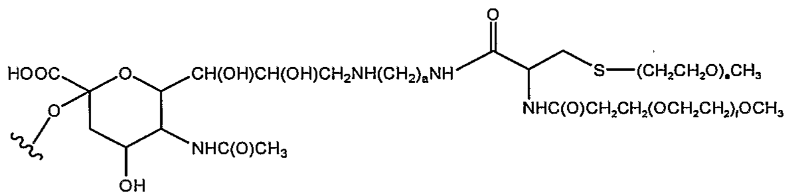
[0221] The embodiments of the invention set forth above are further exemplified by reference to species in which the polymer is a water-soluble polymer, particularly poly(ethylene glycol) ("PEG"), e.g., methoxy-poly(ethylene glycol). Those of skill will appreciate that the focus in the sections that follow is for clarity of illustration and the various motifs set forth using PEG as an exemplary polymer are equally applicable to species in which a polymer other than PEG is utilized.

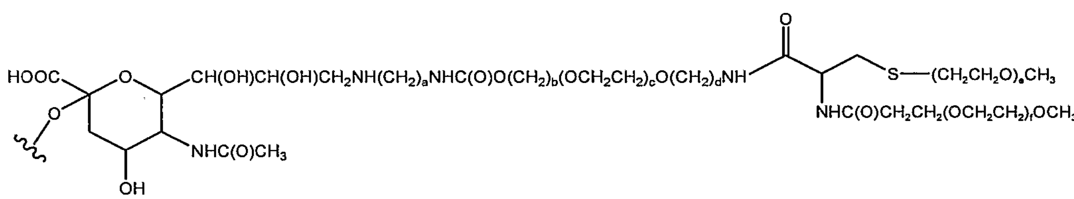
[0222] PEG of any molecular weight, e.g., 1 KDa, 2 KDa, 5 KDa, 10 KDa, 15 KDa, 20 KDa, 25 KDa, 30 KDa, 35 KDa, 40 KDa and 45 KDa is of use in the present invention.

[0223] In an exemplary embodiment, the R^{15} moiety has a formula that is a member selected from the group:


;

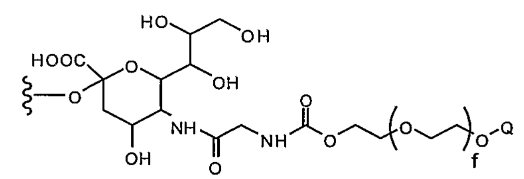

;

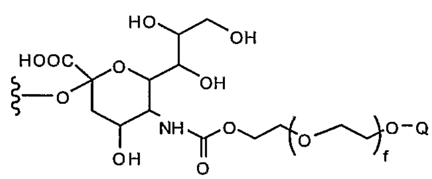

; and

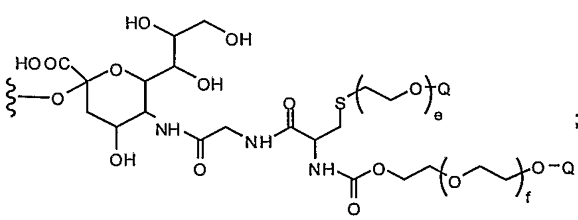


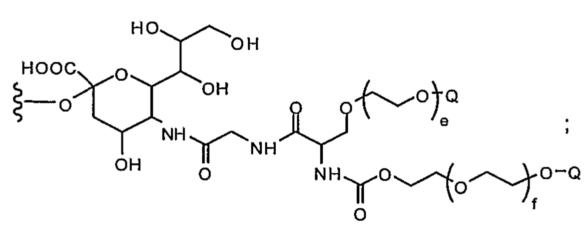
In each of the structures above, the linker fragment $-NH(CH_2)_a-$ can be present or absent.

[0224] In other exemplary embodiments, the peptide conjugate includes an R^{15} moiety selected from the group:

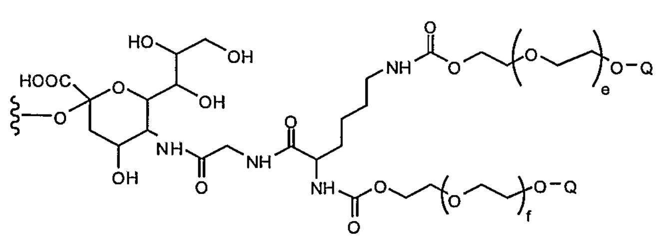

;


;


;

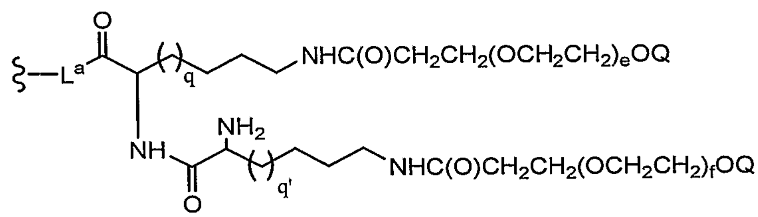

;

and

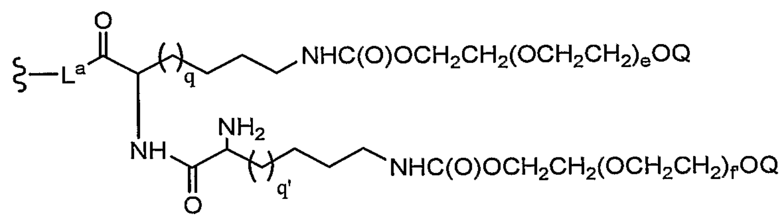


[0225] In each of the formulae above, the indices e and f are independently selected from the integers from 1 to 2500. In further exemplary embodiments, e and f are selected to provide a PEG moiety that is about 1 KDa, 2 KDa, 5 KDa, 10 KDa, 15 KDa, 20 KDa, 25 KDa, 30 KDa, 35 KDa, 40 KDa and 45 KDa. The symbol Q represents substituted or unsubstituted alkyl (e.g., C_1-C_6 alkyl, e.g., methyl), substituted or unsubstituted heteroalkyl or H.

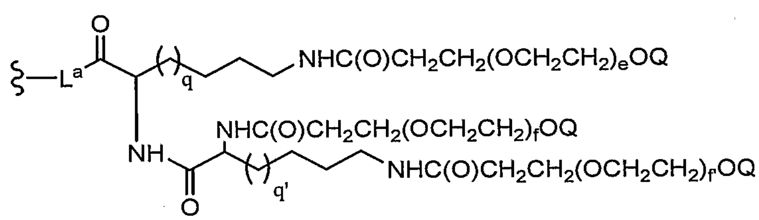
[0226] Other branched polymers have structures based on di-lysine (Lys-Lys) peptides, e.g.:



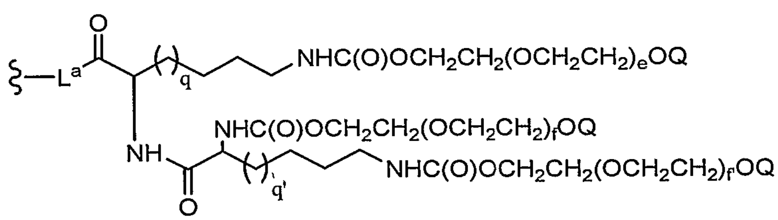
;



;

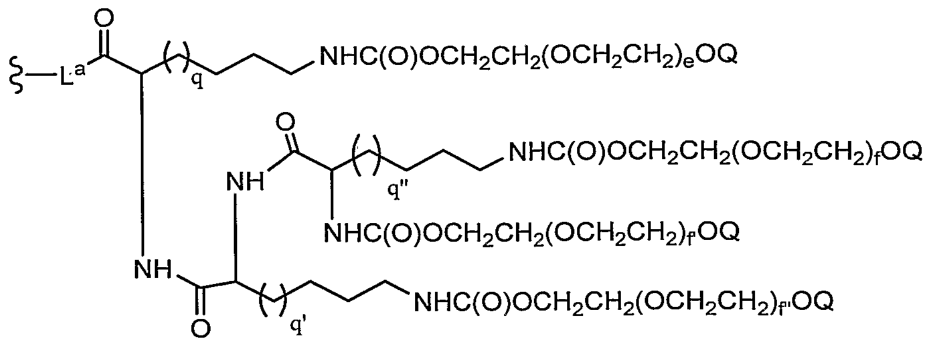


; and

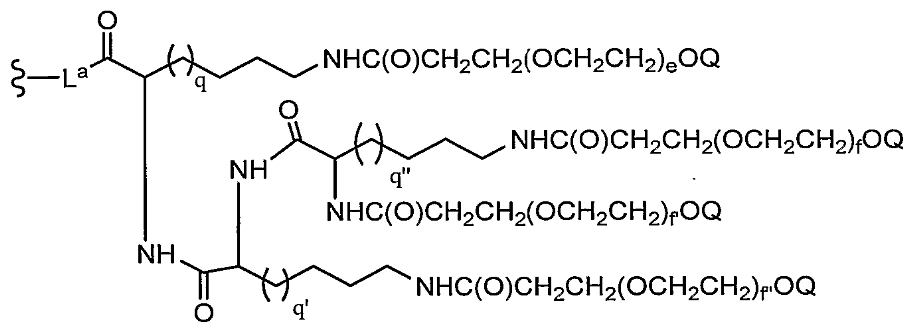


;

and tri-lysine peptides (Lys-Lys-Lys), e.g.:

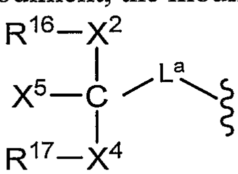


; and

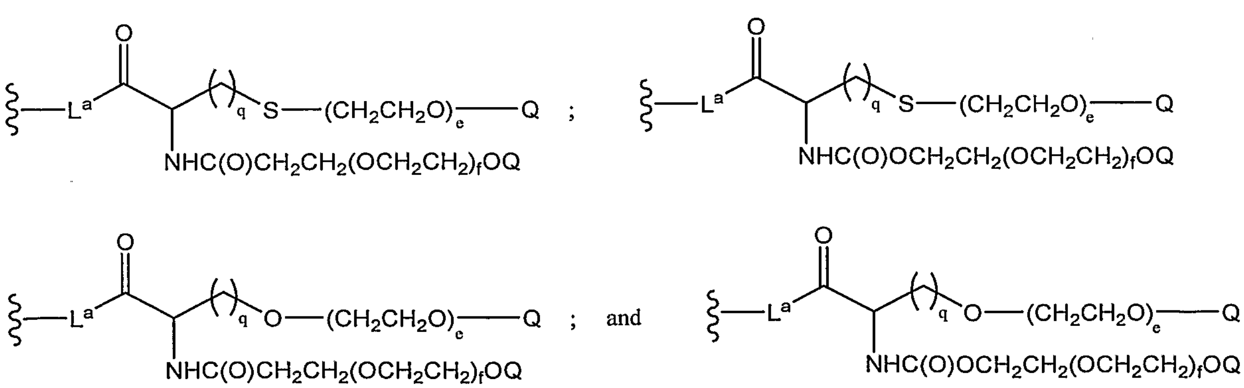


In each of the figures above, the indices e, f, f' and f'' represent integers independently selected from 1 to 2500. The indices q, q' and q'' represent integers independently selected from 1 to 20.

[0227] In another exemplary embodiment, the modifying group:

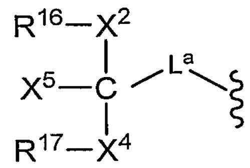


has a formula that is a member selected from:

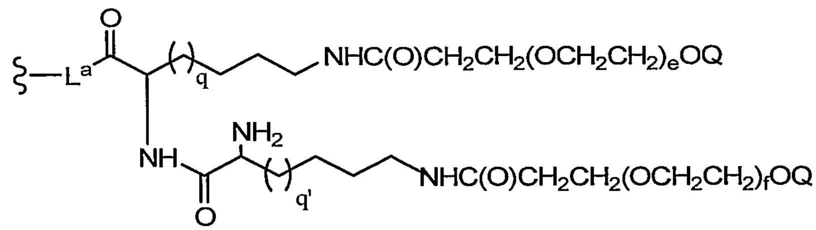


wherein Q is a member selected from H and substituted or unsubstituted C_1-C_6 alkyl. The indices e and f are integers independently selected from 1 to 2500, and the index q is an integer selected from 0 to 20.

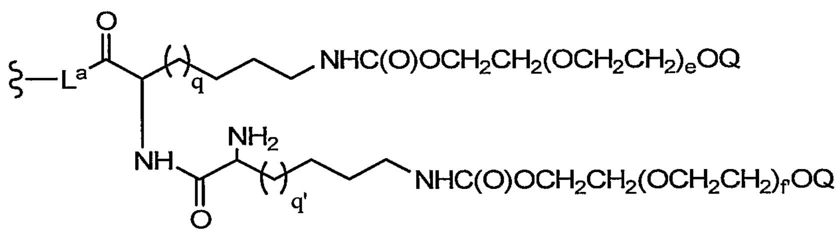
[0228] In another exemplary embodiment, the modifying group:



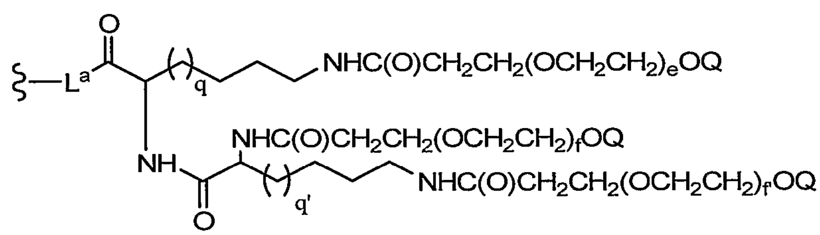
has a formula that is a member selected from:



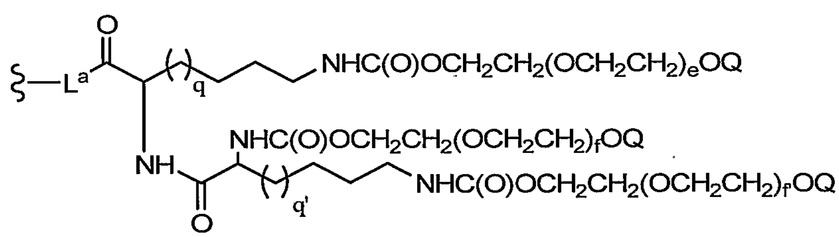
;



;

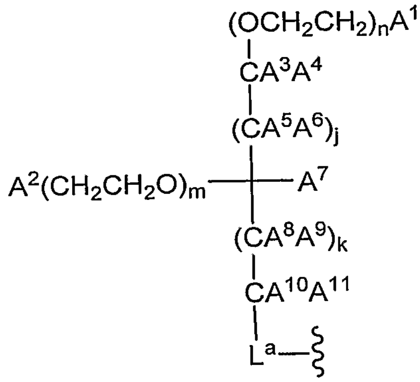


; and



wherein Q is a member selected from H and substituted or unsubstituted C_1-C_6 alkyl. The indices e, f and f' are integers independently selected from 1 to 2500, and q and q' are integers independently selected from 1 to 20.

[0229] In another exemplary embodiment, the branched polymer has a structure according to the following formula:

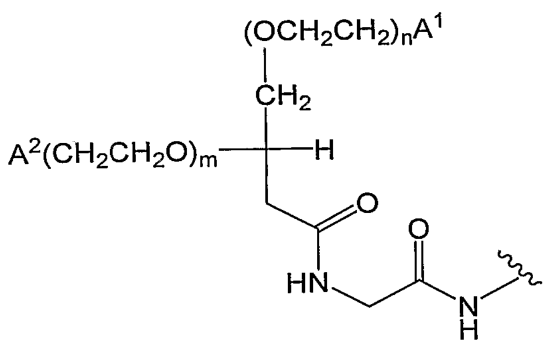


(IIIa)

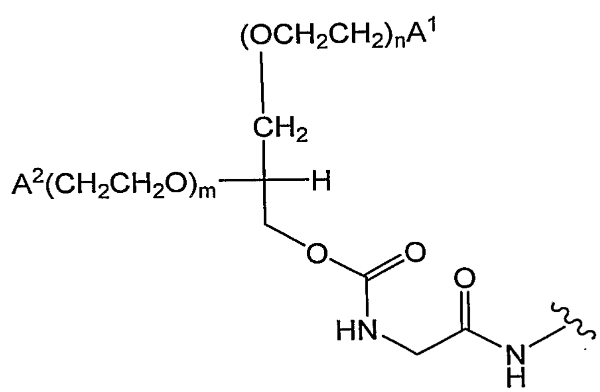
in which the indices m and n are integers independently selected from 0 to 5000. A^1, A^2, A^3, A^4, A^5, A^6, A^7, A^8, A^9, A^{10} and A^{11} are members independently selected from H, substituted or unsubstituted alkyl, substituted or unsubstituted heteroalkyl, substituted or unsubstituted cycloalkyl, substituted or unsubstituted heterocycloalkyl, substituted or unsubstituted aryl, substituted or unsubstituted heteroaryl, $-NA^{12}A^{13}$, $-OA^{12}$ and $-SiA^{12}A^{13}$. A^{12} and A^{13} are members independently selected from substituted or unsubstituted alkyl, substituted or unsubstituted heteroalkyl, substituted or unsubstituted cycloalkyl, substituted or unsubstituted heterocycloalkyl, substituted or unsubstituted aryl, and substituted or unsubstituted heteroaryl.

[0230] Formula IIIa is a subset of Formula III. The structures described by Formula IIIa are also encompassed by Formula III.

[0231] In an exemplary embodiment, the polymeric modifying group has a structure according to the following formulae:

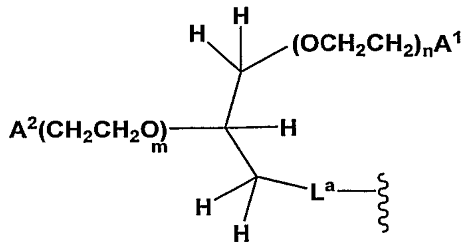


; and



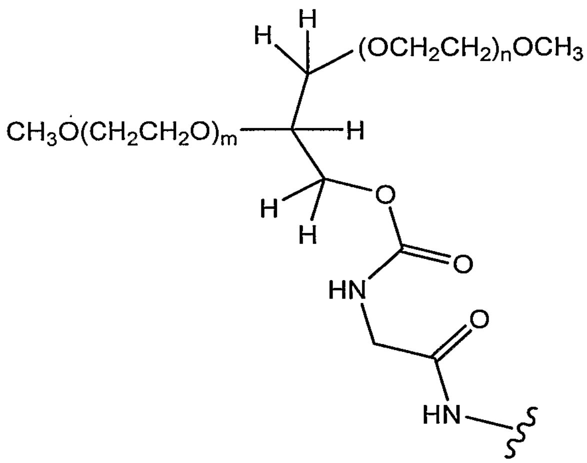
.

[0232] In another exemplary embodiment according to the formula above, the branched polymer has a structure according to the following formula:

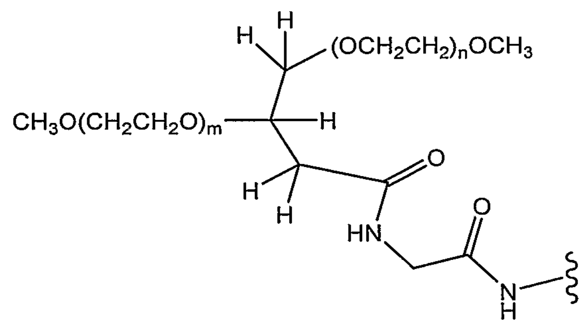


In an exemplary embodiment, A^1 and A^2 are members independently selected from -OH and $-OCH_3$.

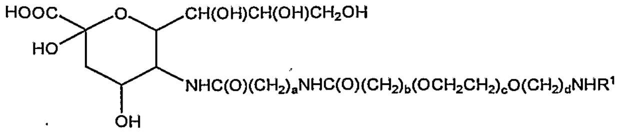
[0233] Exemplary polymeric modifying groups according to this embodiment include:

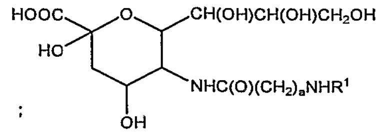


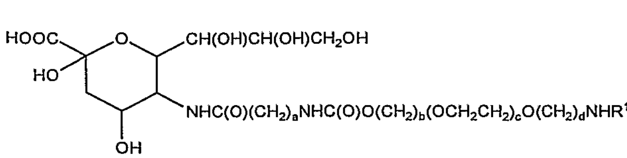
and

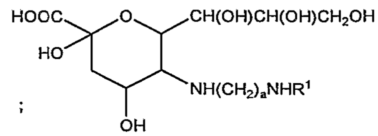


[0234] In an illustrative embodiment, the modified sugar is sialic acid and selected modified sugar compounds of use in the invention have the formulae:


;


;


;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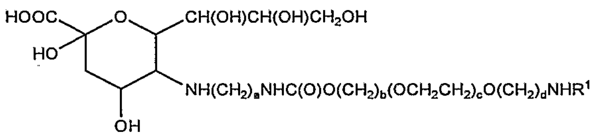

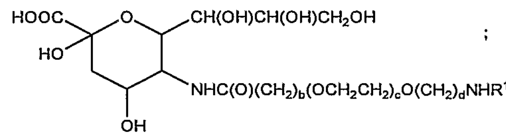
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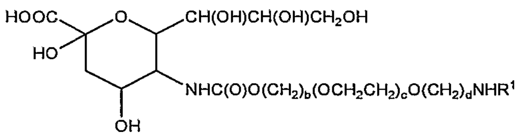
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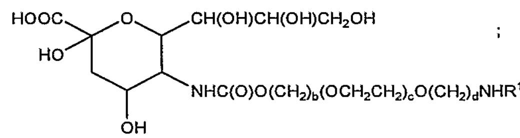
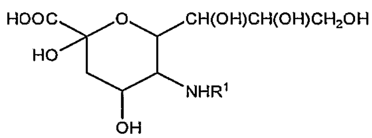
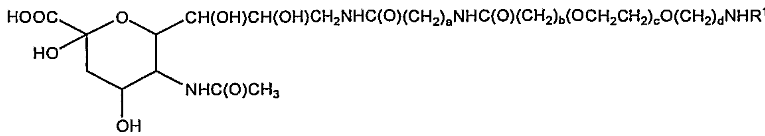

;

and


.

The indices a, b and d are integers from 0 to 20. The index c is an integer from 1 to 2500. The structures set forth above can be components of R^{15}.

[0235] In another illustrative embodiment, a primary hydroxyl moiety of the sugar is functionalized with the modifying group. For example, the 9-hydroxyl of sialic acid can be converted to the corresponding amine and functionalized to provide a compound according to the invention. Formulae according to this embodiment include: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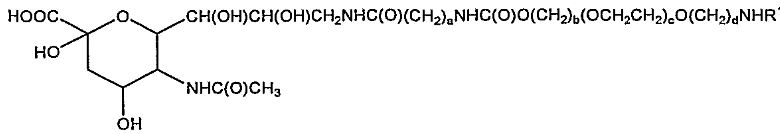
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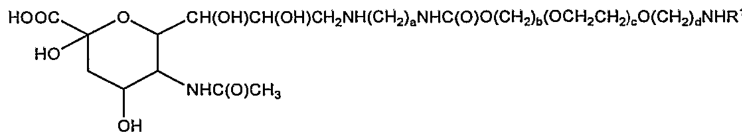
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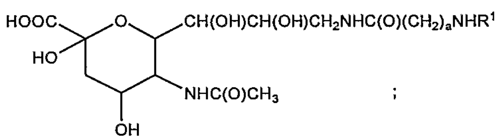
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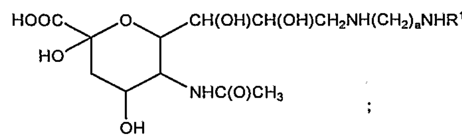
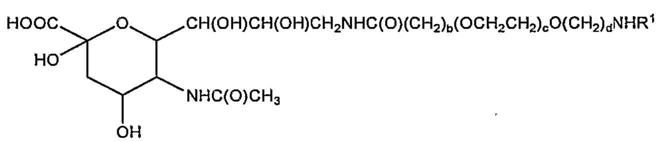
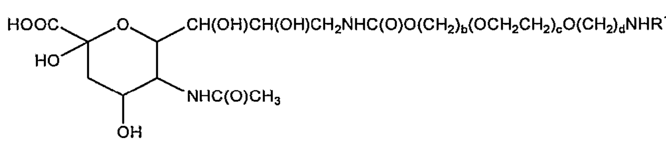
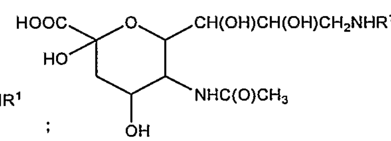

;


;

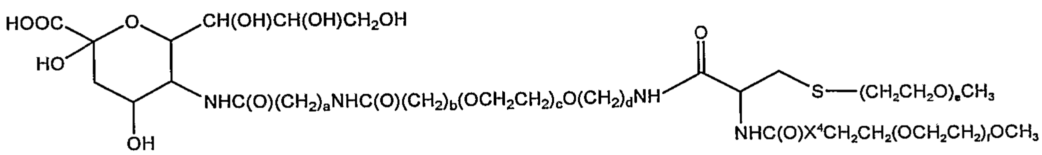

;


.

The structures set forth above can be components of R^{15}.

[0236] Although the present invention is exemplified in the preceding sections by reference to PEG, as those of skill will appreciate, an array of polymeric modifying moieties is of use in the compounds and methods set forth herein.

[0237] In selected embodiments, R^1 or L-R^1 is a branched PEG, for example, one of the species set forth above. In an exemplary embodiment, the branched PEG structure is based on a cysteine peptide. Illustrative modified sugars according to this embodiment include:



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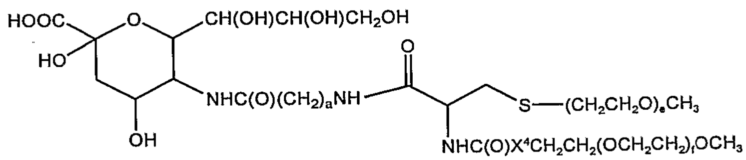

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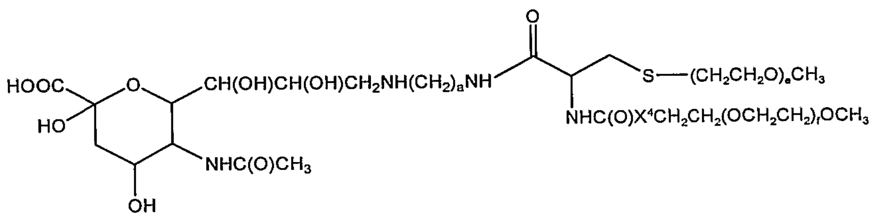
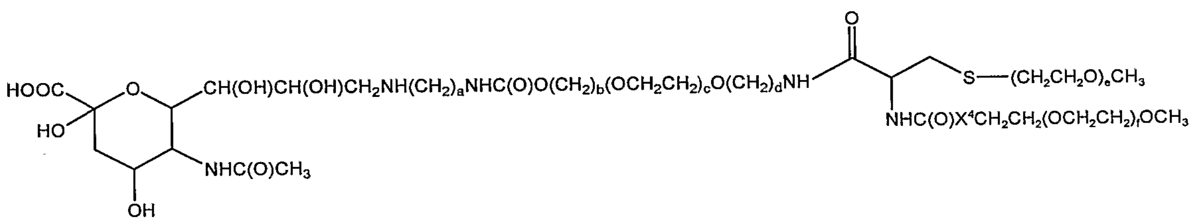
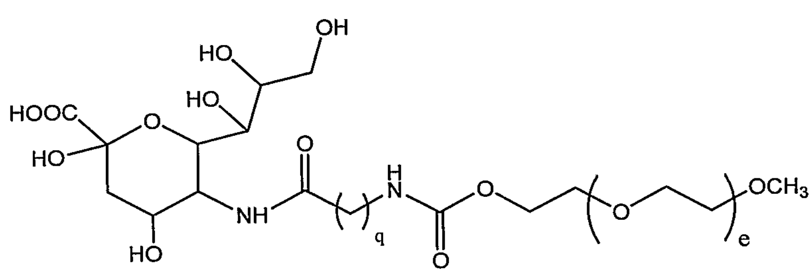


;



in which X^4 is a bond or O. In each of the structures above, the alkylamine linker $-(CH_2)_aNH-$ can be present or absent. The structures set forth above can be components of $R^{15}/R^{15'}$.

[0238] As discussed herein, the polymer-modified sialic acids of use in the invention may also be linear structures. Thus, the invention provides for conjugates that include a sialic acid moiety derived from a structure such as:

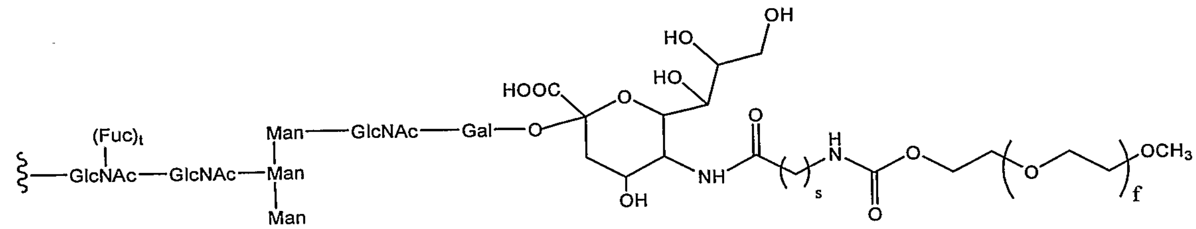


in which the indices q and e are as discussed above.

[0239] Exemplary modified sugars are modified with water-soluble or water-insoluble polymers. Examples of useful polymer are further exemplified below.

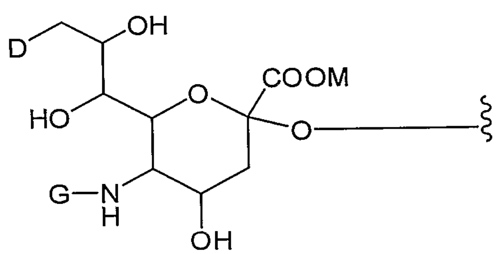
[0240] In another exemplary embodiment, the peptide is derived from insect cells, remodeled by adding GlcNAc and Gal to the mannose core and glycopegylated using a sialic

acid bearing a linear PEG moiety, affording a Factor VII/Factor VIIa peptide that comprises at least one moiety having the formula:



in which the index t is an integer from 0 to 1; the index s represents an integer from 1 to 10; and the index f represents an integer from 1 to 2500.

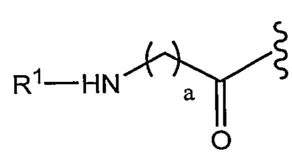
[0241] In one embodiment, the present invention provides a peptide conjugate comprising the following glycosyl linking group:



;

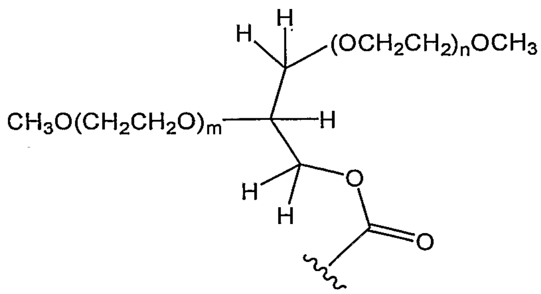
D is a member selected from -OH and R^1-L-HN-;G is a member selected from R^1-L- and -C(O)(C_1-C_6)alkyl-R^1; R^1 is a moiety comprising a member selected from a straight-chain poly(ethylene glycol) residue and branched poly(ethylene glycol) residue; and M is a member selected from H, a salt counterion and a single negative charge; L is a linker which is a member selected from a bond, substituted or unsubstituted alkyl and substituted or unsubstituted heteroalkyl. In an exemplary embodiment, when D is OH, G is R^1-L-. In another exemplary embodiment, when G is –C(O)(C_1-C_6)alkyl, D is R^1-L-NH-.

[0242] In an exemplary embodiment, L-R^1 has the formula:



wherein a is an integer selected from 0 to 20.

[0243] In an exemplary embodiment, R^1 has a structure that is a member selected from:



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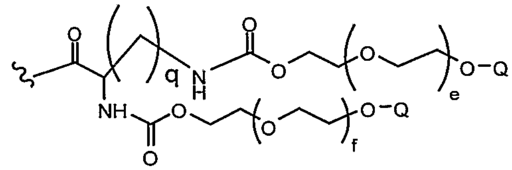

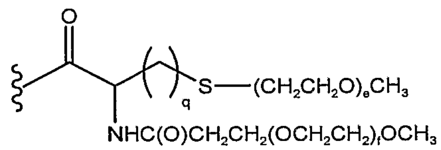

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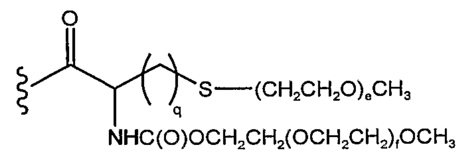
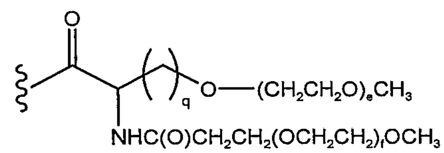
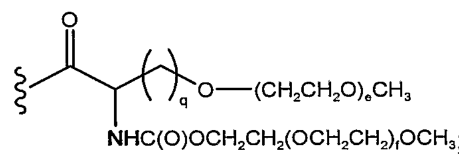


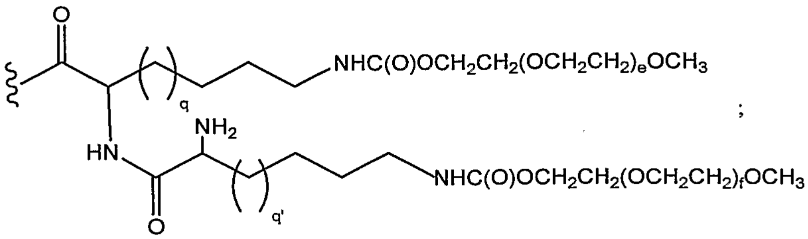


; and



wherein e, f, m and n are integers independently selected from 1 to 2500; and q is an integer selected from 0 to 20.

[0244] In an exemplary embodiment, R^1 has a structure that is a member selected from:



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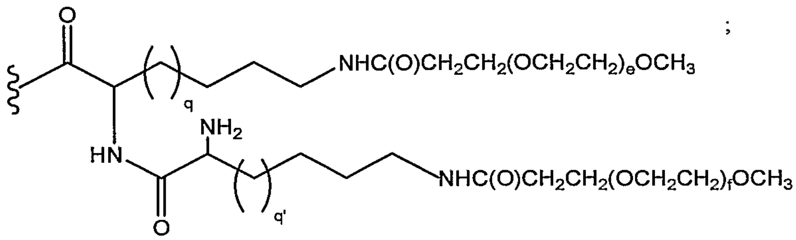

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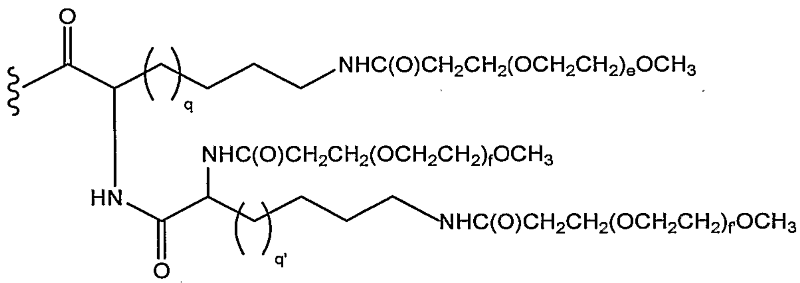
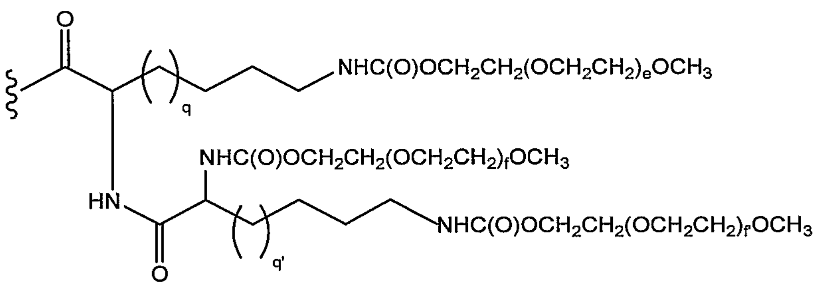
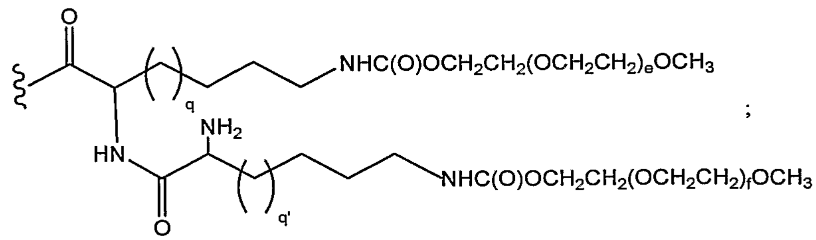


; and



wherein e, f and f' are integers independently selected from 1 to 2500; and q and q' are integers independently selected from 1 to 20.

[0245] In another exemplary embodiment, R^1 has a structure that is a member selected from:



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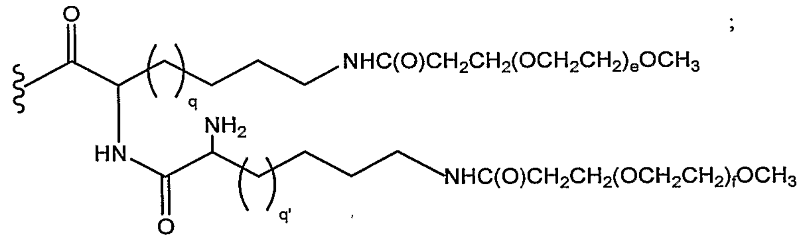

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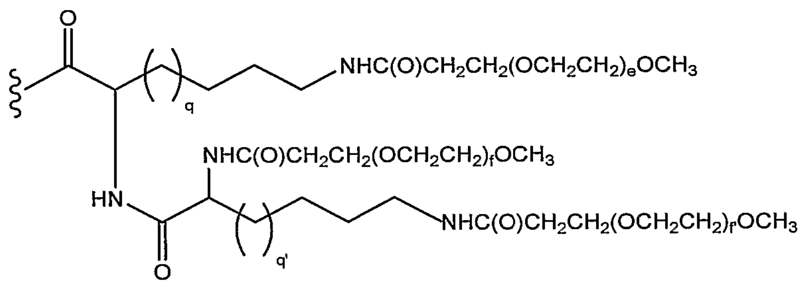
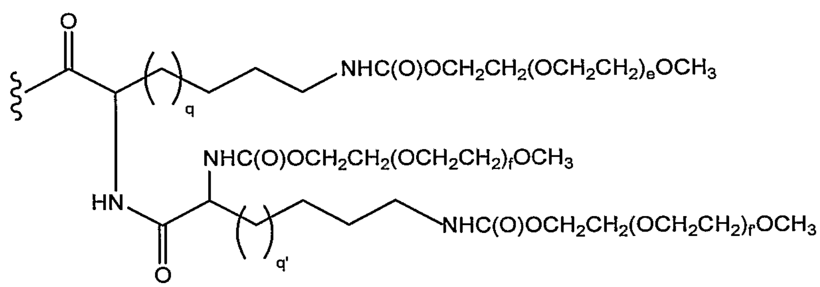


; and



wherein e, f and f' are integers independently selected from 1 to 2500; and q and q' are integers independently selected from 1 to 20.

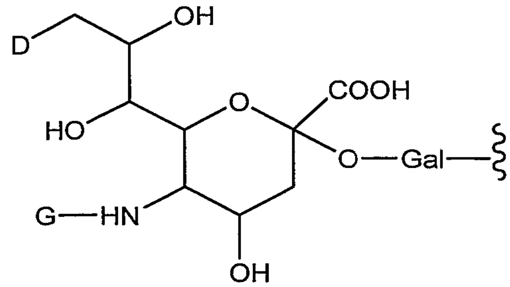
[0246] In another exemplary embodiment, R^1 has a structure that is a member selected from:

$$\text{}\!\!-C(O)CH_2CH_2(OCH_2CH_2)_eOCH_3 \text{ ; and}$$

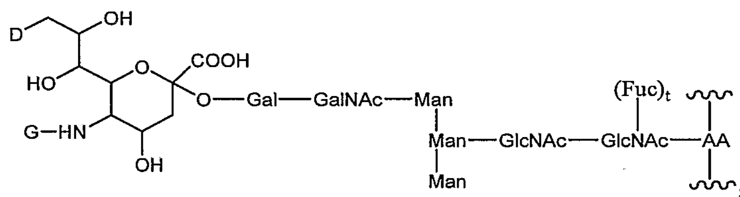
$$\text{}\!\!-C(O)OCH_2CH_2(OCH_2CH_2)_fOCH_3$$

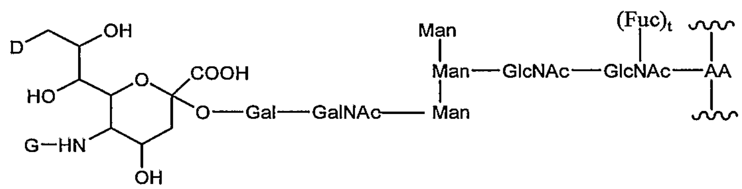
wherein e and f are integers independently selected from 1 to 2500

[0247] In another exemplary embodiment, the glycosyl linker has the formula:



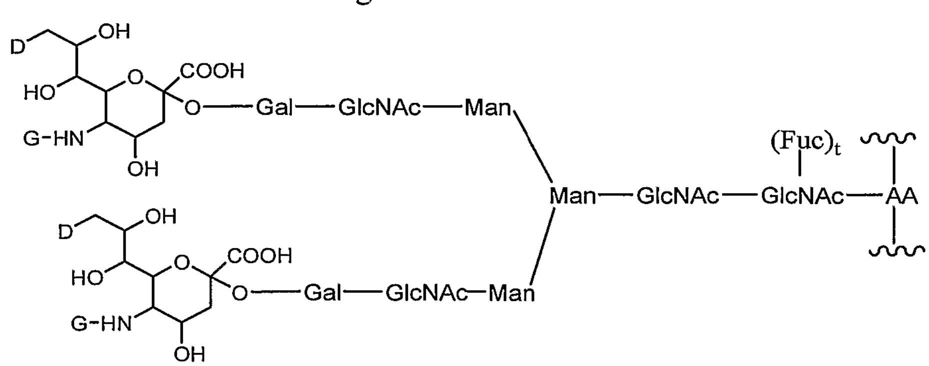
[0248] In another exemplary embodiment, the peptide conjugate comprises at least one of said glycosyl linker according to a formula selected from:

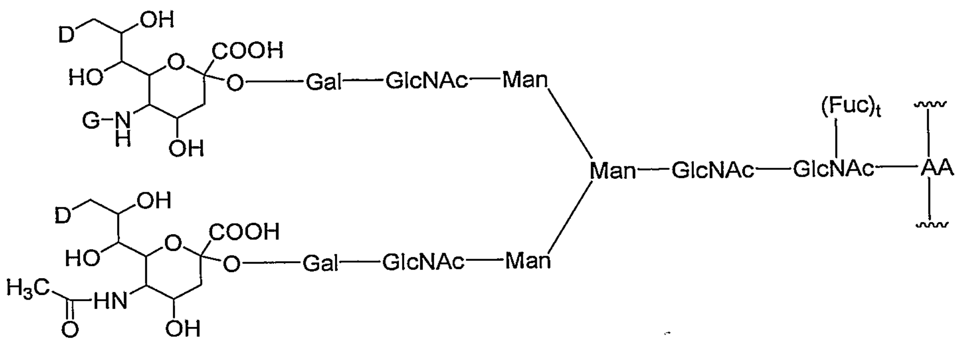




wherein AA is an amino acid residue of said peptide conjugate and t is an integer selected from 0 and 1.

[0249] In another exemplary embodiment, the peptide conjugate comprises at least one of said glycosyl linker wherein each of said glycosyl linker has a structure which is a member independently selected from the following formulae:





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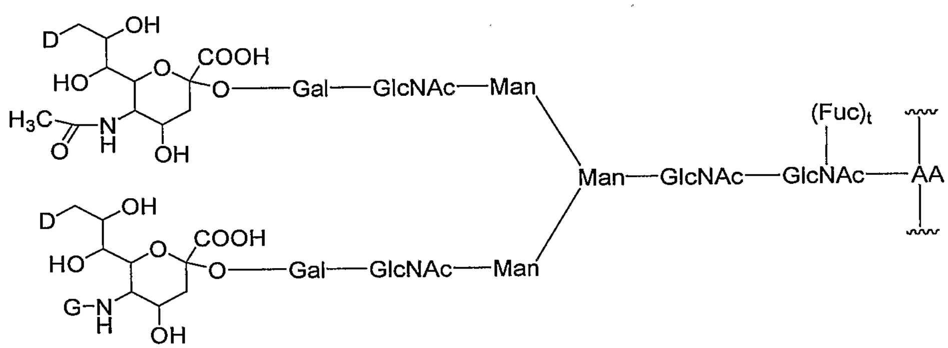

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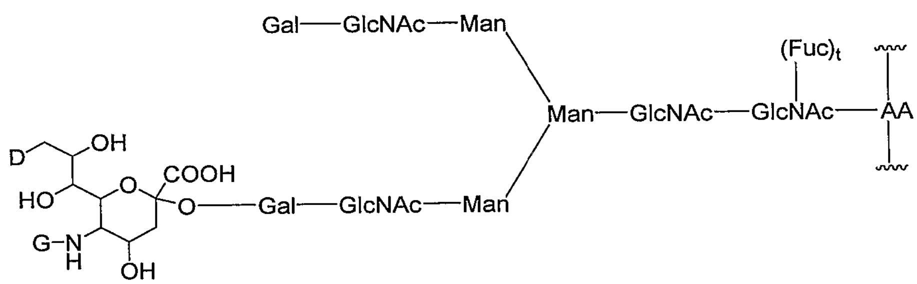

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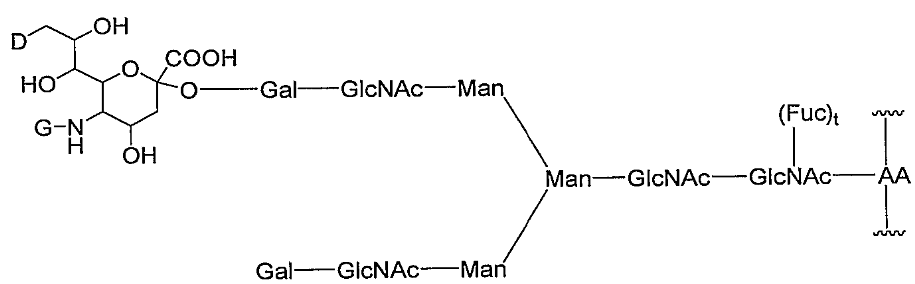
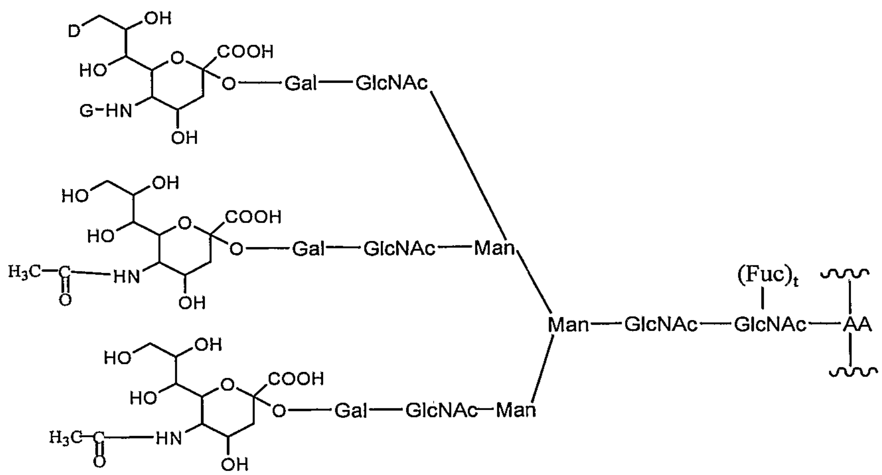
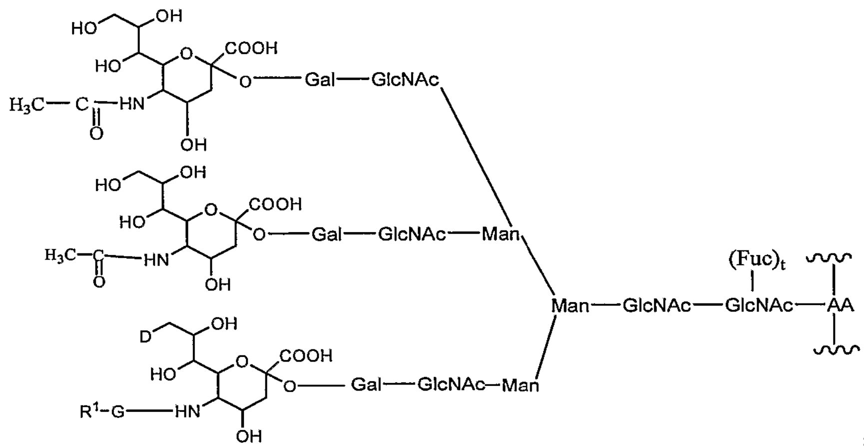


wherein AA is an amino acid residue of said peptide conjugate and t is an integer selected from 0 and 1.

[0250] In another exemplary embodiment, the peptide conjugate comprises at least one of said glycosyl linker according to a formula selected from:





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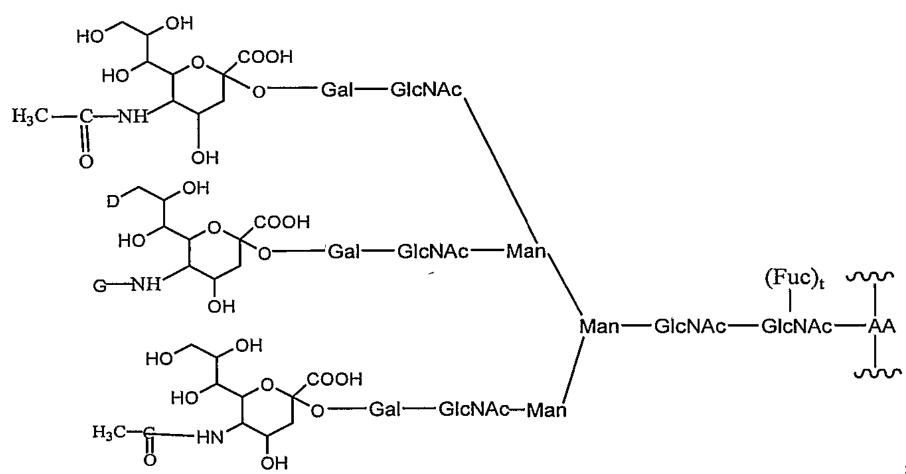

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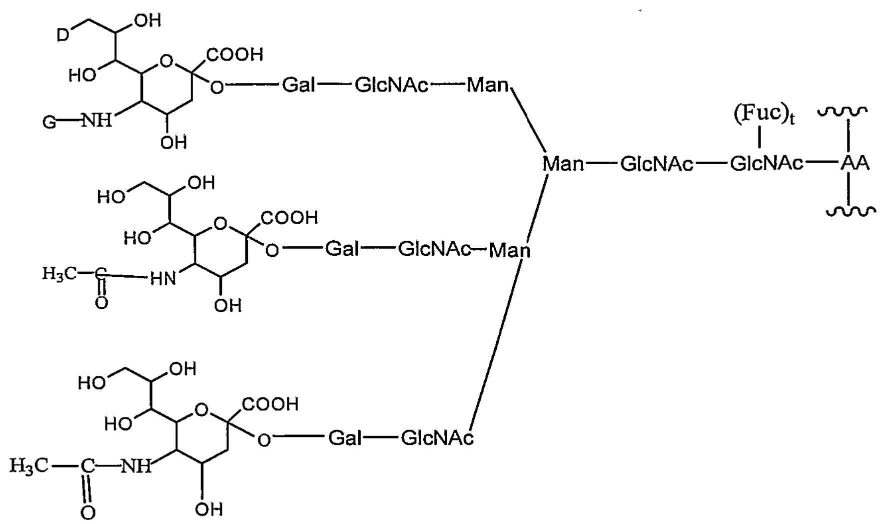

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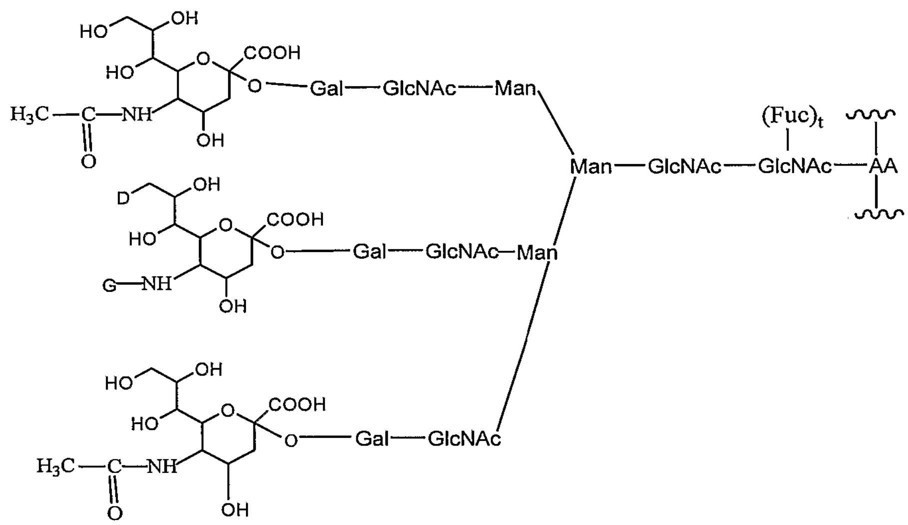
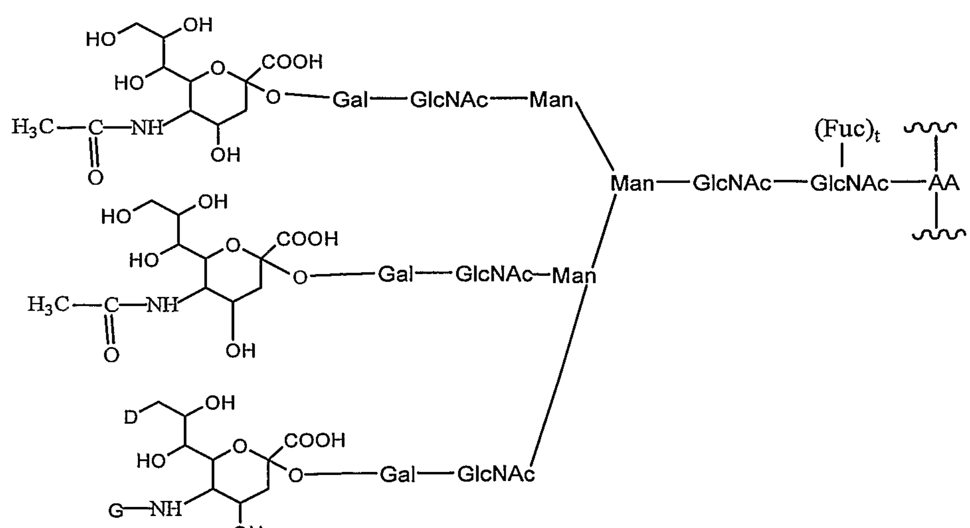


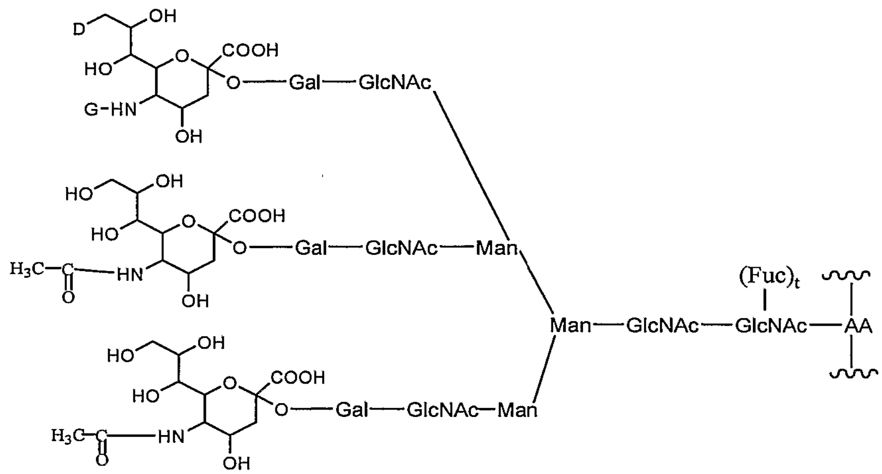
, and



wherein AA is an amino acid residue of said peptide conjugate and t is an integer selected from 0 and 1. In an exemplary embodiment, a member selected from 0 and 2 of the sialyl

moieties which do not comprise G are absent. In an exemplary embodiment, a member selected from 1 and 2 of the sialyl moieties which do not comprise G are absent.

[0251] In another exemplary embodiment, the peptide conjugate comprises at least one of said glycosyl linker according to a formula selected from:



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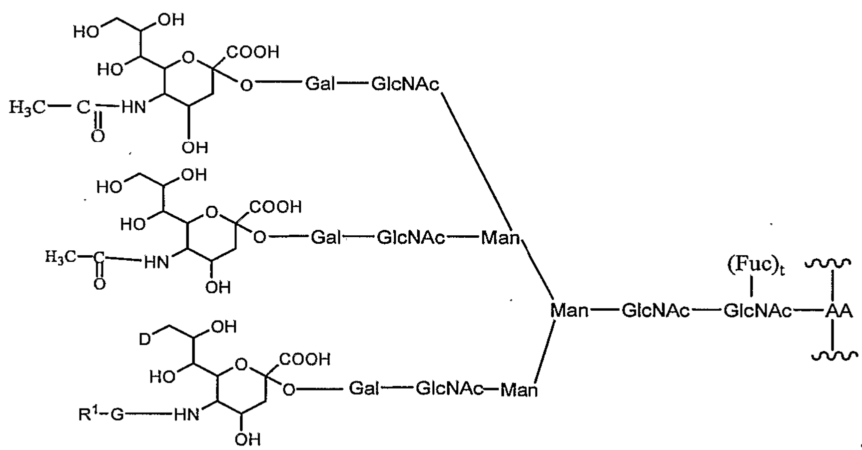

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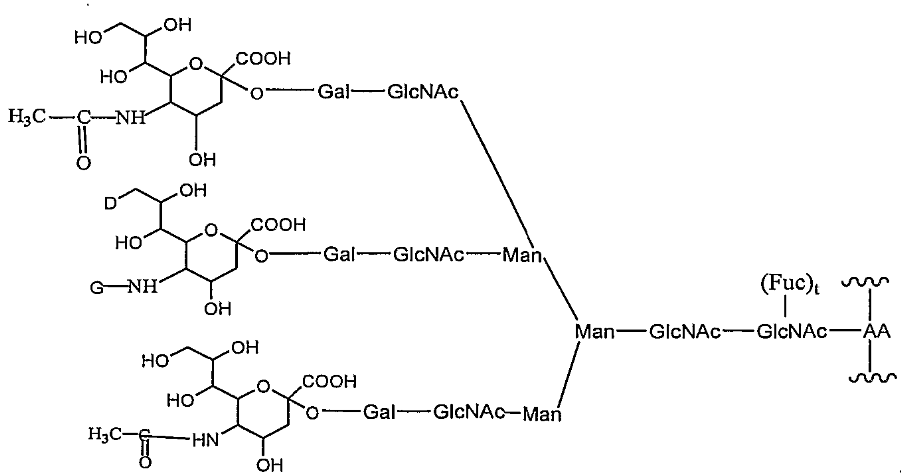

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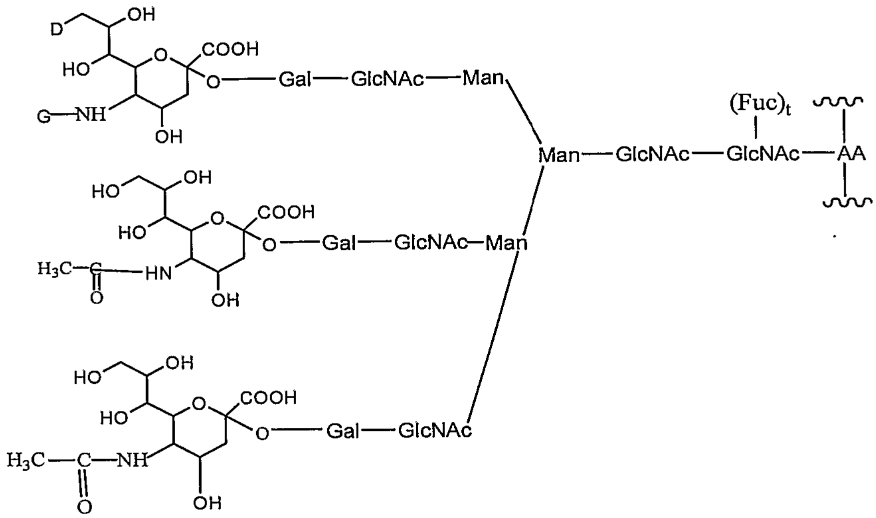

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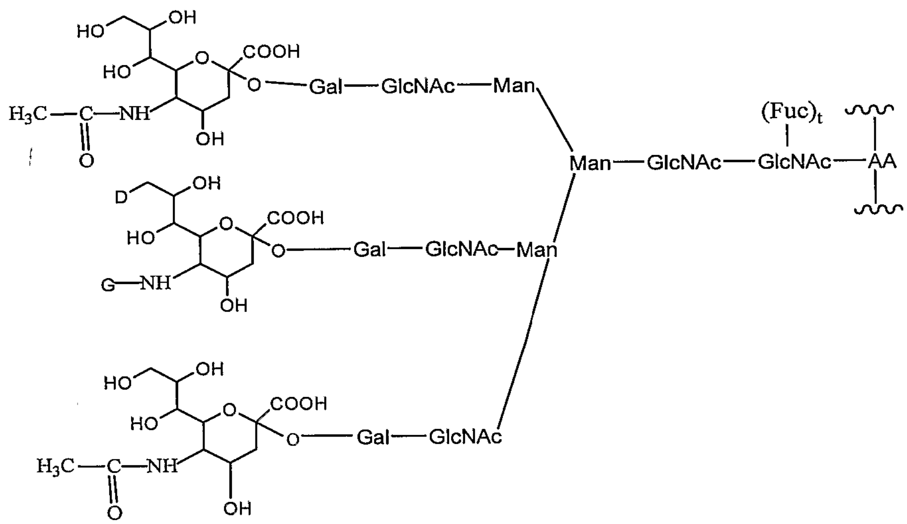
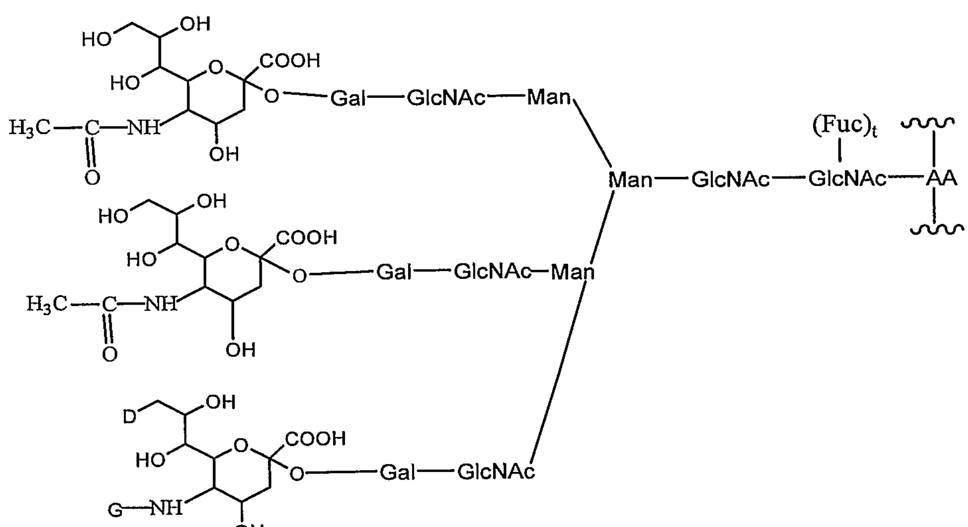


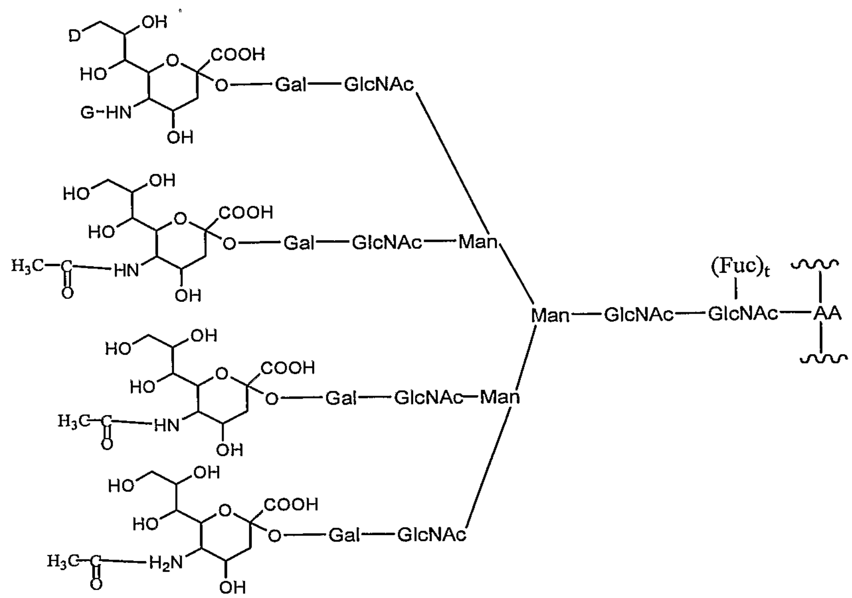
, and



wherein AA is an amino acid residue of said peptide conjugate and t is an integer selected from 0 and 1. In an exemplary embodiment, a member selected from 0 and 2 of the sialyl

moieties which do not comprise G are absent. In an exemplary embodiment, a member selected from 1 and 2 of the sialyl moieties which do not comprise G are absent. .

[0252] In another exemplary embodiment, the peptide conjugate comprises at least one said glycosyl linker according to a formula selected from:



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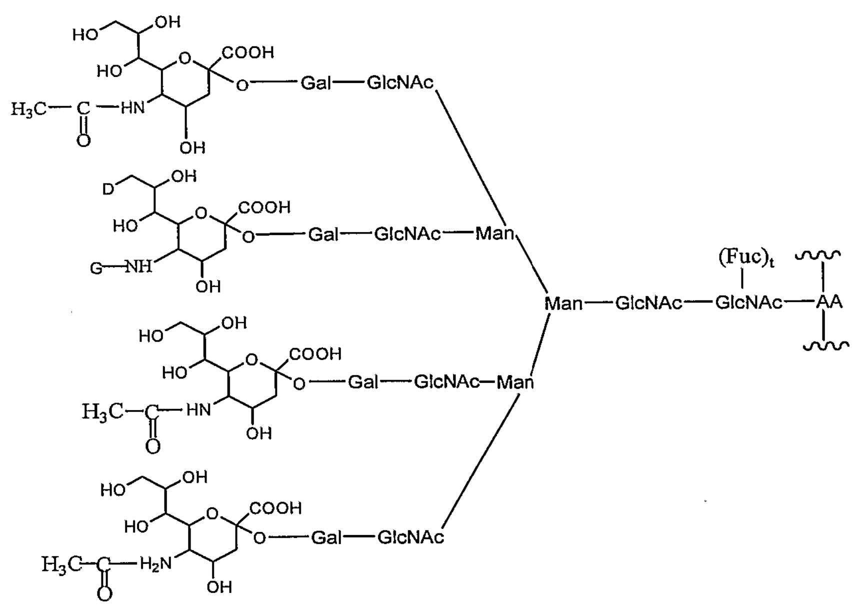
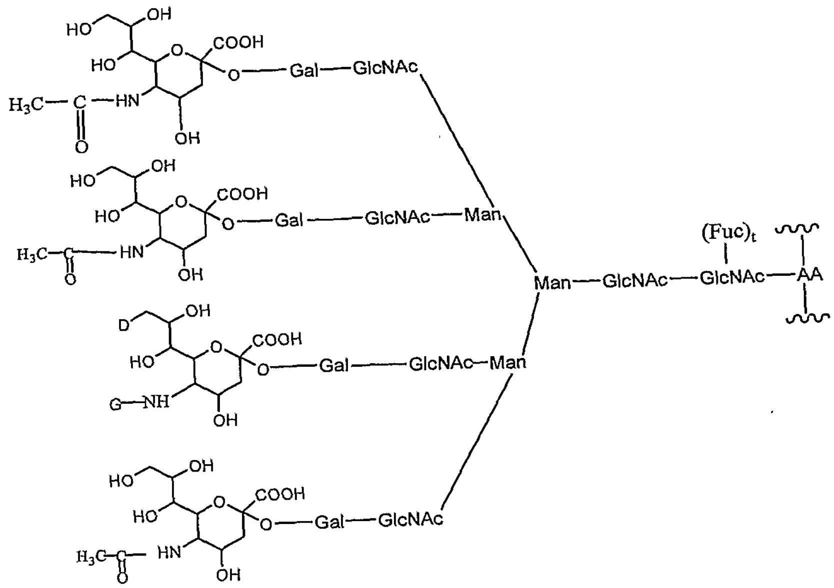
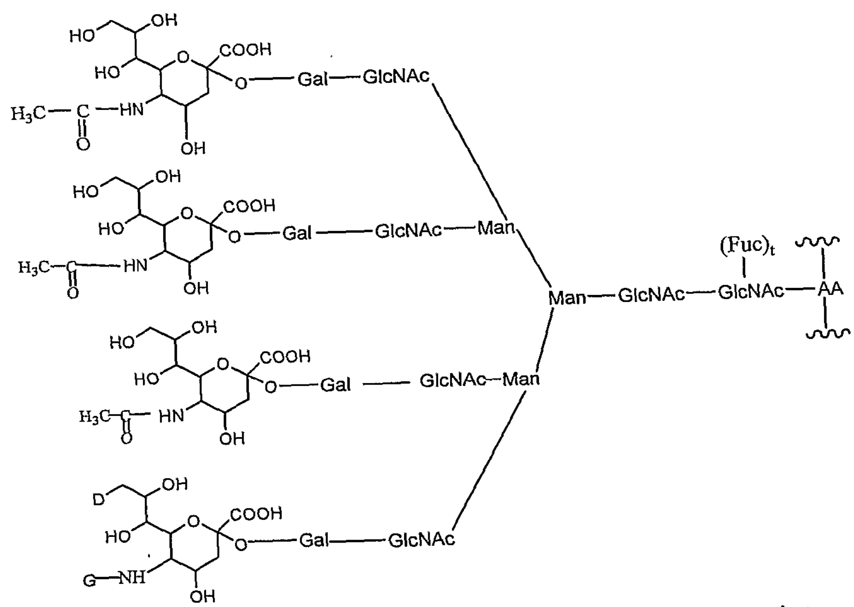




, and



wherein AA is an amino acid residue of said peptide conjugate and t is an integer selected from 0 and 1. In an exemplary embodiment, a member selected from 0 and 2 of the sialyl moieties which do not comprise G are absent. In an exemplary embodiment, a member selected from 1 and 2 of the sialyl moieties which do not comprise G are absent. .

[0253] In another exemplary embodiment,the Factor VII/Factor VIIa peptide has the amino acid sequence of SEQ. ID. NO:1. In another exemplary embodiment,the glycosyl linker is attached to said Factor VII/Factor VIIa peptide through an amino acid residue selected from serine and threonine.

[0254] In another exemplary embodiment, the asparagine residue is a member selected from N152, N322 and combinations thereof.

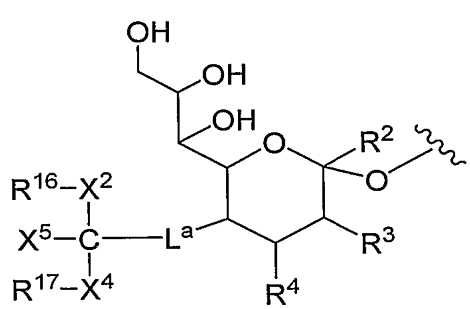
[0255] In another exemplary embodiment, the Factor VIIa peptide is a bioactive Factor VIIa peptide.

[0256] In another exemplary embodiment, the glycosyl linker is attached to said Factor VII/Factor VIIa peptide through an amino acid residue which is an asparagine residue.

[0257] In another exemplary embodiment, the invention provides a Factor VII/Factor VIIa peptide which is produced in a suitable host. The invention also provides methods of expressing this peptide. In another exemplary embodiment, the host is a mammalian expression system.

[0258] In another exemplary embodiment, the invention provides a method of treating a condition in a subject in need thereof, said condition characterized by compromised clotting potency in said subject, said method comprising the step of administering to the subject an amount of the Factor VII/Factor VIIa peptide conjugate of invention, effective to ameliorate said condition in said subject. In another exemplary embodiment, the method comprises administering to said mammal an amount of the Factor VII/Factor VIIa peptide conjugate produced according to the methods described herein.

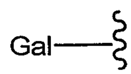
[0259] In another aspect, the invention provides a meethod of making a Factor VII/Factor VIIa peptide conjugate comprising a glycosyl linker comprising a modified sialyl residue having the formula:



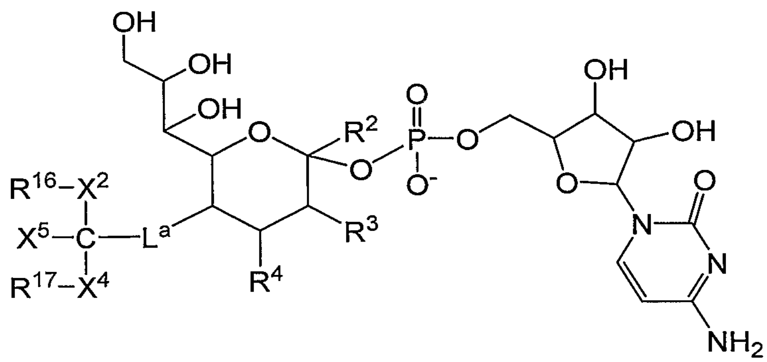
;

wherein R^2 is H, CH_2OR^7, $COOR^7$ or OR^7. R^7 represents H, substituted or unsubstituted alkyl or substituted or unsubstituted heteroalkyl. R^3 and R^4 are members independently selected from H, substituted or unsubstituted alkyl, OR^8, $NHC(O)R^9$. R^8 and R^9 are independently selected from H, substituted or unsubstituted alkyl, substituted or unsubstituted heteroalkyl or sialic acid. R^{16} and R^{17} are independently selected polymeric arms. X^2 and X^4

are independently selected linkage fragments joining polymeric moieties R^{16} and R^{17} to C. X^5 is a non-reactive group and L^a is a linker group. The method comprises (a) contacting a Factor VII/FactorVIIa peptide comprising the glycosyl moiety:

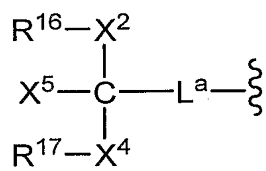


with a PEG-sialic acid donor moiety having the formula:

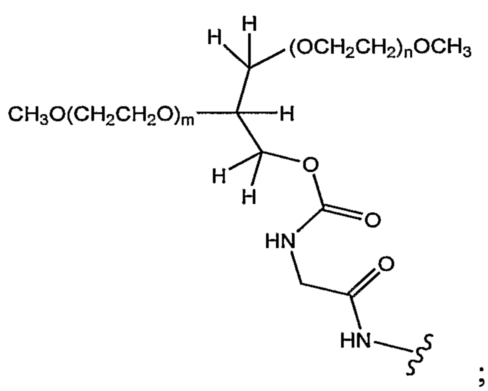


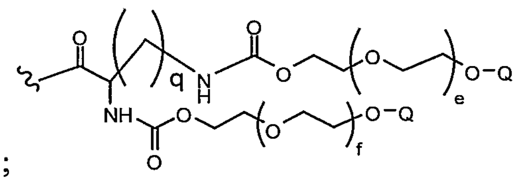
and an enzyme that transfers PEG-sialic acid onto the Gal of said glycosyl moiety, under conditions appropriate for said transfer.

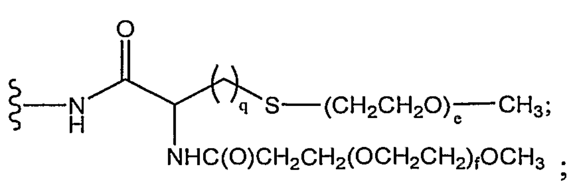
[0260] In another exemplary embodiment, the moiety:

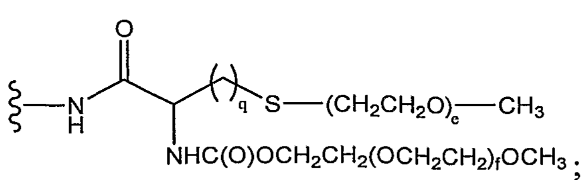


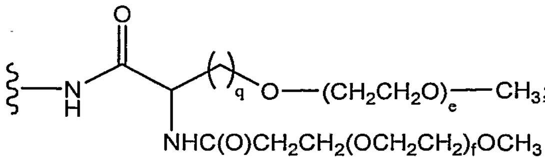
has a formula that is a member selected from:







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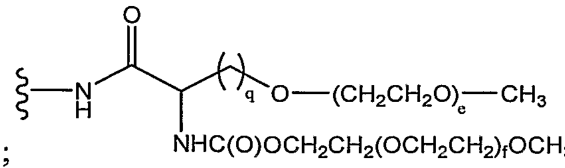
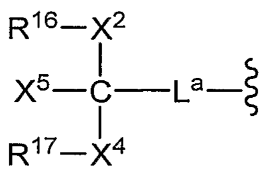


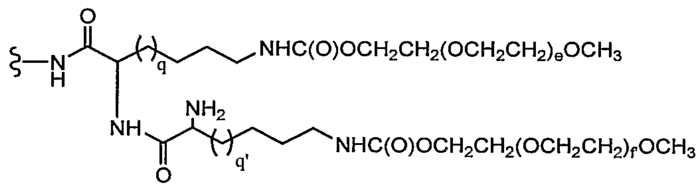
wherein e, f, m and n are integers independently selected from 1 to 2500; and

q is an integer selected from 0 to 20.

[0261] In another exemplary embodiment, the moiety:



has a formula that is a member selected from: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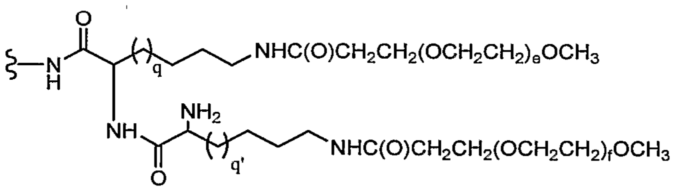
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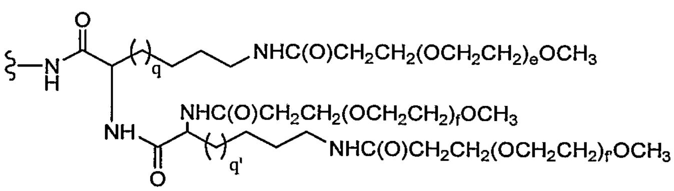
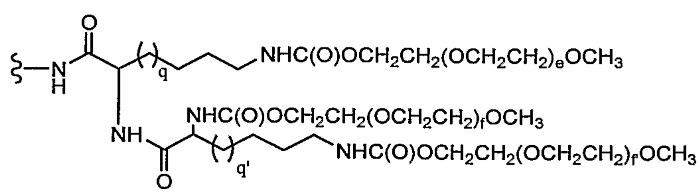
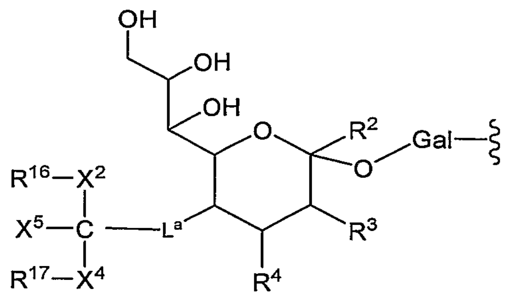

; and



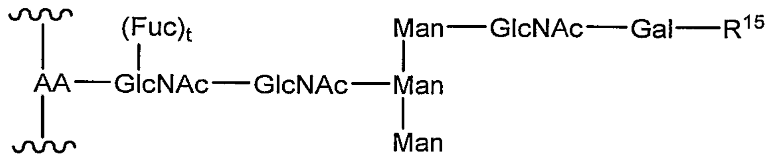
wherein e, f and f' are integers independently selected from 1 to 2500; and
q and q' are integers independently selected from 1 to 20.

[0262] In another exemplary embodiment, the glycosyl linker comprises the formula:



.

[0263] In another exemplary embodiment, the Factor VII/Factor VIIa peptide conjugate comprises at least one glycosyl linker having the formula:



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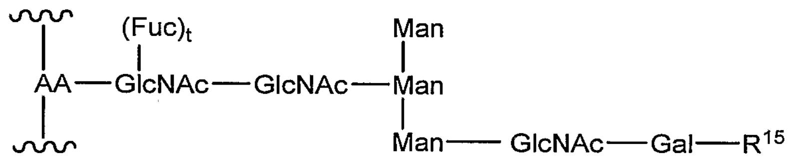

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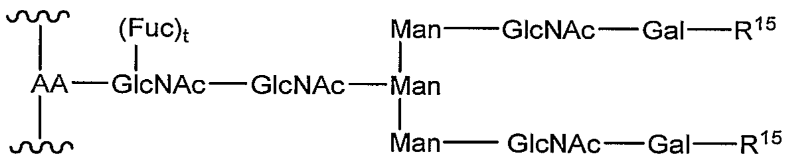

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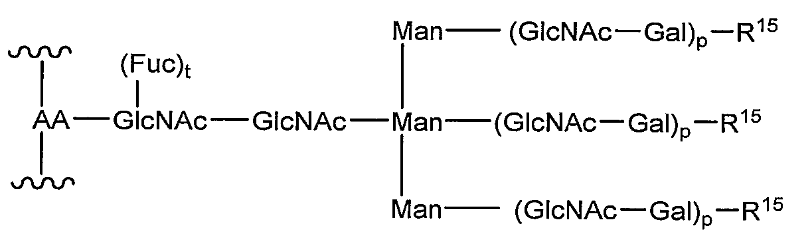

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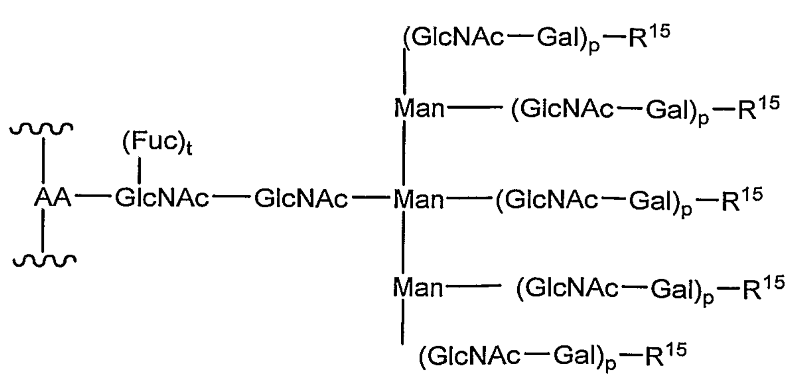


wherein AA is an amino acid residue of said peptide; t is an integer selected from 0 and 1; and R^{15} is the modified sialyl moiety.

[0264] In another exemplary embodiment, the Factor VII/Factor VIIa peptide has the amino acid sequence of SEQ. ID. NO:1.

[0265] In another exemplary embodiment, the glycosyl linker is attached to said Factor VII/Factor VIIa peptide through an amino acid residue which is an asparagine residue.

[0266] In another exemplary embodiment, the asparagine residue is a member selected from N152, N322 and combinations thereof.

[0267] In another exemplary embodiment, the Factor VIIa peptide is a bioactive Factor VIIa peptide.

[0268] In another exemplary embodiment, the method comprises, prior to step (a): (b) expressing the Factor VII/Factor VIIa peptide in a suitable host.

[0269] In another aspect, the invention provides a method of treating a condition in a subject in need thereof, said condition characterized by compromised clotting potency in said subject, said method comprising the step of administering to the subject an amount of the Factor VII/Factor VIIa peptide conjugate produced according to the methods described herein, effective to ameliorate said condition in said subject. In another exemplary embodiment, the method comprises administering to said mammal an amount of the Factor VII/Factor VIIa peptide conjugate produced according to the methods described herein.

[0270] In another aspect, the invention provides a method of synthesizing a Factor VII or Factor VIIa peptide conjugate, said method comprising combining a) sialidase; b) enzyme which is a member selected from glycosyltransferase, exoglycosidase and endoglycosidase; c) modified sugar/modified sialyl residue; d) Factor VII/Factor VIIa peptide thus synthesizing said Factor VII or Factor VIIa peptide conjugate. In an exemplary embodiment, the combining is for a time less than 10 hours. In another exemplary embodiment, the invention further comprising a capping step.

II. D. iv. ***Water-Insoluble Polymers***

[0271] In another embodiment, analogous to those discussed above, the modified sugars include a water-insoluble polymer, rather than a water-soluble polymer. The conjugates of the invention may also include one or more water-insoluble polymers. This embodiment of the invention is illustrated by the use of the conjugate as a vehicle with which to deliver a

therapeutic peptide in a controlled manner. Polymeric drug delivery systems are known in the art. *See*, for example, Dunn *et al.*, Eds. POLYMERIC DRUGS AND DRUG DELIVERY SYSTEMS, ACS Symposium Series Vol. 469, American Chemical Society, Washington, D.C. 1991. Those of skill in the art will appreciate that substantially any known drug delivery system is applicable to the conjugates of the present invention.

[0272] The motifs forth above for R^1, L-R^1, R^{15}, $R^{15'}$ and other radicals are equally applicable to water-insoluble polymers, which may be incorporated into the linear and branched structures without limitation utilizing chemistry readily accessible to those of skill in the art.

[0273] Representative water-insoluble polymers include, but are not limited to, polyphosphazines, poly(vinyl alcohols), polyamides, polycarbonates, polyalkylenes, polyacrylamides, polyalkylene glycols, polyalkylene oxides, polyalkylene terephthalates, polyvinyl ethers, polyvinyl esters, polyvinyl halides, polyvinylpyrrolidone, polyglycolides, polysiloxanes, polyurethanes, poly(methyl methacrylate), poly(ethyl methacrylate), poly(butyl methacrylate), poly(isobutyl methacrylate), poly(hexyl methacrylate), poly(isodecyl methacrylate), poly(lauryl methacrylate), poly(phenyl methacrylate), poly(methyl acrylate), poly(isopropyl acrylate), poly(isobutyl acrylate), poly(octadecyl acrylate) polyethylene, polypropylene, poly(ethylene glycol), poly(ethylene oxide), poly (ethylene terephthalate), poly(vinyl acetate), polyvinyl chloride, polystyrene, polyvinyl pyrrolidone, pluronics and polyvinylphenol and copolymers thereof.

[0274] Synthetically modified natural polymers of use in conjugates of the invention include, but are not limited to, alkyl celluloses, hydroxyalkyl celluloses, cellulose ethers, cellulose esters, and nitrocelluloses. Particularly preferred members of the broad classes of synthetically modified natural polymers include, but are not limited to, methyl cellulose, ethyl cellulose, hydroxypropyl cellulose, hydroxypropyl methyl cellulose, hydroxybutyl methyl cellulose, cellulose acetate, cellulose propionate, cellulose acetate butyrate, cellulose acetate phthalate, carboxymethyl cellulose, cellulose triacetate, cellulose sulfate sodium salt, and polymers of acrylic and methacrylic esters and alginic acid.

[0275] These and the other polymers discussed herein can be readily obtained from commercial sources such as Sigma Chemical Co. (St. Louis, MO.), Polysciences (Warrenton, PA.), Aldrich (Milwaukee, WI.), Fluka (Ronkonkoma, NY), and BioRad (Richmond, CA), or else synthesized from monomers obtained from these suppliers using standard techniques.

[0276] Representative biodegradable polymers of use in the conjugates of the invention include, but are not limited to, polylactides, polyglycolides and copolymers thereof, poly(ethylene terephthalate), poly(butyric acid), poly(valeric acid), poly(lactide-co-caprolactone), poly(lactide-co-glycolide), polyanhydrides, polyorthoesters, blends and copolymers thereof. Of particular use are compositions that form gels, such as those including collagen, pluronics and the like.

[0277] The polymers of use in the invention include "hybrid' polymers that include water-insoluble materials having within at least a portion of their structure, a bioresorbable molecule. An example of such a polymer is one that includes a water-insoluble copolymer, which has a bioresorbable region, a hydrophilic region and a plurality of crosslinkable functional groups per polymer chain.

[0278] For purposes of the present invention, "water-insoluble materials" includes materials that are substantially insoluble in water or water-containing environments. Thus, although certain regions or segments of the copolymer may be hydrophilic or even water-soluble, the polymer molecule, as a whole, does not to any substantial measure dissolve in water.

[0279] For purposes of the present invention, the term "bioresorbable molecule" includes a region that is capable of being metabolized or broken down and resorbed and/or eliminated through normal excretory routes by the body. Such metabolites or break down products are preferably substantially non-toxic to the body.

[0280] The bioresorbable region may be either hydrophobic or hydrophilic, so long as the copolymer composition as a whole is not rendered water-soluble. Thus, the bioresorbable region is selected based on the preference that the polymer, as a whole, remains water-insoluble. Accordingly, the relative properties, *i.e.*, the kinds of functional groups contained by, and the relative proportions of the bioresorbable region, and the hydrophilic region are selected to ensure that useful bioresorbable compositions remain water-insoluble.

[0281] Exemplary resorbable polymers include, for example, synthetically produced resorbable block copolymers of poly(α-hydroxy-carboxylic acid)/poly(oxyalkylene, (*see*, Cohn *et al.*, U.S. Patent No. 4,826,945). These copolymers are not crosslinked and are water-soluble so that the body can excrete the degraded block copolymer compositions. *See*, Younes *et al.*, *J Biomed. Mater. Res.* **21**: 1301-1316 (1987); and Cohn *et al.*, *J Biomed. Mater. Res.* **22**: 993-1009 (1988).

[0282] Presently preferred bioresorbable polymers include one or more components selected from poly(esters), poly(hydroxy acids), poly(lactones), poly(amides), poly(ester-amides), poly (amino acids), poly(anhydrides), poly(orthoesters), poly(carbonates), poly(phosphazines), poly(phosphoesters), poly(thioesters), polysaccharides and mixtures thereof. More preferably still, the biosresorbable polymer includes a poly(hydroxy) acid component. Of the poly(hydroxy) acids, polylactic acid, polyglycolic acid, polycaproic acid, polybutyric acid, polyvaleric acid and copolymers and mixtures thereof are preferred.

[0283] In addition to forming fragments that are absorbed *in vivo* ("bioresorbed"), preferred polymeric coatings for use in the methods of the invention can also form an excretable and/or metabolizable fragment.

[0284] Higher order copolymers can also be used in the present invention. For example, Casey *et al.*, U.S. Patent No. 4,438,253, which issued on March 20, 1984, discloses tri-block copolymers produced from the transesterification of poly(glycolic acid) and an hydroxyl-ended poly(alkylene glycol). Such compositions are disclosed for use as resorbable monofilament sutures. The flexibility of such compositions is controlled by the incorporation of an aromatic orthocarbonate, such as tetra-p-tolyl orthocarbonate into the copolymer structure.

[0285] Other polymers based on lactic and/or glycolic acids can also be utilized. For example, Spinu, U.S. Patent No. 5,202,413, which issued on April 13, 1993, discloses biodegradable multi-block copolymers having sequentially ordered blocks of polylactide and/or polyglycolide produced by ring-opening polymerization of lactide and/or glycolide onto either an oligomeric diol or a diamine residue followed by chain extension with a di-functional compound, such as, a diisocyanate, diacylchloride or dichlorosilane.

[0286] Bioresorbable regions of coatings useful in the present invention can be designed to be hydrolytically and/or enzymatically cleavable. For purposes of the present invention, "hydrolytically cleavable" refers to the susceptibility of the copolymer, especially the bioresorbable region, to hydrolysis in water or a water-containing environment. Similarly, "enzymatically cleavable" as used herein refers to the susceptibility of the copolymer, especially the bioresorbable region, to cleavage by endogenous or exogenous enzymes.

[0287] When placed within the body, the hydrophilic region can be processed into excretable and/or metabolizable fragments. Thus, the hydrophilic region can include, for example, polyethers, polyalkylene oxides, polyols, poly(vinyl pyrrolidine), poly(vinyl

alcohol), poly(alkyl oxazolines), polysaccharides, carbohydrates, peptides, proteins and copolymers and mixtures thereof. Furthermore, the hydrophilic region can also be, for example, a poly(alkylene) oxide. Such poly(alkylene) oxides can include, for example, poly(ethylene) oxide, poly(propylene) oxide and mixtures and copolymers thereof.

[0288] Polymers that are components of hydrogels are also useful in the present invention. Hydrogels are polymeric materials that are capable of absorbing relatively large quantities of water. Examples of hydrogel forming compounds include, but are not limited to, polyacrylic acids, sodium carboxymethylcellulose, polyvinyl alcohol, polyvinyl pyrrolidine, gelatin, carrageenan and other polysaccharides, hydroxyethylenemethacrylic acid (HEMA), as well as derivatives thereof, and the like. Hydrogels can be produced that are stable, biodegradable and bioresorbable. Moreover, hydrogel compositions can include subunits that exhibit one or more of these properties.

[0289] Bio-compatible hydrogel compositions whose integrity can be controlled through crosslinking are known and are presently preferred for use in the methods of the invention. For example, Hubbell *et al.*, U.S. Patent Nos. 5,410,016, which issued on April 25, 1995 and 5,529,914, which issued on June 25, 1996, disclose water-soluble systems, which are crosslinked block copolymers having a water-soluble central block segment sandwiched between two hydrolytically labile extensions. Such copolymers are further end-capped with photopolymerizable acrylate functionalities. When crosslinked, these systems become hydrogels. The water soluble central block of such copolymers can include poly(ethylene glycol); whereas, the hydrolytically labile extensions can be a poly(α-hydroxy acid), such as polyglycolic acid or polylactic acid. *See*, Sawhney *et al.*, *Macromolecules* **26**: 581-587 (1993).

[0290] In another preferred embodiment, the gel is a thermoreversible gel. Thermoreversible gels including components, such as pluronics, collagen, gelatin, hyalouronic acid, polysaccharides, polyurethane hydrogel, polyurethane-urea hydrogel and combinations thereof are presently preferred.

[0291] In yet another exemplary embodiment, the conjugate of the invention includes a component of a liposome. Liposomes can be prepared according to methods known to those skilled in the art, for example, as described in Eppstein *et al.*, U.S. Patent No. 4,522,811. For example, liposome formulations may be prepared by dissolving appropriate lipid(s) (such as stearoyl phosphatidyl ethanolamine, stearoyl phosphatidyl choline, arachadoyl phosphatidyl

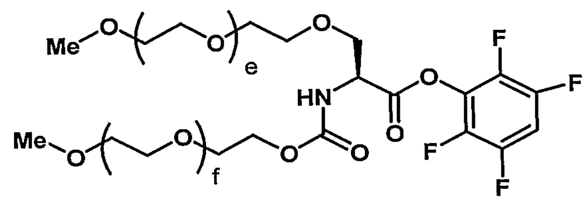
choline, and cholesterol) in an inorganic solvent that is then evaporated, leaving behind a thin film of dried lipid on the surface of the container. An aqueous solution of the active compound or its pharmaceutically acceptable salt is then introduced into the container. The container is then swirled by hand to free lipid material from the sides of the container and to disperse lipid aggregates, thereby forming the liposomal suspension.

[0292] The above-recited microparticles and methods of preparing the microparticles are offered by way of example and they are not intended to define the scope of microparticles of use in the present invention. It will be apparent to those of skill in the art that an array of microparticles, fabricated by different methods, is of use in the present invention.

[0293] The structural formats discussed above in the context of the water-soluble polymers, both straight-chain and branched are generally applicable with respect to the water-insoluble polymers as well. Thus, for example, the cysteine, serine, dilysine, and trilysine branching cores can be functionalized with two water-insoluble polymer moieties. The methods used to produce these species are generally closely analogous to those used to produce the water-soluble polymers.

II. D. v. <u>*Methods of Producing the Polymeric Modifying Groups*</u>

[0294] The polymeric modifying groups can be activated for reaction with a glycosyl or saccharyl moiety or an amino acid moiety. Exemplary structures of activated species (e.g., carbonates and active esters) include: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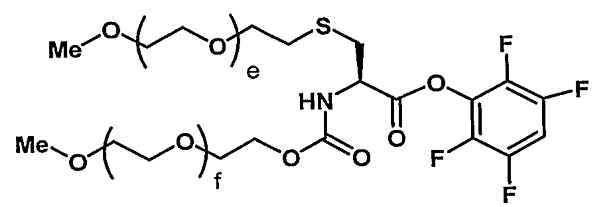
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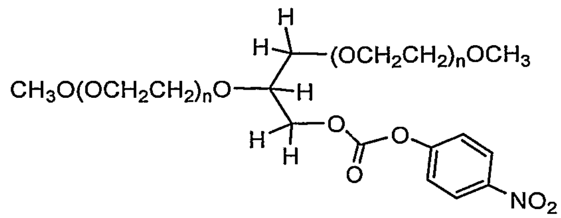

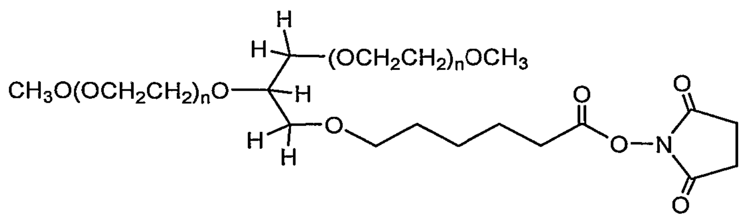

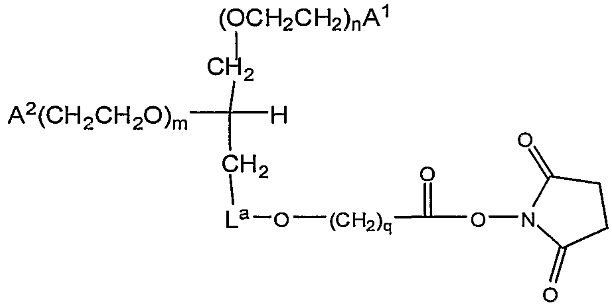

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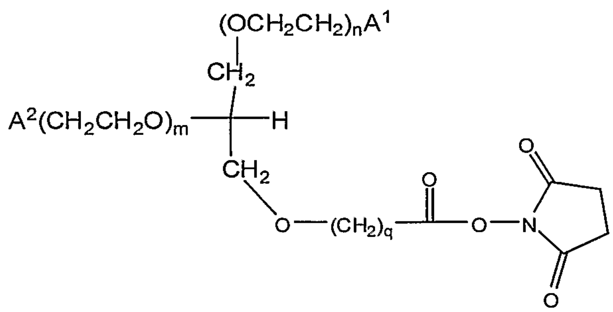

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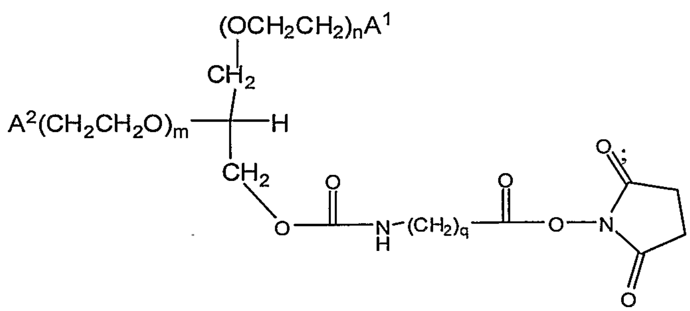

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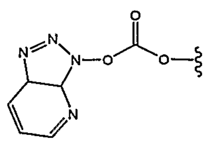
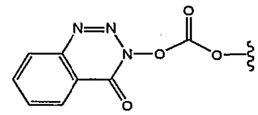
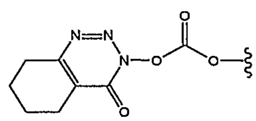
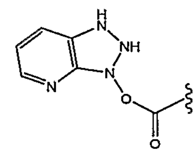
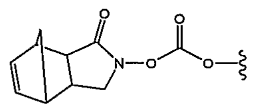
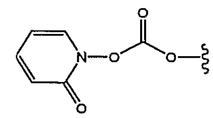
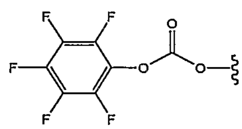
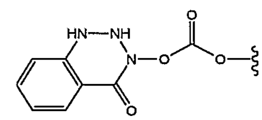
[0295] In the figure above, q is a member selected from 1-40. Other activating, or leaving groups, appropriate for activating linear and branched PEGs of use in preparing the compounds set forth herein include, but are not limited to the species:

 ;  ;

 ;  ;

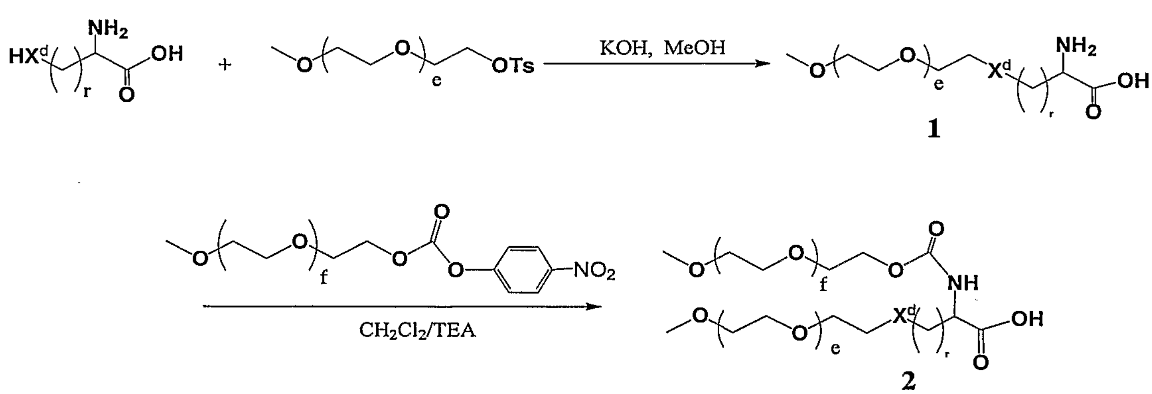
 ;  ;

 ; and  .

PEG molecules that are activated with these and other species and methods of making the activated PEGs are set forth in WO 04/083259.

[0296] Those of skill in the art will appreciate that one or more of the m-PEG arms of the branched polymers shown above can be replaced by a PEG moiety with a different terminus, e.g., OH, COOH, NH_2, C_2-C_{10}-alkyl, etc. Moreover, the structures above are readily modified by inserting alkyl linkers (or removing carbon atoms) between the α-carbon atom and the functional group of the amino acid side chain. Thus, "homo" derivatives and higher homologues, as well as lower homologues are within the scope of cores for branched PEGs of use in the present invention.

[0297] The branched PEG species set forth herein are readily prepared by methods such as that set forth in the scheme below:



in which X^d is O or S and r is an integer from 1 to 5. The indices e and f are independently selected integers from 1 to 2500. In an exemplary embodiment, one or both of these indices

are selected such that the polymer is about 5 KDa, 10 KDa, 15 KDa, 20 KDa, 25 KDa, 30 KDa, 35 KDa, or 40KDa in molecular weight.

[0298] Thus, according to this scheme, a natural or unnatural amino acid is contacted with an activated m-PEG derivative, in this case the tosylate, forming **1** by alkylating the side-chain heteroatom X^d. The mono-functionalize m-PEG amino acid is submitted to N-acylation conditions with a reactive m-PEG derivative, thereby assembling branched m-PEG **2**. As one of skill will appreciate, the tosylate leaving group can be replaced with any suitable leaving group, e.g., halogen, mesylate, triflate, etc. Similarly, the reactive carbonate utilized to acylate the amine can be replaced with an active ester, e.g., N-hydroxysuccinimide, etc., or the acid can be activated *in situ* using a dehydrating agent such as dicyclohexylcarbodiimide, carbonyldiimidazole, etc.

[0299] In other exemplary embodiments, the urea moiety is replaced by a group such as a amide.

II. E. ***<u>Homodisperse Peptide Conjugate Compositions of Matter</u>***

[0300] In addition to providing peptide conjugates that are formed through a chemically or enzymatically added glycosyl linking group, the present invention provides compositions of matter comprising peptide conjugates that are highly homogenous in their substitution patterns. Using the methods of the invention, it is possible to form peptide conjugates in which substantial proportion of the glycosyl linking groups and glycosyl moieties across a population of Factor VII/Factor VIIa conjugates are attached to a structurally identical amino acid or glycosyl residue. Thus, in a second aspect, the invention provides a peptide conjugate having a population of water-soluble polymer moieties, which are covalently bound to the peptide through a glycosyl linking group, e.g., an intact glycosyl linking group. In a an exemplary peptide conjugate of the invention, essentially each member of the water soluble polymer population is bound via the glycosyl linking group to a glycosyl residue of the peptide, and each glycosyl residue of the peptide to which the glycosyl linking group is attached has the same structure.

[0301] The present invention also provides conjugates analogous to those described above in which the peptide is conjugated to a modifying group, e.g. therapeutic moiety, diagnostic moiety, targeting moiety, toxin moiety or the like via a glycosyl linking group. Each of the above-recited modifying groups can be a small molecule, natural polymer (e.g.,

polypeptide) or synthetic polymer. When the modifying group is attached to a sialic acid, it is generally preferred that the modifying group is substantially non-fluorescent.

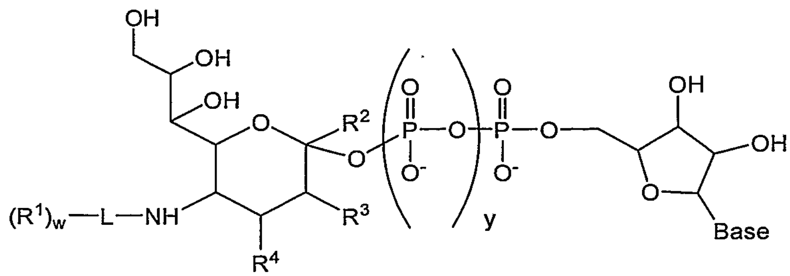
[0302] In an exemplary embodiment, the peptides of the invention include at least one O-linked or N-linked glycosylation site, which is glycosylated with a modified sugar that includes a polymeric modifying group, e.g., a PEG moiety. In an exemplary embodiment, the PEG is covalently attached to the peptide via an intact glycosyl linking group, or via a non-glycosyl linker, e.g., substituted or unsubstituted alkyl, substituted or unsubstituted heteroalkyl. The glycosyl linking group is covalently attached to either an amino acid residue or a glycosyl residue of the peptide. Alternatively, the glycosyl linking group is attached to one or more glycosyl units of a glycopeptide. The invention also provides conjugates in which a glycosyl linking group is attached to both an amino acid residue and a glycosyl residue.

[0303] The glycans on the peptides of the invention generally correspond to those found on a Factor VII/Factor VIIa peptide that is produced by mammalian (BHK, CHO) cells or insect (e.g., Sf-9) cells, following remodeling according to the methods set forth herein. For example insect-derived Factor VII/Factor VIIa peptide that is expressed with a tri-mannosyl core is subsequently contacted with a GlcNAc donor and a GlcNAc transferase and a Gal donor and a Gal transferase. Appending GlcNAc and Gal to the tri-mannosyl core is accomplished in either two steps or a single step. A modified sialic acid is added to at least one branch of the glycosyl moiety as discussed herein. Those Gal moieties that are not functionalized with the modified sialic acid are optionally "capped" by reaction with a sialic acid donor in the presence of a sialyl transferase.

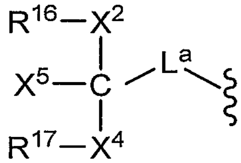
[0304] In an exemplary embodiment, at least 60% of terminal Gal moieties in a population of peptides is capped with sialic acid, preferably at least 70%, more preferably, at least 80%, still more preferably at least 90% and even more preferably at least 95%, 96%, 97%, 98% or 99% are capped with sialic acid.

II. F. ***Nucleotide Sugars***

[0305] In another aspect of the invention, the invention also provides sugar nucleotides. Exemplary species according to this embodiment include:



in which the index y is an integer selected from 0, 1 and 2. Base is a nucleic acid base, such as adenine, thymine, guanine, cytidine and uridine. R^2, R^3 and R^4 are as described above. In an exemplary embodiment, L-$(R^1)_w$ is a member selected from



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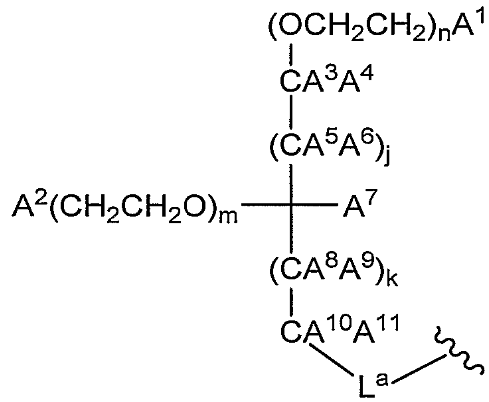
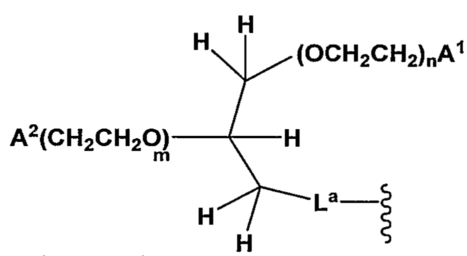


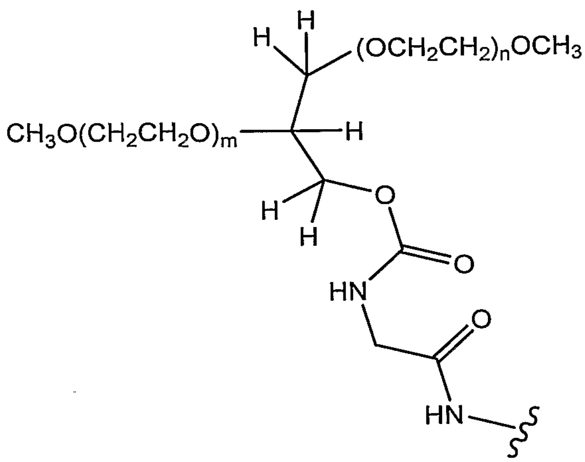
in which the variables are as described above.

[0306] In an exemplary embodiment, L-$(R^1)_w$ has a structure according to the following formula:



In an exemplary embodiment, A^1 and A^2 are each selected from -OH and $-OCH_3$.

[0307] Exemplary polymeric modifying groups according to this embodiment include:



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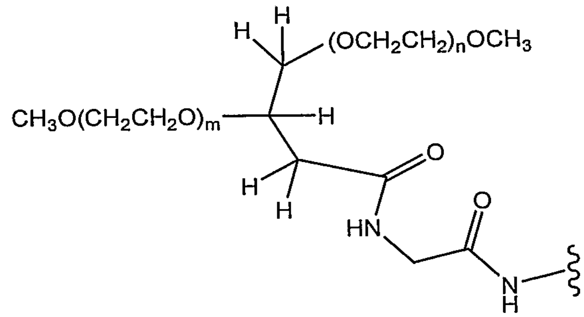
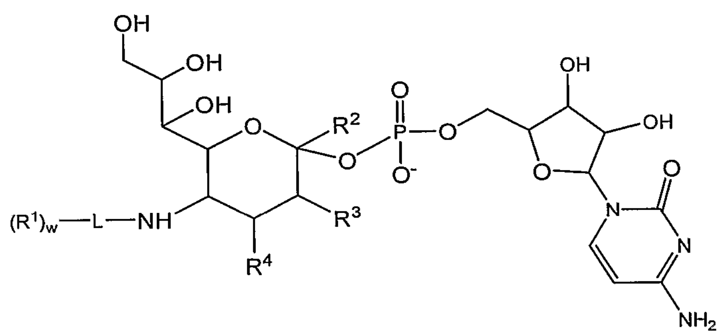


[0308] In another exemplary embodiment, the nucleotide sugars have a formula which is a member selected from:



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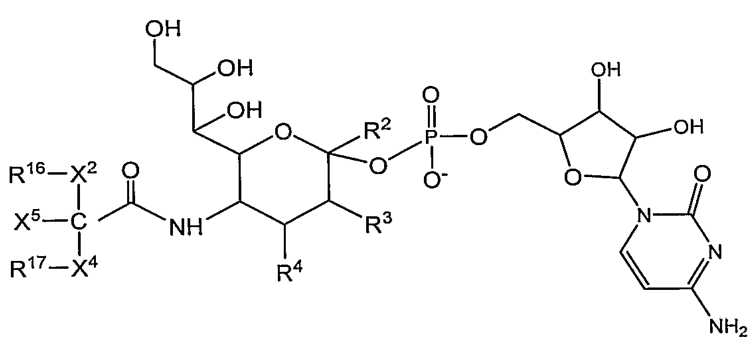

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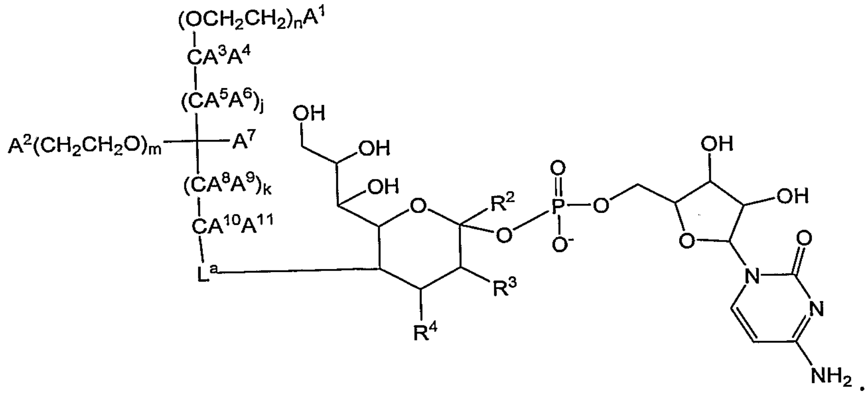
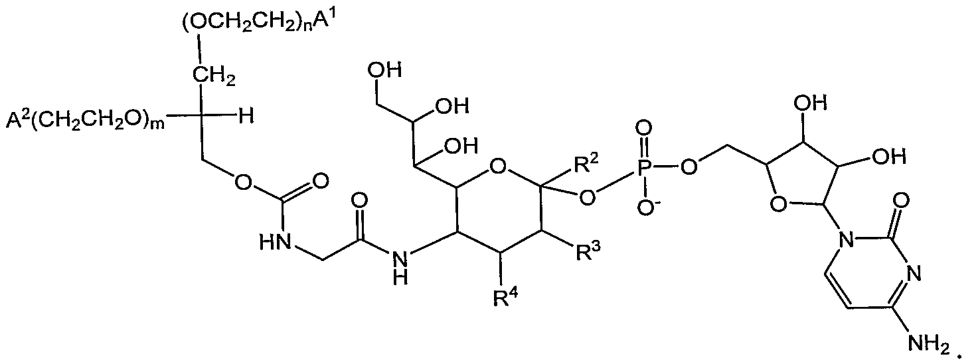
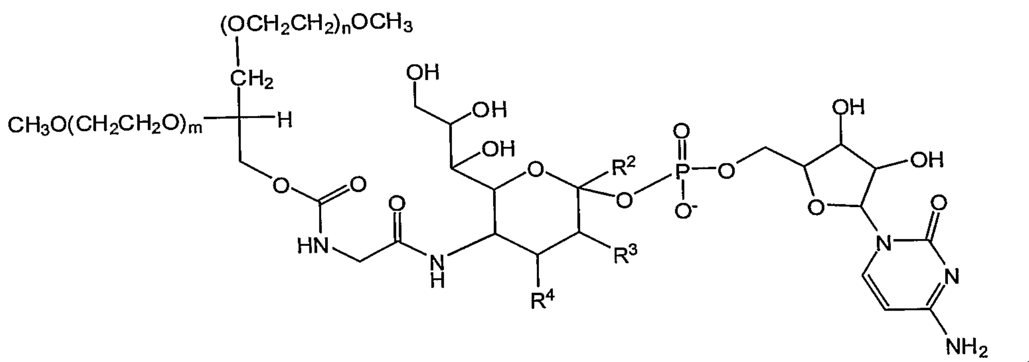


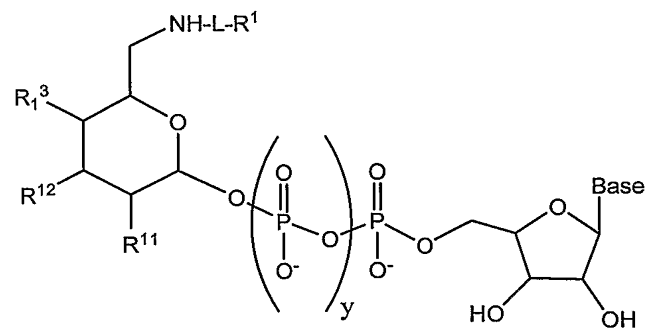
[0309] An exemplary nucleotide sugar according to this embodiment has the structure:



[0310] An exemplary nucleotide sugar according to this embodiment has the structure:

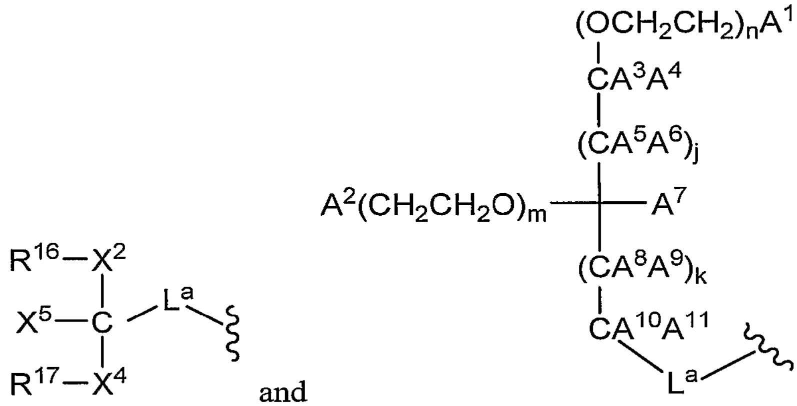


[0311] In another exemplary embodiment, the nucleotide sugar is based upon the following formula:



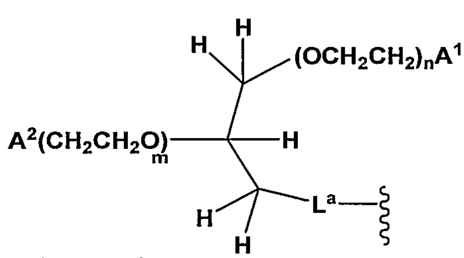
in which the R groups, and L, represent moieties as discussed above. The index "y" is 0, 1 or 2. In an exemplary embodiment, L is a bond between NH and R^1. The base is a nucleic acid base.

[0312] In an exemplary embodiment, L-R^1 is a member selected from



in which the variables are as described above.

[0313] In an exemplary embodiment, L-R^1 has a structure according to the following formula:



In an exemplary embodiment, A^1 and A^2 are each slected from -OH and $-OCH_3$.

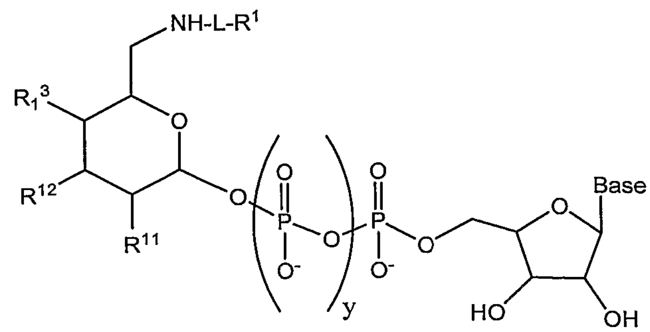
III. *__The Methods__*

[0314] In addition to the conjugates discussed above, the present invention provides methods for preparing these and other conjugates. Moreover, the invention provides methods of preventing, curing or ameliorating a disease state by administering a conjugate of the invention to a subject at risk of developing the disease or a subject that has the disease.

[0315] In exemplary embodiments, the conjugate is formed between a polymeric modifying moiety and a glycosylated or non-glycosylated peptide. The polymer is conjugated to the peptide via a glycosyl linking group, which is interposed between, and covalently linked to both the peptide (or glycosyl residue) and the modifying group (e.g., water-soluble polymer). The method includes contacting the peptide with a mixture containing a modified sugar and an enzyme, e.g., a glycosyltransferase that conjugates the modified sugar to the substrate. The reaction is conducted under conditions appropriate to form a covalent bond between the modified sugar and the peptide. The sugar moiety of the modified sugar is preferably selected from nucleotide sugars. The method of synthesizing a Factor VII/Factor VIIa peptide conjugate, comprising combining a) sialidase; b) an enzyme capable of catalyzing the transfer of a glycosyl linking group such as a glycosyltransferase, exoglycosidase or endoglycosidase; c) modified sugar; d) Factor VII/Factor VIIa peptide, thus synthesizing the Factor VII/Factor VIIa peptide conjugate. The reaction is conducted under conditions appropriate to form a covalent bond between the modified sugar and the peptide. The sugar moiety of the modified sugar is preferably selected from nucleotide sugars.

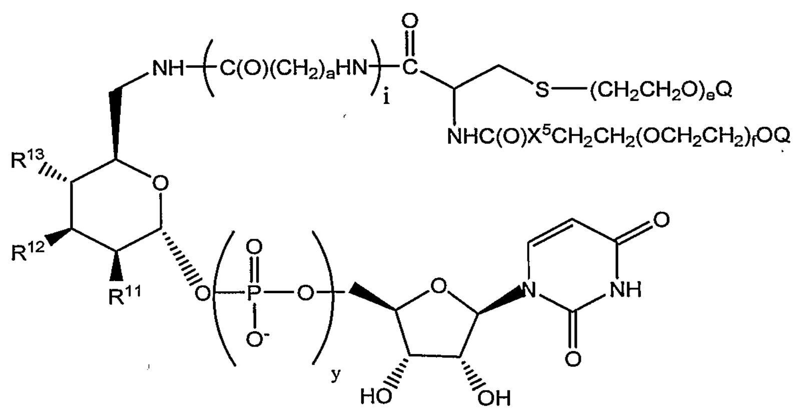
[0316] In an exemplary embodiment, the modified sugar, such as those set forth above, is activated as the corresponding nucleotide sugars. Exemplary sugar nucleotides that are used in the present invention in their modified form include nucleotide mono-, di- or triphosphates or analogs thereof. In a preferred embodiment, the modified sugar nucleotide is selected from a UDP-glycoside, CMP-glycoside, or a GDP-glycoside. Even more preferably, the sugar nucleotide portion of the modified sugar nucleotide is selected from UDP-galactose, UDP-galactosamine, UDP-glucose, UDP-glucosamine, GDP-mannose, GDP-fucose, CMP-sialic acid, or CMP-NeuAc. In an exemplary embodiment, the nucleotide phosphate is attached to C-1.

[0317] The invention also provides for the use of sugar nucleotides modified with L-R^1 at the 6-carbon position. Exemplary species according to this embodiment include:

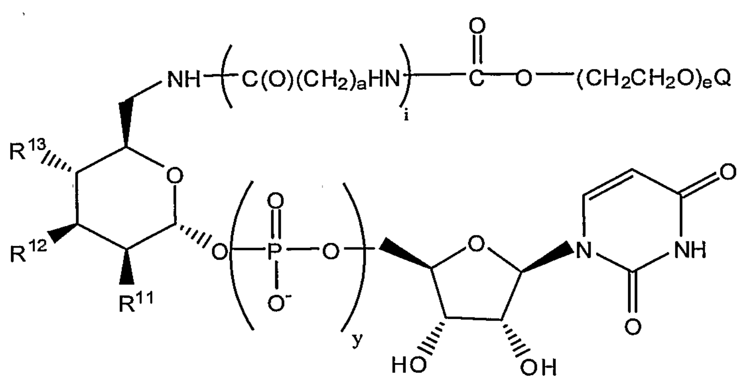


in which the R groups, and L, represent moieties as discussed above. The index "y" is 0, 1 or 2. In an exemplary embodiment, L is a bond between NH and R^1. The base is a nucleic acid base.

[0318] Exemplary nucleotide sugars of use in the invention in which the carbon at the 6-position is modified include species having the stereochemistry of GDP mannose, e.g.:

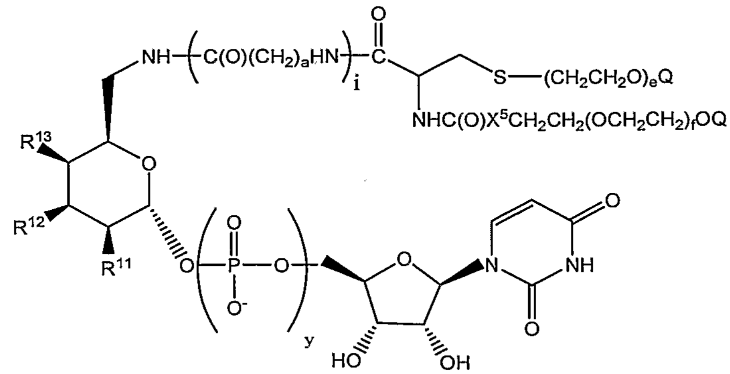


; and

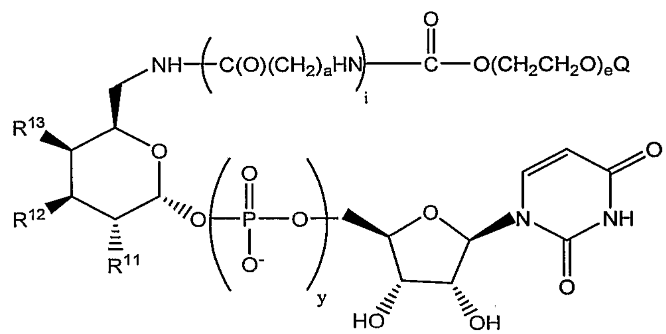


in which X^5 is a bond or O. The index i represents 0 or 1. The index a represents an integer from 1 to 20. The indices e and f independently represent integers from 1 to 2500. Q, as discussed above, is H or substituted or unsubstituted C_1-C_6 alkyl. As those of skill will appreciate, the serine derivative, in which S is replaced with O also falls within this general motif.

[0319] In a still further exemplary embodiment, the invention provides a conjugate in which the modified sugar is based on the stereochemistry of UDP galactose. An exemplary nucleotide sugar of use in this invention has the structure:

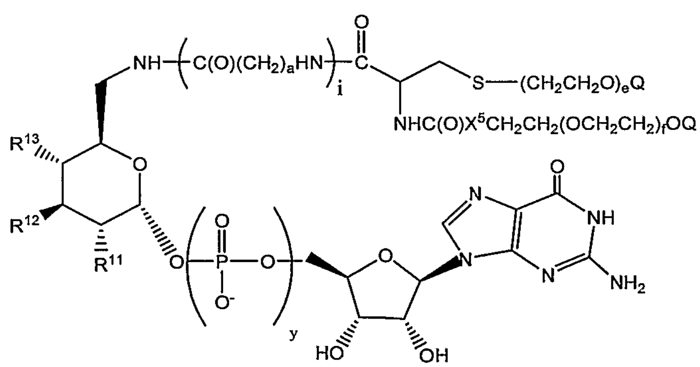


; and

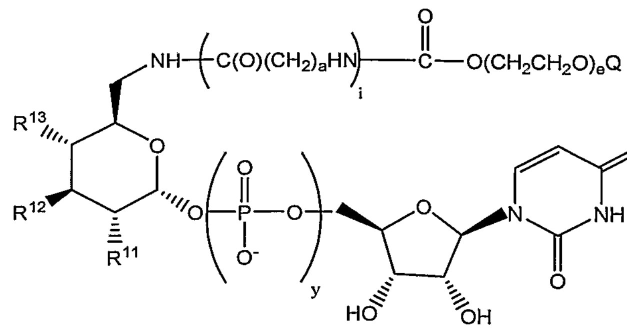


.

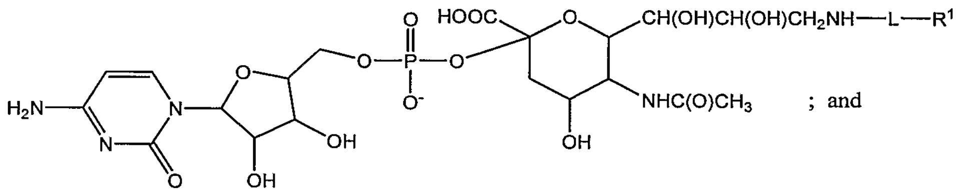
[0320] In another exemplary embodiment, the nucleotide sugar is based on the stereochemistry of glucose. Exemplary species according to this embodiment have the formulae:



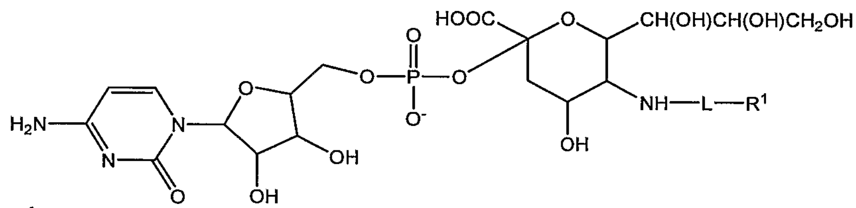
; and



[0321] Thus, in an illustrative embodiment in which the glycosyl moiety is sialic acid, the method of the invention utilizes compounds having the formulae:

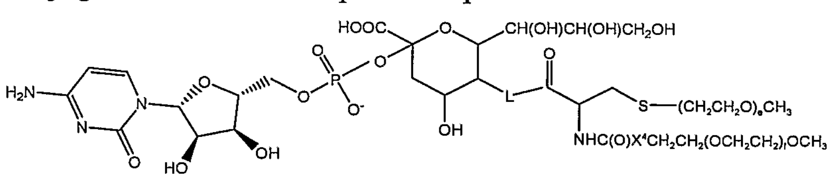


; and

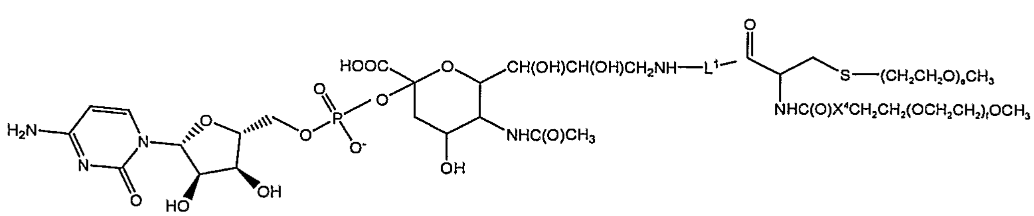


in which L-R^1 is as discussed above, and L^1-R^1 represents a linker bound to the modifying group. As with L, exemplary linker species according to L^1 include a bond, alkyl or heteroalkyl moieties.

[0322] Moreover, as discussed above, the present invention provides for the use of nucleotide sugars that are modified with a water-soluble polymer, which is either straight-chain or branched. For example, compounds having the formula shown below are of use to prepare conjugates within the scope of the present invention:



; and



in which X^4 is O or a bond.

[0323] In general, the sugar moiety or sugar moiety-linker cassette and the PEG or PEG-linker cassette groups are linked together through the use of reactive groups, which are typically transformed by the linking process into a new organic functional group or unreactive species. The sugar reactive functional group(s), is located at any position on the sugar moiety. Reactive groups and classes of reactions useful in practicing the present invention are generally those that are well known in the art of bioconjugate chemistry. Currently favored classes of reactions available with reactive sugar moieties are those, which proceed under relatively mild conditions. These include, but are not limited to nucleophilic substitutions (*e.g.*, reactions of amines and alcohols with acyl halides, active esters), electrophilic substitutions (*e.g.*, enamine reactions) and additions to carbon-carbon and carbon-heteroatom multiple bonds (*e.g.*, Michael reaction, Diels-Alder addition). These and other useful reactions are discussed in, for example, March, ADVANCED ORGANIC CHEMISTRY, 3rd Ed., John Wiley & Sons, New York, 1985; Hermanson, BIOCONJUGATE TECHNIQUES, Academic Press, San Diego, 1996; and Feeney *et al.*, MODIFICATION OF PROTEINS; Advances in Chemistry Series, Vol. 198, American Chemical Society, Washington, D.C., 1982.

[0324] Useful reactive functional groups pendent from a sugar nucleus or modifying group include, but are not limited to:

(a) carboxyl groups and various derivatives thereof including, but not limited to, N-hydroxysuccinimide esters, N-hydroxybenztriazole esters, acid halides, acyl imidazoles, thioesters, p-nitrophenyl esters, alkyl, alkenyl, alkynyl and aromatic esters;

(b) hydroxyl groups, which can be converted to, *e.g.*, esters, ethers, aldehydes, *etc.*

(c) haloalkyl groups, wherein the halide can be later displaced with a nucleophilic group such as, for example, an amine, a carboxylate anion, thiol anion, carbanion, or an alkoxide ion, thereby resulting in the covalent attachment of a new group at the functional group of the halogen atom;

(d) dienophile groups, which are capable of participating in Diels-Alder reactions such as, for example, maleimido groups;

(e) aldehyde or ketone groups, such that subsequent derivatization is possible via formation of carbonyl derivatives such as, for example, imines, hydrazones, semicarbazones or oximes, or via such mechanisms as Grignard addition or alkyllithium addition;

(f) sulfonyl halide groups for subsequent reaction with amines, for example, to form sulfonamides;

(g) thiol groups, which can be, for example, converted to disulfides or reacted with acyl halides;

(h) amine or sulfhydryl groups, which can be, for example, acylated, alkylated or oxidized;

(i) alkenes, which can undergo, for example, cycloadditions, acylation, Michael addition, *etc*; and

(j) epoxides, which can react with, for example, amines and hydroxyl compounds.

[0325] The reactive functional groups can be chosen such that they do not participate in, or interfere with, the reactions necessary to assemble the reactive sugar nucleus or modifying group. Alternatively, a reactive functional group can be protected from participating in the reaction by the presence of a protecting group. Those of skill in the art understand how to protect a particular functional group such that it does not interfere with a chosen set of reaction conditions. For examples of useful protecting groups, *see,* for example, Greene *et al.*, PROTECTIVE GROUPS IN ORGANIC SYNTHESIS, John Wiley & Sons, New York, 1991.

[0326] In the discussion that follows, a number of specific examples of modified sugars that are useful in practicing the present invention are set forth. In the exemplary embodiments, a sialic acid derivative is utilized as the sugar nucleus to which the modifying group is attached. The focus of the discussion on sialic acid derivatives is for clarity of illustration only and should not be construed to limit the scope of the invention. Those of skill in the art will appreciate that a variety of other sugar moieties can be activated and derivatized in a manner analogous to that set forth using sialic acid as an example. For example, numerous methods are available for modifying galactose, glucose, N-acetylgalactosamine and fucose to name a few sugar substrates, which are readily modified by art recognized methods. *See*, for example, Elhalabi *et al.*, *Curr. Med. Chem.* **6**: 93 (1999); and Schafer *et al.*, *J. Org. Chem.* **65**: 24 (2000)).

[0327] In an exemplary embodiment, the modified sugar is based upon a 6-amino-N-acetyl-glycosyl moiety.

[0328] In the scheme above, the index n represents an integer from 1 to 2500. In an exemplary embodiment, this index is selected such that the polymer is about 10 KDa, 15 KDa or 20 KDa in molecular weight. The symbol "A" represents an activating group, e.g., a halo,

a component of an activated ester (e.g., a N-hydroxysuccinimide ester), a component of a carbonate (e.g., p-nitrophenyl carbonate) and the like. Those of skill in the art will appreciate that other PEG-amide nucleotide sugars are readily prepared by this and analogous methods.

[0329] The peptide is typically synthesized *de novo*, or recombinantly expressed in a prokaryotic cell (*e.g.*, bacterial cell, such as *E. coli*) or in a eukaryotic cell such as a mammalian, yeast, insect, fungal or plant cell. The peptide can be either a full-length protein or a fragment. Moreover, the peptide can be a wild type or mutated peptide. In an exemplary embodiment, the peptide includes a mutation that adds one or more N- or O-linked glycosylation sites to the peptide sequence.

[0330] The method of the invention also provides for modification of incompletely glycosylated peptides that are produced recombinantly. Many recombinantly produced glycoproteins are incompletely glycosylated, exposing carbohydrate residues that may have undesirable properties, e.g., immunogenicity, recognition by the RES. Employing a modified sugar in a method of the invention, the peptide can be simultaneously further glycosylated and derivatized with, e.g., a water-soluble polymer, therapeutic agent, or the like. The sugar moiety of the modified sugar can be the residue that would properly be conjugated to the acceptor in a fully glycosylated peptide, or another sugar moiety with desirable properties.

[0331] Those of skill will appreciate that the invention can be practiced using substantially any peptide or glycopeptide from any source. Exemplary peptides with which the invention can be practiced are set forth in WO 03/031464, and the references set forth therein.

[0332] Peptides modified by the methods of the invention can be synthetic or wild-type peptides or they can be mutated peptides, produced by methods known in the art, such as site-directed mutagenesis. Glycosylation of peptides is typically either N-linked or O-linked. An exemplary N-linkage is the attachment of the modified sugar to the side chain of an asparagine residue. The tripeptide sequences asparagine-X-serine and asparagine-X-threonine, where X is any amino acid except proline, are the recognition sequences for enzymatic attachment of a carbohydrate moiety to the asparagine side chain. Thus, the presence of either of these tripeptide sequences in a polypeptide creates a potential glycosylation site. O-linked glycosylation refers to the attachment of one sugar (*e.g.*, N-acetylgalactosamine, galactose, mannose, GlcNAc, glucose, fucose or xylose) to the hydroxy side chain of a hydroxyamino acid, preferably serine or threonine, although unusual or non-natural amino acids, e.g., 5-hydroxyproline or 5-hydroxylysine may also be used.

[0333] Moreover, in addition to peptides, the methods of the present invention can be practiced with other biological structures (*e.g.*, glycolipids, lipids, sphingoids, ceramides, whole cells, and the like, containing a glycosylation site).

[0334] Addition of glycosylation sites to a peptide or other structure is conveniently accomplished by altering the amino acid sequence such that it contains one or more glycosylation sites. The addition may also be made by the incorporation of one or more species presenting an –OH group, preferably serine or threonine residues, within the sequence of the peptide (for O-linked glycosylation sites). The addition may be made by mutation or by full chemical synthesis of the peptide. The peptide amino acid sequence is preferably altered through changes at the DNA level, particularly by mutating the DNA encoding the peptide at preselected bases such that codons are generated that will translate into the desired amino acids. The DNA mutation(s) are preferably made using methods known in the art.

[0335] In an exemplary embodiment, the glycosylation site is added by shuffling polynucleotides. Polynucleotides encoding a candidate peptide can be modulated with DNA shuffling protocols. DNA shuffling is a process of recursive recombination and mutation, performed by random fragmentation of a pool of related genes, followed by reassembly of the fragments by a polymerase chain reaction-like process. *See*, *e.g.*, Stemmer, *Proc. Natl. Acad. Sci. USA* 91:10747-10751 (1994); Stemmer, *Nature* 370:389-391 (1994); and U.S. Patent Nos. 5,605,793, 5,837,458, 5,830,721 and 5,811,238.

[0336] Exemplary peptides with which the present invention can be practiced, methods of adding or removing glycosylation sites, and adding or removing glycosyl structures or substructures are described in detail in WO03/031464 and related U.S. and PCT applications.

[0337] The present invention also takes advantage of adding to (or removing from) a peptide one or more selected glycosyl residues, after which a modified sugar is conjugated to at least one of the selected glycosyl residues of the peptide. The present embodiment is useful, for example, when it is desired to conjugate the modified sugar to a selected glycosyl residue that is either not present on a peptide or is not present in a desired amount. Thus, prior to coupling a modified sugar to a peptide, the selected glycosyl residue is conjugated to the peptide by enzymatic or chemical coupling. In another embodiment, the glycosylation pattern of a glycopeptide is altered prior to the conjugation of the modified sugar by the removal of a carbohydrate residue from the glycopeptide. *See*, for example WO 98/31826.

[0338] Addition or removal of any carbohydrate moieties present on the glycopeptide is accomplished either chemically or enzymatically. An exemplary chemical deglycosylation is brought about by exposure of the polypeptide variant to the compound trifluoromethanesulfonic acid, or an equivalent compound. This treatment results in the cleavage of most or all sugars except the linking sugar (N-acetylglucosamine or N-acetylgalactosamine), while leaving the peptide intact. Chemical deglycosylation is described by Hakimuddin *et al., Arch. Biochem. Biophys.* **259**: 52 (1987) and by Edge *et al., Anal. Biochem.* **118**: 131 (1981). Enzymatic cleavage of carbohydrate moieties on polypeptide variants can be achieved by the use of a variety of endo- and exo-glycosidases as described by Thotakura *et al., Meth. Enzymol.* **138**: 350 (1987).

[0339] In an exemplary embodiment, the peptide is essentially completely desialylated with neuraminidase prior to performing glycoconjugation or remodeling steps on the peptide. Following the glycoconjugation or remodeling, the peptide is optionally re-sialylated using a sialyltransferase. In an exemplary embodiment, the re-sialylation occurs at essentially each (e.g., >80%, preferably greater than 85%, greater than 90%, preferably greater than 95% and more preferably greater than 96%, 97%, 98% or 99%) terminal saccharyl acceptor in a population of sialyl acceptors. In a preferred embodiment, the saccharide has a substantially uniform sialylation pattern (i.e., substantially uniform glycosylation pattern).

[0340] Chemical addition of glycosyl moieties is carried out by any art-recognized method. Enzymatic addition of sugar moieties is preferably achieved using a modification of the methods set forth herein, substituting native glycosyl units for the modified sugars used in the invention. Other methods of adding sugar moieties are disclosed in U.S. Patent No. 5,876,980, 6,030,815, 5,728,554, and 5,922,577.

[0341] Exemplary attachment points for selected glycosyl residue include, but are not limited to: (a) consensus sites for N-linked glycosylation, and sites for O-linked glycosylation; (b) terminal glycosyl moieties that are acceptors for a glycosyltransferase; (c) arginine, asparagine and histidine; (d) free carboxyl groups; (e) free sulfhydryl groups such as those of cysteine; (f) free hydroxyl groups such as those of serine, threonine, or hydroxyproline; (g) aromatic residues such as those of phenylalanine, tyrosine, or tryptophan; or (h) the amide group of glutamine. Exemplary methods of use in the present invention are described in WO 87/05330 published Sep. 11, 1987, and in Aplin and Wriston, CRC CRIT. REV. BIOCHEM., pp. 259-306 (1981).

[0342] In one embodiment, the invention provides a method for linking two or more peptides through a linking group. The linking group is of any useful structure and may be selected from straight- and branched-chain structures. Preferably, each terminus of the linker, which is attached to a peptide, includes a modified sugar (i.e., a nascent intact glycosyl linking group).

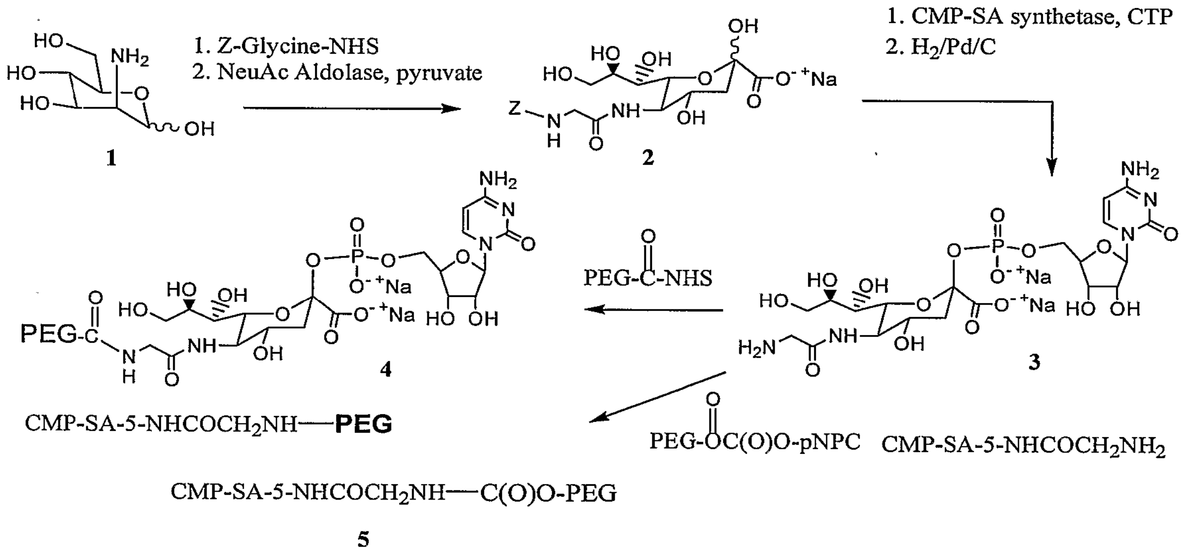
[0343] In an exemplary method of the invention, two peptides are linked together via a linker moiety that includes a polymeric (e.g., PEG linker). The construct conforms to the general structure set forth in the cartoon above. As described herein, the construct of the invention includes two intact glycosyl linking groups (i.e., $s + t = 1$). The focus on a PEG linker that includes two glycosyl groups is for purposes of clarity and should not be interpreted as limiting the identity of linker arms of use in this embodiment of the invention.

[0344] Thus, a PEG moiety is functionalized at a first terminus with a first glycosyl unit and at a second terminus with a second glycosyl unit. The first and second glycosyl units are preferably substrates for different transferases, allowing orthogonal attachment of the first and second peptides to the first and second glycosyl units, respectively. In practice, the $(\text{glycosyl})^1$-PEG-$(\text{glycosyl})^2$ linker is contacted with the first peptide and a first transferase for which the first glycosyl unit is a substrate, thereby forming $(\text{peptide})^1$-$(\text{glycosyl})^1$-PEG-$(\text{glycosyl})^2$. Transferase and/or unreacted peptide is then optionally removed from the reaction mixture. The second peptide and a second transferase for which the second glycosyl unit is a substrate are added to the $(\text{peptide})^1$-$(\text{glycosyl})^1$-PEG-$(\text{glycosyl})^2$ conjugate, forming $(\text{peptide})^1$-$(\text{glycosyl})^1$-PEG-$(\text{glycosyl})^2$-$(\text{peptide})^2$; at least one of the glycosyl residues is either directly or indirectly O-linked. Those of skill in the art will appreciate that the method outlined above is also applicable to forming conjugates between more than two peptides by, for example, the use of a branched PEG, dendrimer, poly(amino acid), polysaccharide or the like.

[0345] In an exemplary embodiment, the peptide that is modified by a method of the invention is a glycopeptide that is produced in mammalian cells (e.g., CHO cells) or in a transgenic animal and thus, contains N- and/or O-linked oligosaccharide chains, which are incompletely sialylated. The oligosaccharide chains of the glycopeptide lacking a sialic acid and containing a terminal galactose residue can be PEGylated, PPGylated or otherwise modified with a modified sialic acid.

[0346] In Scheme 1, the amino glycoside **1**, is treated with the active ester of a protected amino acid (*e.g.*, glycine) derivative, converting the sugar amine residue into the corresponding protected amino acid amide adduct. The adduct is treated with an aldolase to form α-hydroxy carboxylate **2**. Compound **2** is converted to the corresponding CMP derivative by the action of CMP-SA synthetase, followed by catalytic hydrogenation of the CMP derivative to produce compound **3**. The amine introduced via formation of the glycine adduct is utilized as a locus of PEG attachment by reacting compound **3** with an activated PEG or PPG derivative (*e.g.*, PEG-C(O)NHS, PEG-OC(O)O-p-nitrophenyl), producing species such as **4** or **5**, respectively.

Scheme 1



In an exemplary embodiment, a modified sugar can be attached to an O-glycan binding site on a Factor VII/Factor VIIa peptide. The glycosyltransferases which can be used to produce this Factor VII/Factor VIIa peptide conjugate include: for Ser56 (-Glc-(Xyl)n-Gal-SA-PEG – a galactosyltransferase and sialyltransferase; for Ser56 –Glc-(Xyl)n-Xyl-PEG – a xylosyltransferase; and for Ser60-Fuc-GlcNAc-(Gal)n-(SA)m-PEG – a GlcNAc transferase.

III. A. <u>Conjugation of Modified Sugars to Peptides</u>

[0347] The PEG modified sugars are conjugated to a glycosylated or non-glycosylated peptide using an appropriate enzyme to mediate the conjugation. Preferably, the concentrations of the modified donor sugar(s), enzyme(s) and acceptor peptide(s) are selected such that glycosylation proceeds until the acceptor is consumed. The considerations discussed below, while set forth in the context of a sialyltransferase, are generally applicable

to other glycosyltransferase reactions. A list of preferred sialyltransferases for use in the invention is provided in **FIG. 3**.

[0348] A number of methods of using glycosyltransferases to synthesize desired oligosaccharide structures are known and are generally applicable to the instant invention. Exemplary methods are described, for instance, WO 96/32491, Ito *et al., Pure Appl. Chem.* **65**: 753 (1993), U.S. Pat. Nos. 5,352,670, 5,374,541, 5,545,553, commonly owned U.S. Pat. Nos. 6,399,336, and 6,440,703, and commonly owned published PCT applications, WO 03/031464, WO 04/033651, WO 04/099231, which are incorporated herein by reference.

[0349] The present invention is practiced using a single glycosyltransferase or a combination of glycosyltransferases. For example, one can use a combination of a sialyltransferase and a galactosyltransferase. In those embodiments using more than one enzyme, the enzymes and substrates are preferably combined in an initial reaction mixture, or the enzymes and reagents for a second enzymatic reaction are added to the reaction medium once the first enzymatic reaction is complete or nearly complete. By conducting two enzymatic reactions in sequence in a single vessel, overall yields are improved over procedures in which an intermediate species is isolated. Moreover, cleanup and disposal of extra solvents and by-products is reduced.

[0350] In a preferred embodiment, each of the first and second enzyme is a glycosyltransferase. In another preferred embodiment, one enzyme is an endoglycosidase. In an additional preferred embodiment, more than two enzymes are used to assemble the modified glycoprotein of the invention. The enzymes are used to alter a saccharide structure on the peptide at any point either before or after the addition of the modified sugar to the peptide.

[0351] In another embodiment, the method makes use of one or more exo- or endoglycosidase. The glycosidase is typically a mutant, which is engineered to form glycosyl bonds rather than rupture them. The mutant glycanase typically includes a substitution of an amino acid residue for an active site acidic amino acid residue. For example, when the endoglycanase is endo-H, the substituted active site residues will typically be Asp at position 130, Glu at position 132 or a combination thereof. The amino acids are generally replaced with serine, alanine, asparagine, or glutamine.

[0352] The mutant enzyme catalyzes the reaction, usually by a synthesis step that is analogous to the reverse reaction of the endoglycanase hydrolysis step. In these

embodiments, the glycosyl donor molecule (*e.g.*, a desired oligo- or mono-saccharide structure) contains a leaving group and the reaction proceeds with the addition of the donor molecule to a GlcNAc residue on the protein. For example, the leaving group can be a halogen, such as fluoride. In other embodiments, the leaving group is a Asn, or a Asn-peptide moiety. In further embodiments, the GlcNAc residue on the glycosyl donor molecule is modified. For example, the GlcNAc residue may comprise a 1,2 oxazoline moiety.

[0353] In a preferred embodiment, each of the enzymes utilized to produce a conjugate of the invention are present in a catalytic amount. The catalytic amount of a particular enzyme varies according to the concentration of that enzyme's substrate as well as to reaction conditions such as temperature, time and pH value. Means for determining the catalytic amount for a given enzyme under preselected substrate concentrations and reaction conditions are well known to those of skill in the art.

[0354] The temperature at which an above process is carried out can range from just above freezing to the temperature at which the most sensitive enzyme denatures. Preferred temperature ranges are about 0 °C to about 55 °C, and more preferably about 20 ° C to about 37 °C. In another exemplary embodiment, one or more components of the present method are conducted at an elevated temperature using a thermophilic enzyme.

[0355] The reaction mixture is maintained for a period of time sufficient for the acceptor to be glycosylated, thereby forming the desired conjugate. Some of the conjugate can often be detected after a few h, with recoverable amounts usually being obtained within 24 h or less. Those of skill in the art understand that the rate of reaction is dependent on a number of variable factors (*e.g*, enzyme concentration, donor concentration, acceptor concentration, temperature, solvent volume), which are optimized for a selected system.

[0356] The present invention also provides for the industrial-scale production of modified peptides. As used herein, an industrial scale generally produces at least one gram of finished, purified conjugate.

[0357] In the discussion that follows, the invention is exemplified by the conjugation of modified sialic acid moieties to a glycosylated peptide. The exemplary modified sialic acid is labeled with PEG. The focus of the following discussion on the use of PEG-modified sialic acid and glycosylated peptides is for clarity of illustration and is not intended to imply that the invention is limited to the conjugation of these two partners. One of skill understands that the discussion is generally applicable to the additions of modified glycosyl moieties other

than sialic acid. Moreover, the discussion is equally applicable to the modification of a glycosyl unit with agents other than PEG including other PEG moieties, therapeutic moieties, and biomolecules.

[0358] An enzymatic approach can be used for the selective introduction of PEGylated or PPGylated carbohydrates onto a peptide or glycopeptide. The method utilizes modified sugars containing PEG, PPG, or a masked reactive functional group, and is combined with the appropriate glycosyltransferase or glycosynthase. By selecting the glycosyltransferase that will make the desired carbohydrate linkage and utilizing the modified sugar as the donor substrate, the PEG or PPG can be introduced directly onto the peptide backbone, onto existing sugar residues of a glycopeptide or onto sugar residues that have been added to a peptide.

[0359] In an exemplary embodiment, an acceptor for a sialyltransferase is present on the peptide to be modified either as a naturally occurring structure or it is placed there recombinantly, enzymatically or chemically. Suitable acceptors, include, for example, galactosyl acceptors such as Galβ1,4GlcNAc, Galβ1,4GalNAc, Galβ1,3GalNAc, lacto-N-tetraose, Galβ1,3GlcNAc, Galβ1,3Ara, Galβ1,6GlcNAc, Galβ1,4Glc (lactose), and other acceptors known to those of skill in the art (*see, e.g.*, Paulson *et al., J. Biol. Chem.* **253**: 5617-5624 (1978)). Exemplary sialyltransferases are set forth herein.

[0360] In one embodiment, an acceptor for the sialyltransferase is present on the glycopeptide to be modified upon *in vivo* synthesis of the glycopeptide. Such glycopeptides can be sialylated using the claimed methods without prior modification of the glycosylation pattern of the glycopeptide. Alternatively, the methods of the invention can be used to sialylate a peptide that does not include a suitable acceptor; one first modifies the peptide to include an acceptor by methods known to those of skill in the art. In an exemplary embodiment, a GalNAc residue is added by the action of a GalNAc transferase.

[0361] In an exemplary embodiment, the galactosyl acceptor is assembled by attaching a galactose residue to an appropriate acceptor linked to the peptide, e.g., a GlcNAc. The method includes incubating the peptide to be modified with a reaction mixture that contains a suitable amount of a galactosyltransferase (*e.g.*, Galβ1,3 or Galβ1,4), and a suitable galactosyl donor (*e.g.*, UDP-galactose). The reaction is allowed to proceed substantially to completion or, alternatively, the reaction is terminated when a preselected amount of the

galactose residue is added. Other methods of assembling a selected saccharide acceptor will be apparent to those of skill in the art.

[0362] In yet another embodiment, glycopeptide-linked oligosaccharides are first "trimmed," either in whole or in part, to expose either an acceptor for the sialyltransferase or a moiety to which one or more appropriate residues can be added to obtain a suitable acceptor. Enzymes such as glycosyltransferases and endoglycosidases (*see*, for example U.S. Patent No. 5,716,812) are useful for the attaching and trimming reactions. In another embodiment of this method, the sialic acid moieties of the peptide are essentially completely removed (e.g., at least 90, at least 95 or at least 99%), exposing an acceptor for a modified sialic acid.

[0363] In the discussion that follows, the method of the invention is exemplified by the use of modified sugars having a PEG moiety attached thereto. The focus of the discussion is for clarity of illustration. Those of skill will appreciate that the discussion is equally relevant to those embodiments in which the modified sugar bears a therapeutic moiety, biomolecule or the like.

[0364] In an exemplary embodiment of the invention in which a carbohydrate residue is "trimmed" prior to the addition of the modified sugar high mannose is trimmed back to the first generation biantennary structure. A modified sugar bearing a PEG moiety is conjugated to one or more of the sugar residues exposed by the "trimming back." In one example, a PEG moiety is added via a GlcNAc moiety conjugated to the PEG moiety. The modified GlcNAc is attached to one or both of the terminal mannose residues of the biantennary structure. Alternatively, an unmodified GlcNAc can be added to one or both of the termini of the branched species.

[0365] In another exemplary embodiment, a PEG moiety is added to one or both of the terminal mannose residues of the biantennary structure via a modified sugar having a galactose residue, which is conjugated to a GlcNAc residue added onto the terminal mannose residues. Alternatively, an unmodified Gal can be added to one or both terminal GlcNAc residues.

[0366] In yet a further example, a PEG moiety is added onto a Gal residue using a modified sialic acid such as those discussed above.

[0367] In another exemplary embodiment, a high mannose structure is "trimmed back" to the mannose from which the biantennary structure branches. In one example, a PEG moiety is added via a GlcNAc modified with the polymer. Alternatively, an unmodified GlcNAc is added to the mannose, followed by a Gal with an attached PEG moiety. In yet another embodiment, unmodified GlcNAc and Gal residues are sequentially added to the mannose, followed by a sialic acid moiety modified with a PEG moiety.

[0368] A high mannose structure can also be trimmed back to the elementary tri-mannosyl core.

[0369] In a further exemplary embodiment, high mannose is "trimmed back" to the GlcNAc to which the first mannose is attached. The GlcNAc is conjugated to a Gal residue bearing a PEG moiety. Alternatively, an unmodified Gal is added to the GlcNAc, followed by the addition of a sialic acid modified with a water-soluble sugar. In yet a further example, the terminal GlcNAc is conjugated with Gal and the GlcNAc is subsequently fucosylated with a modified fucose bearing a PEG moiety.

[0370] High mannose may also be trimmed back to the first GlcNAc attached to the Asn of the peptide. In one example, the GlcNAc of the GlcNAc-$(Fuc)_a$ residue is conjugated wit ha GlcNAc bearing a water soluble polymer. In another example, the GlcNAc of the GlcNAc-$(Fuc)_a$ residue is modified with Gal, which bears a water soluble polymer. In a still further embodiment, the GlcNAc is modified with Gal, followed by conjugation to the Gal of a sialic acid modified with a PEG moiety.

[0371] Other exemplary embodiments are set forth in commonly owned U.S. Patent application Publications: 20040132640; 20040063911; 20040137557; U.S. Patent application Nos: 10/369,979; 10/410,913; 10/360,770; 10/410,945 and PCT/US02/32263 each of which is incorporated herein by reference.

[0372] The Examples set forth above provide an illustration of the power of the methods set forth herein. Using the methods described herein, it is possible to "trim back" and build up a carbohydrate residue of substantially any desired structure. The modified sugar can be added to the termini of the carbohydrate moiety as set forth above, or it can be intermediate between the peptide core and the terminus of the carbohydrate.

[0373] In an exemplary embodiment, an existing sialic acid is removed from a glycopeptide using a sialidase, thereby unmasking all or most of the underlying galactosyl

residues. Alternatively, a peptide or glycopeptide is labeled with galactose residues, or an oligosaccharide residue that terminates in a galactose unit. Following the exposure of or addition of the galactose residues, an appropriate sialyltransferase is used to add a modified sialic acid.

[0374] In another exemplary embodiment, an enzyme that transfers sialic acid onto sialic acid is utilized. This method can be practiced without treating a sialylated glycan with a sialidase to expose glycan residues beneath the sialic acid. An exemplary polymer-modified sialic acid is a sialic acid modified with poly(ethylene glycol). Other exemplary enzymes that add sialic acid and modified sialic acid moieties onto glycans that include a sialic acid residue or exchange an existing sialic acid residue on a glycan for these species include ST3Gal3, CST-II, ST8Sia-II, ST8Sia-III and ST8Sia-IV.

[0375] In yet a further approach, a masked reactive functionality is present on the sialic acid. The masked reactive group is preferably unaffected by the conditions used to attach the modified sialic acid to the Factor VII/Factor VIIa peptide. After the covalent attachment of the modified sialic acid to the peptide, the mask is removed and the peptide is conjugated with an agent such as PEG. The agent is conjugated to the peptide in a specific manner by its reaction with the unmasked reactive group on the modified sugar residue.

[0376] Any modified sugar can be used with its appropriate glycosyltransferase, depending on the terminal sugars of the oligosaccharide side chains of the glycopeptide. As discussed above, the terminal sugar of the glycopeptide required for introduction of the PEGylated structure can be introduced naturally during expression or it can be produced post expression using the appropriate glycosidase(s), glycosyltransferase(s) or mix of glycosidase(s) and glycosyltransferase(s).

[0377] In a further exemplary embodiment, UDP-galactose-PEG is reacted with β1,4-galactosyltransferase, thereby transferring the modified galactose to the appropriate terminal N-acetylglucosamine structure. The terminal GlcNAc residues on the glycopeptide may be produced during expression, as may occur in such expression systems as mammalian, insect, plant or fungus, but also can be produced by treating the glycopeptide with a sialidase and/or glycosidase and/or glycosyltransferase, as required.

[0378] In another exemplary embodiment, a GlcNAc transferase, such as GNT1-5, is utilized to transfer PEGylated-GlcNAc to a terminal mannose residue on a glycopeptide. In a still further exemplary embodiment, an the N- and/or O-linked glycan structures are

enzymatically removed from a glycopeptide to expose an amino acid or a terminal glycosyl residue that is subsequently conjugated with the modified sugar. For example, an endoglycanase is used to remove the N-linked structures of a glycopeptide to expose a terminal GlcNAc as a GlcNAc-linked-Asn on the glycopeptide. UDP-Gal-PEG and the appropriate galactosyltransferase is used to introduce the PEG-galactose functionality onto the exposed GlcNAc.

[0379] In an alternative embodiment, the modified sugar is added directly to the peptide backbone using a glycosyltransferase known to transfer sugar residues to the peptide backbone. Exemplary glycosyltransferases useful in practicing the present invention include, but are not limited to, GalNAc transferases (GalNAc T1-14), GlcNAc transferases, fucosyltransferases, glucosyltransferases, xylosyltransferases, mannosyltransferases and the like. Use of this approach allows the direct addition of modified sugars onto peptides that lack any carbohydrates or, alternatively, onto existing glycopeptides. In both cases, the addition of the modified sugar occurs at specific positions on the peptide backbone as defined by the substrate specificity of the glycosyltransferase and not in a random manner as occurs during modification of a protein's peptide backbone using chemical methods. An array of agents can be introduced into proteins or glycopeptides that lack the glycosyltransferase substrate peptide sequence by engineering the appropriate amino acid sequence into the polypeptide chain.

[0380] In each of the exemplary embodiments set forth above, one or more additional chemical or enzymatic modification steps can be utilized following the conjugation of the modified sugar to the peptide. In an exemplary embodiment, an enzyme (*e.g.*, fucosyltransferase) is used to append a glycosyl unit (*e.g.*, fucose) onto the terminal modified sugar attached to the peptide. In another example, an enzymatic reaction is utilized to "cap" sites to which the modified sugar failed to conjugate. Alternatively, a chemical reaction is utilized to alter the structure of the conjugated modified sugar. For example, the conjugated modified sugar is reacted with agents that stabilize or destabilize its linkage with the peptide component to which the modified sugar is attached. In another example, a component of the modified sugar is deprotected following its conjugation to the peptide. One of skill will appreciate that there is an array of enzymatic and chemical procedures that are useful in the methods of the invention at a stage after the modified sugar is conjugated to the peptide. Further elaboration of the modified sugar-peptide conjugate is within the scope of the invention.

[0381] Enzymes and reaction conditions for preparing the conjugates of the present invention are discussed in detail in the parent of the instant application as well as co-owned published PCT patent applications WO 03/031464, WO 04/033651, WO 04/099231.

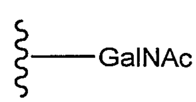
[0382] In a selected embodiment, a Factor VII/Factor VIIa peptide, expressed in insect cells, is remodeled such that glycans on the remodeled glycopeptide include a GlcNAc-Gal glycosyl residue. The addition of GlcNAc and Gal can occur as separate reactions or as a single reaction in a single vessel. In this example, GlcNAc-transferase I and Gal-transferase I are used. The modified sialyl moiety is added using ST3Gal-III.

[0383] In another embodiment, the addition of GlcNAc, Gal and modified Sia can also occur in a single reaction vessel, using the enzymes set forth above. Each of the enzymatic remodeling and glycoPEGylation steps are carried out individually.

[0384] When the peptide is expressed in mammalian cells, different methods are of use. In one embodiment, the peptide is conjugated without need for remodeling prior to conjugation by contacting the peptide with a sialyltransferase that transfers the modified sialic acid directly onto a sialic acid on the peptide forming Sia-Sia-L-R^1, or exchanges a sialic acid on the peptide for the modified sialic acid, forming Sia-L-R^1. An exemplary enzyme of use in this method is CST-II. Other enzymes that add sialic acid to sialic acid are known to those of skill in the art and examples of such enzymes are set forth the figures appended hereto.

[0385] In yet another method of preparing the conjugates of the invention, the peptide expressed in a mammalian system is desialylated using a sialidase. The exposed Gal residue is sialylated with a modified sialic acid using a sialyltransferase specific for O-linked glycans, providing a Factor VII/Factor VIIa peptide with an O-linked modified glycan. The desialylated, modified Factor VII/Factor VIIa peptide is optionally partially or fully re-sialylated by using a sialyltransferase such as ST3GalIII.

[0386] In another aspect, the invention provides a method of making a PEGylated Factor VII/Factor VIIa peptide conjugate of the invention. The method includes: (a) contacting a Factor VII/Factor VIIa peptide comprising a glycosyl group selected from:



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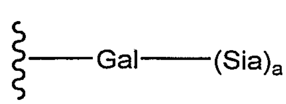
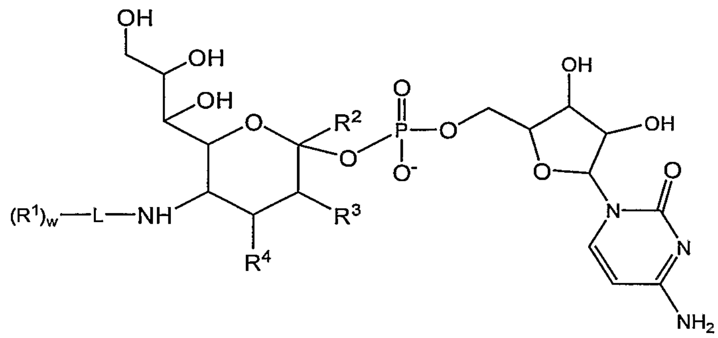


with a PEG-sialic acid donor having the formula which is a member selected from



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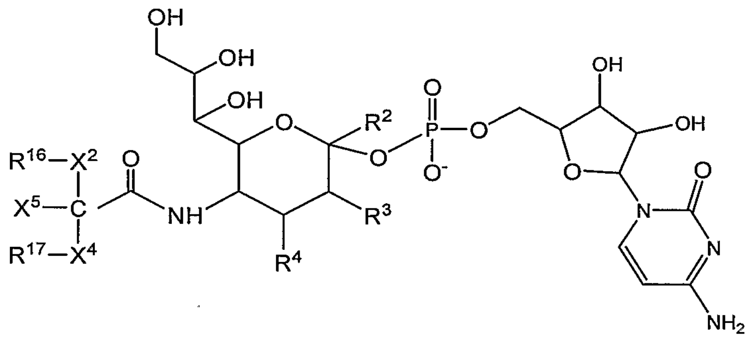

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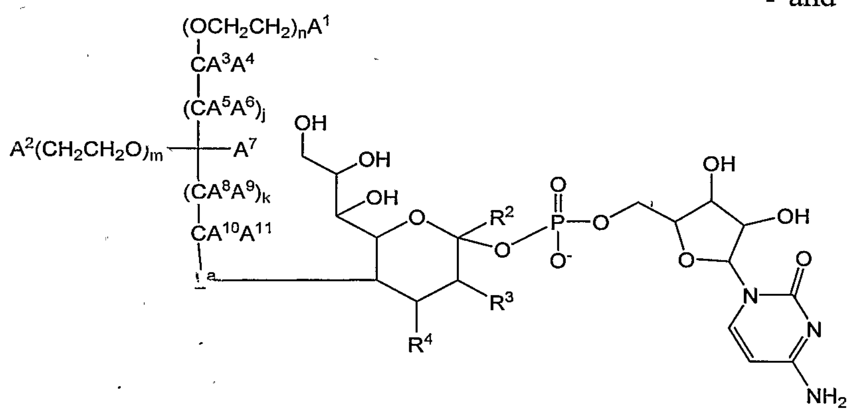

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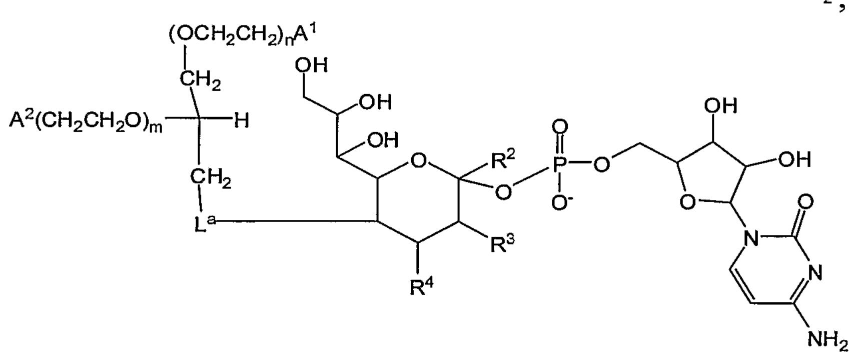
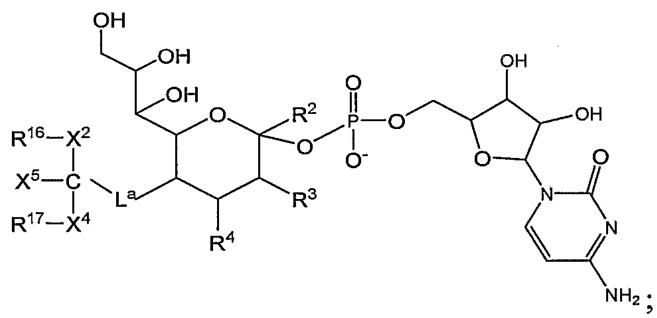


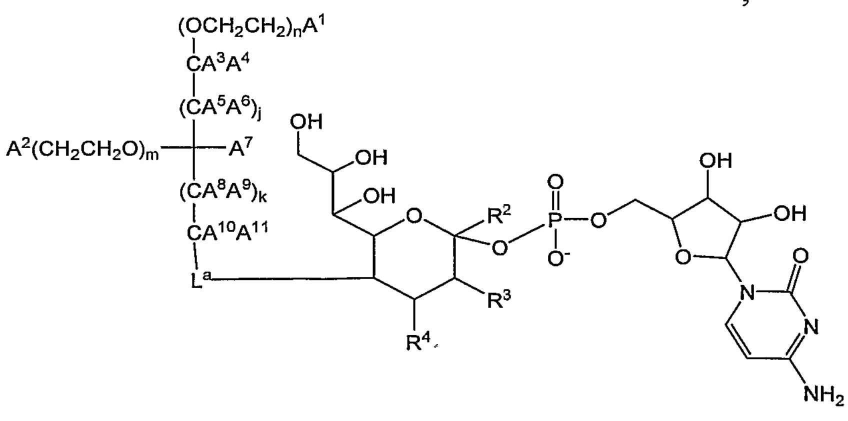
and an enzyme that transfers PEG-sialic acid from said donor onto a member selected from the GalNAc, Gal and the Sia of said glycosyl group, under conditions appropriate for said transfer. An exemplary modified sialic acid donor is CMP-sialic acid modified, through a linker moiety, with a polymer, e.g., a straight chain or branched poly(ethylene glycol) moiety. As discussed herein, the peptide is optionally glycosylated with GalNAc and/or Gal and/or

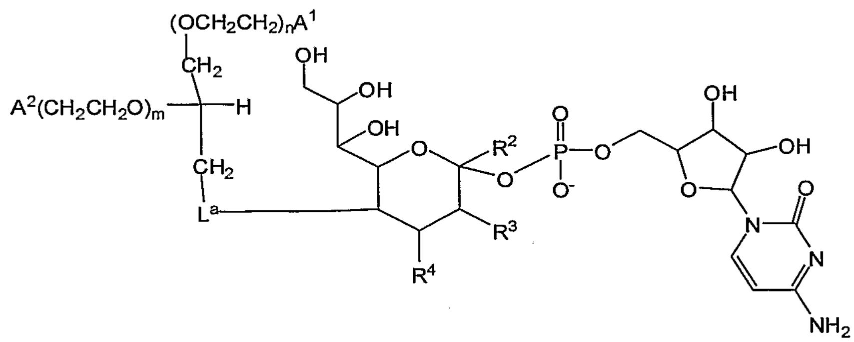
Sia ("Remodeled") prior to attaching the modified sugar. The remodeling steps can occur in sequence in the same vessel without purification of the glycosylated peptide between steps. Alternatively, following one or more remodeling step, the glycosylated peptide can be purified prior to submitting it to the next glycosylation or glycPEGylation step. In an exemplary embodiment, the method further comprises expressing the peptide in a host. In an exemplary embodiment, the host is a mammalian cell or an insect cell. In another exemplary embodiment, the mammalian cell is a member selected from a BHK cell and a CHO cell and the insect cell is a *Spodoptera frugiperda* cell.

[0387] As illustrated in the examples and discussed further below, placement of an acceptor moiety for the PEG-sugar is accomplished in any desired number of steps. For example, in one embodiment, the addition of GalNAc to the peptide can be followed by a second step in which the PEG-sugar is conjugated to the GalNAc in the same reaction vessel. Alternatively, these two steps can be carried out in a single vessel approximately simultaneously.

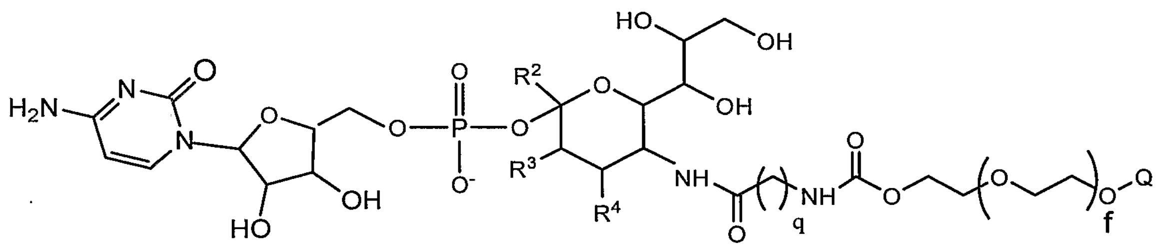
[0388] In an exemplary embodiment, the PEG-sialic acid donor has the formula:





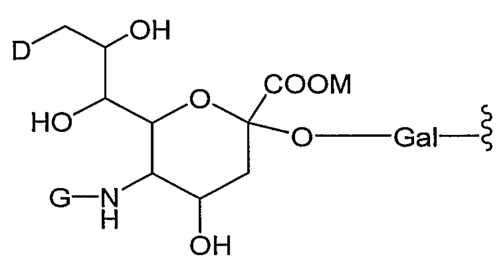


[0389] In another exemplary emodiment, the PEG-sialic acid donor has the formula:



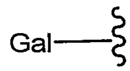
[0390] In a further exemplary embodiment, the Factor VII/Factor VIIa peptide is expressed in an appropriate expression system prior to being glycopegylated or remodeled. Exemplary expression systems include Sf-9/baculovirus and Chinese Hamster Ovary (CHO) cells.

[0391] In an exemplary embodiment, the invention provides a method of making a Factor VII/Factor VIIa peptide conjugate comprising a glycosyl linker comprising a modified sialyl residue having the formula:

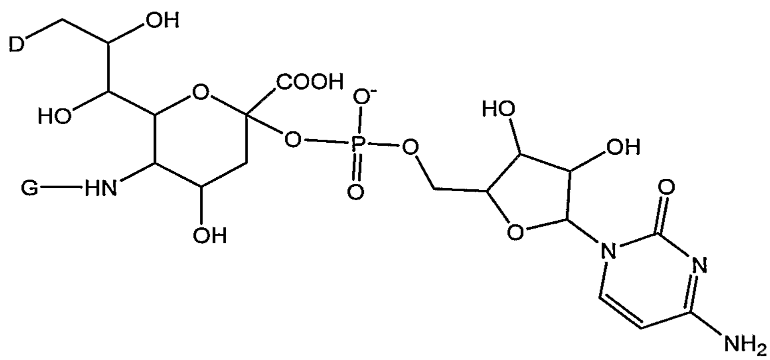


wherein D is a member selected from -OH and R^1-L-HN-; G is a member selected from R^1-L- and -C(O)(C_1-C_6)alkyl-R^1; R^1 is a moiety comprising a member selected from a straight-chain poly(ethylene glycol) residue and branched poly(ethylene glycol) residue; M is a member selected from H, a metal and a single negative charge; L is a linker which is a member selected from a bond, substituted or unsubstituted alkyl and substituted or unsubstituted heteroalkyl, such that when D is OH, G is R^1-L-, and when G is –C(O)(C_1-C_6)alkyl, D is R^1-L-NH-

said method comprising: (a) contacting a Factor VII/Factor VIIa peptide comprising the glycosyl moiety:

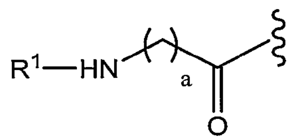


with a PEG-sialic acid donor moiety having the formula:



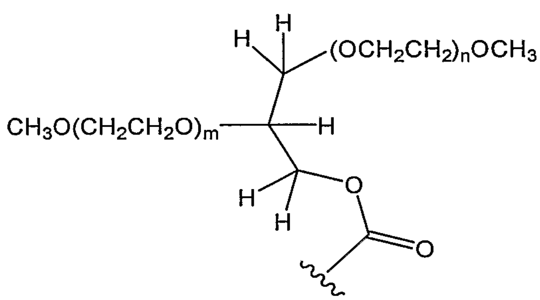
and an enzyme that transfers said PEG-sialic acid onto the Gal of said glycosyl moiety, under conditions appropriate for said transfer.

[0392] In an exemplary embodiment, L-R^1 has the formula:



wherein a is an integer selected from 0 to 20.

[0393] In another exemplary embodiment, R^1 has a structure that is a member selected from:



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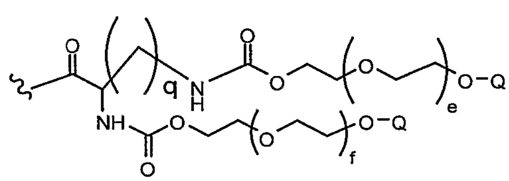
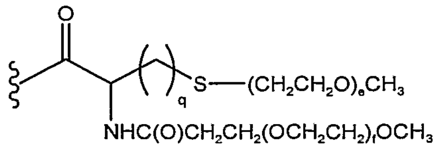
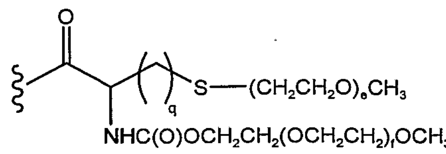
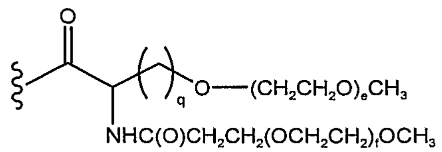
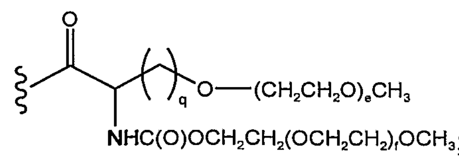


 ; 

 ; and 

wherein e, f, m and n are integers independently selected from 1 to 2500; and q is an integer selected from 0 to 20.

[0394] Large scale or small scale amounts of Factor VII/Factor VIIa peptide conjugate can be produced by the methods described herein. In an exemplary embodiment, the amount of Factor VII/Factor VIIa peptide is a member selected from about 0.5 mg to about 100kg. In an exemplary embodiment, the amount of Factor VII/Factor VIIa peptide is a member selected from about 0.1 kg to about 1 kg. In an exemplary embodiment, the amount of Factor VII/Factor VIIa peptide is a member selected from about 0.5 kg to about 10kg. In an exemplary embodiment, the amount of Factor VII/Factor VIIa peptide is a member selected from about 0.5 kg to about 3kg. In an exemplary embodiment, the amount of Factor VII/Factor VIIa peptide is a member selected from about 0.1 kg to about 5kg. In an exemplary embodiment, the amount of Factor VII/Factor VIIa peptide is a member selected from about 0.08 kg to about 0.2 kg. In an exemplary embodiment, the amount of Factor VII/Factor VIIa peptide is a member selected from about 0.05 kg to about 0.4kg. In an exemplary embodiment, the amount of Factor VII/Factor VIIa peptide is a member selected from about 0.1 kg to about 0.7kg. In an exemplary embodiment, the amount of Factor VII/Factor VIIa peptide is a member selected from about 0.3 kg to about 1.75 kg. In an exemplary embodiment, the amount of Factor VII/Factor VIIa peptide is a member selected from about 25 kg to about 65kg.

[0395] The concentration of Factor VII/Factor VIIa peptide utilized in the reactions described herein is a member selected from about 0.5 to about 10 mg Factor VII/Factor VIIa peptide/mL reaction mixture. In an exemplary embodiment, the Factor VII/Factor VIIa peptide concentration is a member selected from about 0.5 to about 1 mg Factor VII/Factor VIIa peptide/mL reaction mixture. In an exemplary embodiment, the Factor VII/Factor VIIa peptide concentration is a member selected from about 0.8 to about 3 mg Factor VII/Factor

VIIa peptide/mL reaction mixture. In an exemplary embodiment, the Factor VII/Factor VIIa peptide concentration is a member selected from about 2 to about 6 mg Factor VII/Factor VIIa peptide/mL reaction mixture. In an exemplary embodiment, the Factor VII/Factor VIIa peptide concentration is a member selected from about 4 to about 9 mg Factor VII/Factor VIIa peptide/mL reaction mixture. In an exemplary embodiment, the Factor VII/Factor VIIa peptide concentration is a member selected from about 1.2 to about 7.8 mg Factor VII/Factor VIIa peptide/mL reaction mixture. In an exemplary embodiment, the Factor VII/Factor VIIa peptide concentration is a member selected from about 6 to about 9.5 mg Factor VII/Factor VIIa peptide/mL reaction mixture.

[0396] The concentration of CMP-SA-PEG that can be utilized in the reactions described herein is a member selected from about 0.1 to about 1.0 mM. Factors which may increase or decrease the concentration include the size of the PEG, time of incubation, temperature, buffer components, as well as the type, and concentration, of glycosyltransferase used. In an exemplary embodiment, CMP-SA-PEG concentration is a member selected from about 0.1 to about 1.0 mM. In an exemplary embodiment, CMP-SA-PEG concentration is a member selected from about 0.1 to about 0.5 mM. In an exemplary embodiment, CMP-SA-PEG concentration is a member selected from about 0.1 to about 0.3 mM. In an exemplary embodiment, CMP-SA-PEG concentration is a member selected from about 0.2 to about 0.7 mM. In an exemplary embodiment, CMP-SA-PEG concentration is a member selected from about 0.3 to about 0.5 mM. In an exemplary embodiment, CMP-SA-PEG concentration is a member selected from about 0.4 to about 1.0 mM. In an exemplary embodiment, CMP-SA-PEG concentration is a member selected from about 0.5 to about 0.7 mM. In an exemplary embodiment, CMP-SA-PEG concentration is a member selected from about 0.8 to about 0.95 mM. In an exemplary embodiment, CMP-SA-PEG concentration is a member selected from about 0.55 to about 1.0 mM.

[0397] The molar equivalents of CMP-SA-PEG that can be utilized in the reactions described herein are based on the theoretical number of SA-PEGs that can be added to the Factor VII/Factor VIIa protein. The theoretical number of SA-PEGs is based on the theoretical number of sialation sites on the Factor VII/Factor VIIa protein as well as the MW of the Factor VII/Factor VIIa protein when compared to the MW and therefore moles of CMP-SA-PEG. For Factor VII/Factor VIIa, that is about four or five PEGs based on N-glycans that are primarily bi- and tri-antennary with only two glycan sites. In an exemplary embodiment, the molar equivalents of CMP-SA-PEG is an integer selected from 1 to 20. In

an exemplary embodiment, the molar equivalents of CMP-SA-PEG is an integer selected from 1 to 20. In an exemplary embodiment, the molar equivalents of CMP-SA-PEG is an integer selected from 2 to 6. In an exemplary embodiment, the molar equivalents of CMP-SA-PEG is an integer selected from 3 to 17. In an exemplary embodiment, the molar equivalents of CMP-SA-PEG is an integer selected from 4 to 11. In an exemplary embodiment, the molar equivalents of CMP-SA-PEG is an integer selected from 5 to 20. In an exemplary embodiment, the molar equivalents of CMP-SA-PEG is an integer selected from 1 to 10. In an exemplary embodiment, the molar equivalents of CMP-SA-PEG is an integer selected from 12 to 20. In an exemplary embodiment, the molar equivalents of CMP-SA-PEG is an integer selected from 14 to 17. In an exemplary embodiment, the molar equivalents of CMP-SA-PEG is an integer selected from 7 to 15. In an exemplary embodiment, the molar equivalents of CMP-SA-PEG is an integer selected from 8 to 16.

III. B. Simultaneous Desialylation and GlycoPEGylation of Factor VII/Factor VIIa

[0398] The present invention provides a "one-pot" method of glycopegylating Factor VII/Factor VIIa. The one-pot method is distinct from other exemplary processes to make a Factor VII/Factor VIIa peptide conjugate, which employ a sequential de-sialylation with sialidase, subsequent purification of the asialo Factor VII/Factor VIIa on an anion exchange column, then glycoPEGylation using CMP-sialic acid-PEG and a glycosyltransferase (such as ST3Gal3), exoglycosidase or an endoglycosidase. The Factor VII/Factor VIIa peptide conjugate is then purified via anion exchange followed by size exclusion chromatography to produce the purified Factor VII/Factor VIIa peptide conjugate.

[0399] The one-pot method is an improved method to manufacture a Factor VII/Factor VIIa peptide conjugate. In this method, the de-sialylation and glycoPEGylation reactions are combined in a one-pot reaction which obviates the first anion exchange chromatography step used in the previously described process to purify the asialo Factor VII/Factor VIIa peptide. This reduction in process steps produces several advantages. First, the number of process steps required to produce the Factor VII/Factor VIIa peptide conjugate is reduced, which also reduces the operating complexity of the process. Second, the process time for the production of the peptide conjugates is reduced e.g., from 4 to 2 days. This reduces the raw material requirements and quality control costs associated with in-process controls. Third, the invention utilizes less sialidase, e.g., up to 20-fold less sialidase, e.g., 500 mU/L is required to produce the Factor VII/Factor VIIa peptide conjugate relative to the process. This reduction in the use of sialidase significantly reduces the amount of contaminants, such as sialidase, in the reaction mixture.

[0400] In an exemplary embodiment, a Factor VII/Factor VIIa peptide conjugate is prepared by the following method. In a first step, a Factor VII/Factor VIIa peptide is combined with a sialidase, a modified sugar of the invention, and an enzyme capable of catalyzing the transfer of the glycosyl linking group from the modified sugar to the peptide, thus preparing the Factor VII/Factor VIIa peptide conjugate. Any sialidase may be used in this method. Exemplary sialidases of use in the invention can be found in the CAZY database (*see* http://afmb.cnrs-mrs.fr/CAZY/index.html and **www.cazy.org/CAZY**). Exemplary sialidases can be purchased from any number of sources (QA-Bio, Calbiochem, Marukin, Prozyme, etc.). In an exemplary embodiment, the sialidase is a member selected from cytoplasmic sialidases, lysosomal sialidases, exo-α sialidases, and endosialidases. In another exemplary embodiment, the sialidase used is produced from bacteria such as *Clostridium perfringens* or *Streptococcus pneumoniae,* or from a virus such as an adenovirus. In an exemplary embodiment, the enzyme capable of catalyzing the transfer of the glycosyl linking group from the modified sugar to the peptide is a member selected from a glycosyltransferase, such as sialyltransferases and fucosyltransferases, as well as exoglycosidases and endoglycosidases. In an exemplary embodiment, the enzyme is a glycosyltransferase, which is ST3Gal3. In another exemplary embodiment, the enzyme used is produced from bacteria such as *Escherichia Coli* or a fungus such as *Aspergillus niger*. In another exemplary embodiment, the sialidase is added to the Factor VII/Factor VIIa peptide before the glycosyltransferase for a specified time, allowing the sialidase reaction to proceed before initiating the GlycoPEGylation reaction with addition of the PEG-sialic acid reagent and the glycosyltransferase. Many of these examples are discussed herein. Finally, any modified sugar described herein can be utilized in this reaction.

[0401] In another exemplary embodiment, the method further comprises a 'capping' step. In this step, additional non-PEGylated sialic acid is added to the reaction mixture. In an exemplary embodiment, this sialic acid is added to the Factor VII/Factor VIIa peptide or peptide conjugate thus preventing further addition of PEG-sialic acid. In another exemplary embodiment, this sialic acid impedes the function of the glycosyltransferase in the reaction mixture, effectively stopping the addition of glycosyl linking groups to the Factor VII/Factor VIIa peptides or peptide conjugates. Most importantly, the sialic acid that is added to the reaction mixture caps the unglycoPEGylated glycans thereby providing a Factor VII/Factor VIIa peptide conjugate that has improved pharmaceokinetics. In addition, this sialidase can

be added directly the the glycoPEGylation reaction mixture when the extent of PEGylation to certain amounts is desired without prior purification.

[0402] In an exemplary embodiment, after the capping step, less than about 50% of the sialylation sites on the Factor VII/Factor VIIa peptide or peptide conjugate does not comprise a sialyl moiety. In an exemplary embodiment, after the capping step, less than about 40% of the sialylation sites on the Factor VII/Factor VIIa peptide or peptide conjugate does not comprise a sialyl moiety. In an exemplary embodiment, after the capping step, less than about 30% of the sialylation sites on the Factor VII/Factor VIIa peptide or peptide conjugate does not comprise a sialyl moiety. In an exemplary embodiment, after the capping step, less than about 20% of the sialylation sites on the Factor VII/Factor VIIa peptide or peptide conjugate does not comprise a sialyl moiety. In an exemplary embodiment, after the capping step, less than about 10% of the sialylation sites on the Factor VII/Factor VIIa peptide or peptide conjugate does not comprise a sialyl moiety. In an exemplary embodiment, between about 20% and about 5% of the sialylation sites on the Factor VII/Factor VIIa peptide or peptide conjugate does not comprise a sialyl moiety. In an exemplary embodiment, between about 25% and about 10% of the sialylation sites on the Factor VII/Factor VIIa peptide or peptide conjugate does not comprise a sialyl moiety. In an exemplary embodiment, after the capping step, essentially all of the sialylation sites on the Factor VII/Factor VIIa peptide or peptide conjugate comprise a sialyl moiety.

III. C. Desialylation and Selective Modification of Factor VII/Factor VIIa Peptides

[0403] In another exemplary embodiment, the present invention provides a method for desialylating a Factor VII/Factor VIIa peptide. The method preferably provides a Factor VII/Factor VIIa peptide that is at least about 40%, preferably 45%, preferably about 50%, preferably about 55%, preferably about 60%, preferably about 65%, preferably about 70%, preferably about 75%, preferably about 80%, preferably at least 85%, more preferably at least 90%, still more preferably, at least 92%, preferably at least 94%, even more preferably at least 96%, still more preferably at least 98%, and still more preferably 100% disialylated.

[0404] The method includes contacting the Factor VII/Factor VIIa peptide with a sialidase, preferably for a time period. The preselected time period is sufficient to desialylate the Factor VII/Factor VIIa peptide to the degree desired. In a preferred embodiment, the desialylated Factor VII/Factor VIIa peptide is separated from the sialidase when the desired

degree of desialylation is achieved. An exemplary desialylation reaction and purification cycle is set forth herein.

[0405] Those of skill are able to determine an appropriate preselected time period over which to conduct the desialylation reaction. In an exemplary embodiment, the period is less than 24 hours, preferably less than 8 hours, more preferably less than 6 hours, more preferably less than 4 hours, still more preferably less than 2 hours and even more preferably less than 1 hour.

[0406] In another exemplary embodiment, in the Factor VII/Factor VIIa preparation at the end of the desialylation reaction, at least 10% of the members of the population of Factor VII/Factor VIIa peptides has only a single sialic acid attached thereto, preferably at least 20%, more preferably at least 30%, still more preferably at least 40%, even still more preferably at least 50% and more preferably at least 60%, and still more preferably completely desialylated.

[0407] In yet a further exemplary embodiment, in the Factor VII/Factor VIIa preparation at the end of the desialylation reaction, at least 10% of the members of the population of Factor VII/Factor VIIa peptides is fully desialylated, preferably at least 20%, more preferably at least 30%, even more preferably at least 40%, still more preferably at least 50% and even still more preferably at least 60%.

[0408] In still another exemplary embodiment, in the Factor VII/Factor VIIa preparation at the end of the desialylation reaction, at least 10%, 20%, 30%, 40%, 50% or 60% of the members of the Factor VII/Factor VIIa peptide population has only a single sialic acid, and at least 10%, 20%, 30%, 40%, 50% or 60% of the Factor VII/Factor VIIa peptide is fully disialylated.

[0409] In a preferred embodiment, in the Factor VII/Factor VIIa preparation at the end of the desialylation reaction, at least 50% of the population of Factor VII/Factor VIIa peptides is fully disialylated and at least 40% of the members of the Factor VII/Factor VIIa peptide population bears only a single sialic acid moiety.

[0410] Following desialylation, the Factor VII/Factor VIIa peptide is optionally conjugated with a modified sugar. An exemplary modified sugar includes a saccharyl moiety bound to a branched or linear poly(ethylene glycol) moiety. The conjugation is catalyzed by an enzyme that transfers the modified sugar from a modified sugar donor onto an amino acid or glycosyl

residue of the Factor VII/Factor VIIa peptide. An exemplary modified sugar donor is a CMP-sialic acid that bears a branched or linear poly(ethylene glycol) moiety. An exemplary poly(ethylene glycol) moiety has a molecular weight of at least about 2 KDa, more preferably at least about 5 KDa, more preferably at least about 10 KDa, preferably at least about 20 KDa, more preferably at least about 30 KDa, and more preferably at least about 40 KDa.

[0411] In an exemplary embodiment, the enzyme utilized to transfer the modified sugar moiety from the modified sugar donor is a glycosyltransferase, e.g., sialyltransferase. An exemplary sialyltransferase of use in the methods of the invention is ST3Gal3.

[0412] An exemplary method of the invention results in a modified Factor VII/Factor VIIa peptide bearing at least one, preferably at least two, preferably at least three modifying groups. In one embodiment, the Factor VII/Factor VIIa peptide produced bears a single modifying group on the light chain of the Factor VII/Factor VIIa peptide. In another embodiment, the method provides a modified Factor VII/Factor VIIa peptide that bears a single modifying group on the heavy chain. In still another embodiment, the method provides a modified Factor VII/Factor VIIa peptide with a single modifying group on the light chain and a single modifying group on the heavy chain.

[0413] In another aspect, the invention provides a method of preparing a modified Factor VII/Factor VIIa peptide. The method includes contacting the Factor VII/Factor VIIa peptide with a modified sugar donor bearing a modifying group and an enzyme capable of transferring a modified sugar moiety from the modified sugar donor onto an amino acid or glycosyl residue of the peptide.

[0414] In an exemplary embodiment, the method provides a population of modified Factor VII/Factor VIIa peptides in which at least 40%, preferably at least 50%, preferably at least 60%, more preferably at least 70% and even more preferably at least 80% of the population members are mono-conjugated on the light chain of the Factor VII/Factor VIIa peptide.

[0415] In an exemplary embodiment, the method provides a population of modified Factor VII/Factor VIIa peptides in which at least 40%, preferably at least 50%, preferably at least 60%, more preferably at least 70% and even more preferably at least 80% of the population members are di-conjugated on the light chain of the Factor VII/Factor VIIa peptide.

[0416] In an exemplary embodiment of this aspect, the method provides a population of modified Factor VII/Factor VIIa peptides in which no more than 50%, preferably no more

than 30%, preferably no more than 20%, more preferably no more than 10% of the population members are mono-conjugated on the heavy chain of the Factor VII/Factor VIIa peptide.

[0417] In an exemplary embodiment of this aspect, the method provides a population of modified Factor VII/Factor VIIa peptides in which no more than 50%, preferably no more than 30%, preferably no more than 20%, more preferably no more than 10% of the population members are di-conjugated on the heavy chain of the Factor VII/Factor VIIa peptide.

[0418] The Factor VII/Factor VIIa peptide can be subjected to the action of a sialidase prior to the contacting step, or the peptide can be used without prior desialylation. When the peptide is contacted with a sialidase it can be either essentially completely desialylated or only partially desialylated. In a preferred embodiment, the Factor VII/Factor VIIa peptide is at least partially desialylated prior to the contacting step. The Factor VII/Factor VIIa peptide may be essentially completely desialylated (essentially asialo) or only partially desialylated. In a preferred embodiment, the desialylated Factor VII/Factor VIIa peptide is one of the desialylated embodiments described hereinabove.

III. D. Additional aliquots of reagents added in the synthesis of Factor VII/Factor VIIa Peptide Conjugates

[0419] In an exemplary embodiment of the synthesis of the peptide conjugates described herein, one or more additional aliquots of a reaction component/reagent is added to the reaction mixture after a selected period of time. In an exemplary embodiment, the peptide conjugate is a Factor VII/Factor VIIa peptide conjugate. In another exemplary embodiment, the reaction component/reagent added is a modified sugar nucleotide. Introduction of a modified sugar nucleotide into the reaction will increase the likelihood of driving the GlycoPEGylation reaction to completion. In an exemplary embodiment, the nucleotide sugar is a CMP-SA-PEG described herein. In an exemplary embodiment, the reaction component/reagent added is a sialidase. In an exemplary embodiment, the reaction component/reagent added is a glycosyltransferase. In an exemplary embodiment, the reaction component/reagent added is magnesium. In an exemplary embodiment, the additional aliquot added represents about 10%, or 20%, or 30%, or 40%, or 50%, or 60%, or 70%, or 80% or 90% of the original amount in added at the start of the reaction. In an exemplary embodiment, the reaction component/reagent is added to the reaction about 3 hours, or 6

hours, or 8 hours, or 10 hours, or 12 hours, or 18 hours, or 24 hours, or 30 hours, or 36 hours after its start.

III. E. ***Selective production of light chain PEGylated Factor VII/Factor VIIa Peptide Conjugates***

[0420] In an exemplary embodiment, the invention provides a method of increasing the production of Factor VIIa peptide conjugates which are modified on the light chain over the heavy chain. This method involves the inactivation or sequestering of the heavy chain, thus allowing GlycoPEGylation to preferentially occur on the light chain. The serine protease activity of the heavy chain of Factor VIIa can be exploited as the basis for this sequestration. Adding a benzamidine matrix and/or pseudoaffinity resin for serine proteases to a GlycoPEGylation reaction mixture results in sequestration of the heavy chain, while GlycoPEGylation proceeds on the light chain. The light chain can then be purified away from the heavy chain by standard techniques known in the art. The heavy chain can be removed from the matrix by the addition of benzamidine or removed from the resin by lowering the pH of the solution. Benzamidine impurities introduced in this step can be removed by diafiltration.

III. E. ***Purification of Factor VII/Factor VIIa Peptide Conjugates***

[0421] The products produced by the above processes can be used without purification. However, it is usually preferred to recover the product and one or more of the intermediates, e.g., nucleotide sugars, branched and linear PEG species, modified sugars and modified nucleotide sugars. Standard, well-known techniques for recovery of glycosylated peptides such as thin or thick layer chromatography, column chromatography, ion exchange chromatography, or membrane filtration can be used. It is preferred to use membrane filtration, more preferably utilizing a reverse osmotic membrane, or one or more column chromatographic techniques for the recovery as is discussed hereinafter and in the literature cited herein. For instance, membrane filtration wherein the membranes have molecular weight cutoff of about 3000 to about 10,000 can be used to remove proteins such as glycosyl transferases. In certain instances, the molecular weight cutoff differences between the impurity and the product will be utilized in order to ensure product purification. For example, in order to purify product Factor VIIa-SA-PEG-40KDa from unreacted CMP-SA-PEG-40KDa, a filter must be chosen that will allow, for example, Factor VIIa-SA-PEG-40KDa to remain in the retentate while allowing CMP-SA-PEG-40KDa to flow into the filtrate. Nanofiltration or reverse osmosis can then be used to remove salts and/or purify the

product saccharides (*see, e.g.,* WO 98/15581). Nanofilter membranes are a class of reverse osmosis membranes that pass monovalent salts but retain polyvalent salts and uncharged solutes larger than about 100 to about 2,000 Daltons, depending upon the membrane used. Thus, in a typical application, saccharides prepared by the methods of the present invention will be retained in the membrane and contaminating salts will pass through.

[0422] If the peptide is produced intracellularly, as a first step, the particulate debris, either host cells or lysed fragments, is removed. Following glycoPEGylation, the PEGylated peptide is purified by art-recognized methods , for example, by centrifugation or ultrafiltration; optionally, the protein may be concentrated with a commercially available protein concentration filter, followed by separating the polypeptide variant from other impurities by one or more steps selected from immunoaffinity chromatography, ion-exchange column fractionation (*e.g.*, on diethylaminoethyl (DEAE) or matrices containing carboxymethyl or sulfopropyl groups), chromatography on Blue-Sepharose, CM Blue-Sepharose, MONO-Q, MONO-S, lentil lectin-Sepharose, WGA-Sepharose, Con A-Sepharose, Ether Toyopearl, Butyl Toyopearl, Phenyl Toyopearl, or protein A Sepharose, SDS-PAGE chromatography, silica chromatography, chromatofocusing, reverse phase HPLC (*e.g.*, silica gel with appended aliphatic groups), gel filtration using, *e.g.*, Sephadex molecular sieve or size-exclusion chromatography, chromatography on columns that selectively bind the polypeptide, and ethanol or ammonium sulfate precipitation. Purification can be used to separate one chain of the Factor VII/Factor VIIa peptide conjugate from the other, as further described later in this section.

[0423] Modified glycopeptides produced in culture are usually isolated by initial extraction from cells, enzymes, etc., followed by one or more concentration, salting-out, aqueous ion-exchange, or size-exclusion chromatography steps. Additionally, the modified glycoprotein may be purified by affinity chromatography. Finally, HPLC may be employed for final purification steps.

[0424] A protease inhibitor may be included in any of the foregoing steps to inhibit proteolysis and antibiotics or preservatives may be included to prevent the growth of adventitious contaminants. The protease inhibitors used in the foregoing steps may be low molecular weight inhibitors, including antipain, alpha-1-antitrypsin, anti-thrombin, leupeptin, amastatin, chymostatin, banzamidin, as well as other serine protease inhibitors (i.e. serpins). Generally, serine protease inhibitors should be used in concentrations ranging from 0.5 – 100

μM, although chymostatin in cell culture may be used in concentrations upward of 200 μM. Other serine protease inhibitors will include inhibitors specific to the chymotrypsin-like, the subtilisin-like, the alpha/beta hydrolase, or the signal peptidase clans of serine proteases. Besides serine proteases, other types of protease inhibitors may also be used, including cysteine protease inhibitors (1 - 10 μM) and aspartic protease inhibitors (1 -5 μM), as well as non-specific protease inhibitors such as pepstatin (.1 – 5 μM). Protease inhibitors used in this invention may also include natural protease inhibitors, such as the hirustasin inhibitor isolated from leech. In some embodiments, protease inhibitors will comprise synthetic peptides or antibodies that are able to bind with specificity to the protease catalytic site to stabilize Factor VII/Factor VIIa without interfering with a glycoPEGylation reaction.

[0425] Within another embodiment, supernatants from systems which produce the modified glycopeptide of the invention are first concentrated using a commercially available protein concentration filter, for example, an Amicon or Millipore Pellicon ultrafiltration unit. Following the concentration step, the concentrate may be applied to a suitable purification matrix. For example, a suitable affinity matrix may comprise a ligand for the peptide, a lectin or antibody molecule bound to a suitable support. Alternatively, an anion-exchange resin may be employed, for example, a matrix or substrate having pendant DEAE groups. Suitable matrices include acrylamide, agarose, dextran, cellulose, or other types commonly employed in protein purification. Alternatively, a cation-exchange step may be employed. Suitable cation exchangers include various insoluble matrices comprising sulfopropyl or carboxymethyl groups. Sulfopropyl groups are particularly preferred.

[0426] Other methods of use in purification include size exclusion chromatography (SEC), hydroxyapatite chromatography, hydrophobic interaction chromatography and chromatography on Blue Sepharose. These and other useful methods are illustrated in co-assigned U.S. Provisional Patent No. (Attorney Docket No. 40853-01-5168-P1, filed May 6, 2005).

[0427] One or more RP-HPLC steps employing hydrophobic RP-HPLC media, *e.g.*, silica gel having pendant methyl or other aliphatic groups, may be employed to further purify a polypeptide conjugate composition. Some or all of the foregoing purification steps, in various combinations, can also be employed to provide a homogeneous or essentially homogeneous modified glycoprotein.

[0428] The modified glycopeptide of the invention resulting from a large-scale fermentation may be purified by methods analogous to those disclosed by Urdal *et al., J. Chromatog.* **296**: 171 (1984). This reference describes two sequential, RP-HPLC steps for purification of recombinant human IL-2 on a preparative HPLC column. Alternatively, techniques such as affinity chromatography may be utilized to purify the modified glycoprotein.

[0429] In an exemplary embodiment, the purification is accomplished by the methods set forth in commonly owned, co-assigned U.S. Provisional Patent No. 60/665,588, filed March 24, 2005.

[0430] According to the present invention, pegylated peptides, *e.g.,* Factor VII, Factor VIIa peptide or peptide conjugate produced either via sequential de-sialylation or simultaneous sialylation can be purified or resolved by using magnesium chloride gradient.

[0431] In an exemplary embodiment, the Factor VII/Factor VIIa peptide conjugates can be separated into a light chain and a heavy chain, and one chain can be purified away from the other. In another exemplary embodiment, a product is obtained in which at least 80% of the Factor VII/Factor VIIa peptide conjugate in the product is the light chain portion of the Factor VII/Factor VIIa peptide conjugate. In another exemplary embodiment, a product is obtained in which at least 90% of the Factor VII/Factor VIIa peptide conjugate in the product is the light chain portion of the Factor VII/Factor VIIa peptide conjugate. In another exemplary embodiment, a product is obtained in which at least 95% of the Factor VII/Factor VIIa peptide conjugate in the product is the light chain portion of the Factor VII/Factor VIIa peptide conjugate. In another exemplary embodiment, a product is obtained in which essentially all of the Factor VII/Factor VIIa peptide conjugate in the product is the light chain portion of the Factor VII/Factor VIIa peptide conjugate. This product is possible for any compound of the invention.

[0432] In another exemplary embodiment, a product is obtained in which at least 80% of the Factor VII/Factor VIIa peptide conjugate in the product is the heavy chain portion of the Factor VII/Factor VIIa peptide conjugate. In another exemplary embodiment, a product is obtained in which at least 90% of the Factor VII/Factor VIIa peptide conjugate in the product is the heavy chain portion of the Factor VII/Factor VIIa peptide conjugate. In another exemplary embodiment, a product is obtained in which at least 95% of the Factor VII/Factor VIIa peptide conjugate in the product is the heavy chain portion of the Factor VII/Factor VIIa

peptide conjugate. In another exemplary embodiment, a product is obtained in which essentially all of the Factor VII/Factor VIIa peptide conjugate in the product is the heavy chain portion of the Factor VII/Factor VIIa peptide conjugate. This product is possible for any compound of the invention.

III. F. Properties of Factor VII/Factor VIIa Conjugates

[0433] In an exemplary embodiment, the Factor VII/Factor VIIa peptide conjugates of the invention possess essentially the same biochemical properties (e.g. clotting) as a native Factor VII/Factor VIIa peptide. In an exemplary embodiment, the Factor VII/Factor VIIa peptide conjugates of the invention possess reduced, or enhanced biochemical properties (e.g. clotting) over a native Factor VII/Factor VIIa peptide depending on the site of PEGylation, the size of the PEG added and the number of PEGs added.

[0434] Factor VII/Factor VIIa peptide conjugates are involved in the blood clotting process. In an exemplary embodiment, Factor VII/Factor VIIa peptide conjugates retain about 20%, or about 25%, or about 30%, or about 35%, or about 40%, or about 45%, or about 50%, or about 55%, or about 60%, or about 65%, or about 70%, or about 75%, or about 80%, or about 85%, or about 90%, or about 95% of the clotting activity of native Factor VII/Factor VIIa.

[0435] Factor VII/Factor VIIa peptide conjugates posssess amidolytic activity. In an exemplary embodiment, Factor VII/Factor VIIa peptide conjugates retain about 20%, or about 25%, or about 30%, or about 35%, or about 40%, or about 45%, or about 50%, or about 55%, or about 60%, or about 65%, or about 70%, or about 75%, or about 80%, or about 85%, or about 90%, or about 95% of the amidolytic activity of native Factor VII/Factor VIIa.

[0436] Factor VII/Factor VIIa peptide conjugates are able to convert Factor X to Factor Xa. In an exemplary embodiment, Factor VII/Factor VIIa peptide conjugates retain about 20%, or about 25%, or about 30%, or about 35%, or about 40%, or about 45%, or about 50%, or about 55%, or about 60%, or about 65%, or about 70%, or about 75%, or about 80%, or about 85%, or about 90%, or about 95% of the Factor X conversion activity of native Factor VII/Factor VIIa.

IV. Pharmaceutical Compositions

[0437] In another aspect, the invention provides a pharmaceutical composition. The pharmaceutical composition includes a pharmaceutically acceptable diluent and a covalent

conjugate between a non-naturally-occurring, PEG moiety, therapeutic moiety or biomolecule and a glycosylated or non-glycosylated peptide. The polymer, therapeutic moiety or biomolecule is conjugated to the peptide via an intact glycosyl linking group interposed between and covalently linked to both the peptide and the polymer, therapeutic moiety or biomolecule.

[0438] Pharmaceutical compositions of the invention are suitable for use in a variety of drug delivery systems. Suitable formulations for use in the present invention are found in *Remington's Pharmaceutical Sciences*, Mace Publishing Company, Philadelphia, PA, 17th ed. (1985). For a brief review of methods for drug delivery, *see,* Langer, *Science* **249**:1527-1533 (1990).

[0439] In an exemplary embodiment, the pharmaceutical formulation comprises a Factor VII/Factor VIIa peptide conjugate and a pharmaceutically acceptable diluent which is a member selected from sodium chloride, calcium chloride dihydrate, glycylglycine, polysorbate 80, and mannitol. In another exemplary embodiment, the pharmaceutically acceptable diluent is sodium chloride and glycylglycine. In another exemplary embodiment, the pharmaceutically acceptable diluent is calcium chloride dihydrate and polysorbate 80. In another exemplary embodiment, the pharmaceutically acceptable diluent is mannitol.

[0440] The pharmaceutical compositions may be formulated for any appropriate manner of administration, including for example, topical, oral, nasal, intravenous, intracranial, intraperitoneal, subcutaneous or intramuscular administration. For parenteral administration, such as subcutaneous injection, the carrier preferably comprises water, saline, alcohol, a fat, a wax or a buffer. For oral administration, any of the above carriers or a solid carrier, such as mannitol, lactose, starch, magnesium stearate, sodium saccharine, talcum, cellulose, glucose, sucrose, and magnesium carbonate, may be employed. Biodegradable microspheres (*e.g.*, polylactate polyglycolate) may also be employed as carriers for the pharmaceutical compositions of this invention. Suitable biodegradable microspheres are disclosed, for example, in U.S. Patent Nos. 4,897,268 and 5,075,109.

[0441] Commonly, the pharmaceutical compositions are administered parenterally, *e.g.*, intravenously. Thus, the invention provides compositions for parenteral administration that include the compound dissolved or suspended in an acceptable carrier, preferably an aqueous carrier, *e.g.*, water, buffered water, saline, PBS and the like. The compositions may contain pharmaceutically acceptable auxiliary substances as required to approximate physiological

conditions, such as pH adjusting and buffering agents, tonicity adjusting agents, wetting agents, detergents and the like.

[0442] These compositions may be sterilized by conventional sterilization techniques, or may be sterile filtered. The resulting aqueous solutions may be packaged for use as is, or lyophilized, the lyophilized preparation being combined with a sterile aqueous carrier prior to administration. The pH of the preparations typically will be between 3 and 11, more preferably from 5 to 9 and most preferably from 7 and 8.

[0443] In some embodiments the glycopeptides of the invention can be incorporated into liposomes formed from standard vesicle-forming lipids. A variety of methods are available for preparing liposomes, as described in, *e.g.*, Szoka *et al.*, *Ann. Rev. Biophys. Bioeng.* **9**: 467 (1980), U.S. Pat. Nos. 4,235,871, 4,501,728 and 4,837,028. The targeting of liposomes using a variety of targeting agents (*e.g.*, the sialyl galactosides of the invention) is well known in the art (*see, e.g.*, U.S. Patent Nos. 4,957,773 and 4,603,044).

[0444] Standard methods for coupling targeting agents to liposomes can be used. These methods generally involve incorporation into liposomes of lipid components, such as phosphatidylethanolamine, which can be activated for attachment of targeting agents, or derivatized lipophilic compounds, such as lipid-derivatized glycopeptides of the invention.

[0445] Targeting mechanisms generally require that the targeting agents be positioned on the surface of the liposome in such a manner that the target moieties are available for interaction with the target, for example, a cell surface receptor. The carbohydrates of the invention may be attached to a lipid molecule before the liposome is formed using methods known to those of skill in the art (*e.g.,* alkylation or acylation of a hydroxyl group present on the carbohydrate with a long chain alkyl halide or with a fatty acid, respectively). Alternatively, the liposome may be fashioned in such a way that a connector portion is first incorporated into the membrane at the time of forming the membrane. The connector portion must have a lipophilic portion, which is firmly embedded and anchored in the membrane. It must also have a reactive portion, which is chemically available on the aqueous surface of the liposome. The reactive portion is selected so that it will be chemically suitable to form a stable chemical bond with the targeting agent or carbohydrate, which is added later. In some cases it is possible to attach the target agent to the connector molecule directly, but in most instances it is more suitable to use a third molecule to act as a chemical bridge, thus linking

the connector molecule which is in the membrane with the target agent or carbohydrate which is extended, three dimensionally, off of the vesicle surface.

[0446] The compounds prepared by the methods of the invention may also find use as diagnostic reagents. For example, labeled compounds can be used to locate areas of inflammation or tumor metastasis in a patient suspected of having an inflammation. For this use, the compounds can be labeled with ^{125}I, ^{14}C, or tritium.

[0447] The active ingredient used in the pharmaceutical compositions of the present invention is Factor VII/Factor VIIa peptide conjugates having the biological properties of stimulating blood clot production. Preferably, the Factor VII/Factor VIIa peptide conjugate are administered parenterally (e.g. IV, IM, SC or IP). Effective dosages are expected to vary considerably depending on the condition being treated and the route of administration but are expected to be in the range of about 0.1 (~7U) to 100 (~7000U) μg/kg body weight of the active material. Preferable doses for treatment of anemic conditions are about 50 to about 300 Units/kg three times a week. Because the present invention provides a composition of matter comprising a Factor VII/Factor VIIa peptide with an enhanced in vivo residence time, the stated dosages are optionally lowered when a composition of the invention is administered.

[0448] Preparative methods for species of use in preparing the compositions of the invention are generally set forth in various patent publications, e.g., US 20040137557; WO 04/083258; and WO 04/033651. The following examples are provided to illustrate the conjugates, and methods and of the present invention, but not to limit the claimed invention.

EXAMPLES

EXAMPLE 1

Desialylation of Factor VIIa.

[0449] Factor VIIa which was expressed in serum-free media, Factor VIIa which was produced in serum containing media, plus three Factor VIIa mutants N145Q, N322Q, and analogue DVQ (V158D/E296V/M298Q).

[0450] In preparation for enzymatic desialylation, Factor VIIa was dialyzed into MES, 150 mM NaCl, 5 mM $CaCl_2$, 50mM MES, pH 6 overnight at 4°C in Snakeskin dialysis tubing with a MWCO of 10 KDa. Desialylation of Factor VIIa (1 mg/mL) was

performed with 10 U/L soluble sialidase from *Arthrobacter ureafaciens* (Calbiochem) at 32°C for 18 hours in the exchanged buffer.

EXAMPLE 2

Sialyl-PEGylation of Factor VIIa.

[0451] Sialyl-PEGylation ("GlycoPEGylation") was performed on asialo-Factor VIIa (1 mg/mL) with 100 U/L ST3Gal-III and 200 μM CMP-sialic acid-PEG (40 KDa, 20 KDa, 10 KDa, 5 KDa, and 2 KDa) at 32°C in the desialylation buffer for 2-6 hours. After the proper reaction time had expired, the PEGylated sample was immediately purified to minimize further GlycoPEGylation.

[0452] To cap GlycoPEGylated Factor VII/Factor VIIa with samples capped with sialic acid, the sialidase was first removed from the asialo-Factor VIIa by anion-exchange chromatography as indicated below. Excess CMP-sialic acid (5 mM) was added and incubated at 32°C for 2 hours, capping GlycoPEGylated Factor VIIa with sialic acid. The sialyl-PEGylated forms of Factor VIIa were analyzed by non-reducing SDS-PAGE (Tris-glycine gels and/or NuPAGE gels) and a Colloidal Blue Staining Kit, as described by Invitrogen.

EXAMPLE 3

Purification of PEGylated Factor VIIa.

[0453] GlycoPEGylated samples of Factor VIIa were purified with a modified anion-exchange method. Samples were handled at 5°C. Immediately before loading the column, 1 g Chelex 100 (BioRad) per 10 mL Factor VIIa solution was added to the remodeled sample. After stirring for 10 min, the suspension was filtered on a cellulose acetate membrane (0.2 μm) with a vacuum system. The retained chelator resin on the filter was washed once with 1-2 mL water per 10 mL bulk. The conductivity of the filtrate was adjusted to 10 mS/cm at 5°C, and adjusted to pH 8.6, if necessary.

[0454] Anion exchange was performed at 8-10°C. A column containing Q Sepharose FF was prepared before loading by washing with 1 M NaOH (10 column volumes), water (5 column volumes), 2 M NaCl, 50 mM HOAc, pH 3 (10 column volumes), and equilibrating with 175 mM NaCl, 10 mM glycylglycine, pH 8.6 (10 column volumes). For each PEGylation reaction, 15-20 mg Factor VIIa was loaded on to an XK16 column (Amersham Biosciences) with 10 mL Q Sepharose FF (no more than 2 mg protein per mL

resin) at a flow rate of 100 cm/h. For the 2 KDa linear PEG, 20 mg Factor VIIa was loaded on to an XK26 column (Amersham Biosciences) with 40 mL Q Sepharose FF (0.5 mg protein per mg resin) at a flow rate of 100 cm/h.

[0455] After loading, the column was washed with 175 mM NaCl, 10 mM glycylglycine, pH 8.6 10 column volumes) and 50 mM NaCl, 10 mM glycylglycine, pH 8.6 (2 column volumes). Elution was performed with a step gradient of 15 mM $CaCl_2$ by using 50 mM NaCl, 10 mM glycylglycine, 15 mM $CaCl_2$, pH 8.6 (5 column volumes). The column was then washed with 1 M NaCl, 10 mM glycylglycine, pH 8.6 (5 column volumes). The effluent was monitored by absorbance at 280 nm. Fractions (5 mL) were collected during the flow-through and the two washes; 2.5 mL fractions were collected during the $CaCl_2$ and 1M salt elutions. Fractions containing Factor VIIa were analyzed by non-reducing SDS-PAGE (Tris-glycine gels and/or NUPAGE gels) and a Colloidal Blue Staining Kit. The appropriate fractions with Factor VIIa were pooled, and the pH was adjusted to 7.2 with 4 M HCl.

[0456] Factor VIIa-SA-PEG-10KDa was purified as described above, except for the following changes. EDTA (10 mM) was added to to the PEGylated Factor VIIa solution, the pH was adjusted to pH 6, and the conductivity was adjusted to 5mS/cm, at 5°C. About 20 mg of Factor VIIa-SA-PEG-10KDa was loaded on to an XK16 column (Amersham Biosciences) with 10 mL Poros 50 Micron HQ resin (no more than 2 mg protein per mL, resin) at a flow rate of 100 cm/h. After loading, the column was washed with 175 mM NaCl, 10 mM histidine pH 6 (10 column volumes) and 50 mM NaCl, 10 mM histidine, pH 6 (2 column volumes). Elution was performed with a step gradient of 20 mM $CaCl_2$ in 50 mM NaCl, 10 mM histidine, pH 6 (5 column volumes). The column was then washed with 1 M NaCl, 10 mM histidine, pH 6 (5 column volumes).

[0457] The anion-exchange eluate containing Factor VIIa-SA-PEG-10KDa (25mL) was concentrated to 5-7 mL by using an Amicon Ultra-15 10K centrifugal filter device, according to the manufacturer's directions (Millipore). Following concentration, size exclusion chromatography was performed. The sample (5-7 mL) was loaded onto a column containing Superdex 200 (HiLoad 16/60, prep grade; Amersham Biosciences) equilibrated in 50 mM NaCl, 10 mM glycylglycine, 15 mM $CaCl_2$, pH 7.2 for most of the PEGylated variants. Factor VIIa-SA-PEG-10KDa was separated from the unmodified, asialo-Factor VIIa at a flow rate of 1 mL/min, and the absorbance was monitored at 280

nm. Fractions (1 mL) containing Factor VIIa were collected and analyzed by non-reducing SDS-PAGE (Tris-glycine gels and/or NuPAGE gels) and a Colloidal Blue Staining Kit. Fractions containing the targeted PEGylated isoform and devoid of the unmodified, asialo-Factor VIIa were pooled and concentrated to 1 mg/mL using an Amicon Ultra-15 10K centrifugal filter device. Protein concentration was determined from absorbance readings at 280 nm using an extinction coefficient of 1.37 $(mg/mL)^{-1}cm^{-1}$.

EXAMPLE 4

Determination of PEGylated Isoforms by Reversed phase HPLC analysis.

[0458] PEGylated Factor VIIa was analyzed by HPLC on a reversed-phase column (Zorbax 300SB-C3, 5 µm particle size, 2.1 x 150 mm). The eluants were A) 0.1 TFA in water and B) 0.09 % TFA in acetonitrile. Detection was at 214 nm. The gradient, flow rate, and column temperature depended on the PEG length (40 KDa, 20 KDa, and 10 KDa PEG: 35-65 %B in 30 min, 0.5 mL/min, 45°C; 10 KDa PEG: 35-60 %B in 30 min, 0.5 mL/min, 45°C; 5 KDa: 40-50 %B in 40 min, 0.5 mL/min, 45°C; 2 KDa: 38-43 %B in 67 min, 0.6 mL/min, 55°C). The identity of each peak was assigned based on two or more of four different pieces of evidence: the known retention time of native Factor VIIa, the SDS-PAGE migration of the isolated peak, the MALDI-TOF mass spectrum of the isolated peak, and the orderly progression of the retention time of each peak with increasing number of attached PEG.

EXAMPLE 5

Determination of Site of PEG Attachment by Reversed-phase HPLC.

[0459] Factor VIIa and PEGylated Factor VIIa variants were reduced by mixing sample (10 µL at a concentration of 1 mg/mL) with reducing buffer (40 µL, 50 mM NaCl, 10 mM glycylglycine, 15 mM EDTA, 8 M urea, 20 mM DTT, pH 8.6) for 15 min at room temperature. Water (50 µL) was added and the sample cooled to 4°C until injected on the HPLC (< 12 hrs). The HPLC column, eluants, and detection were as described above for non-reduced samples. The flow rate was 0.5 mL/min and the gradient was 30-55 %B in 90 min, followed by a brief wash cycle up to 90 %B. The identity of each peak was assigned as described in Example 4.

EXAMPLE 6

Factor VIIa Clotting Assay.

[0460] PEGylated samples and standards were tested in duplicate, and were diluted in 100mM NaCl, 5mM $CaCl_2$. 0.1% BSA (wt/vol), 50mM Tris, pH 7.4. The standard and samples were assayed over a range from 0.1 to 10 ng/mL. Equal volumes of diluted standards and samples were mixed with Factor VIIa deficient plasma (Diagnostica Stago), and stored on ice for no greater than 4 hours before they were assayed.

[0461] Clotting times were measured with a STart4 coagulometer (Diagnostica Stago). The coagulometer measured the time elapsed until an *in vitro* clot was formed, as indicated by the stopping of the gentle back-and-forth movement of a magnetic ball in a sample cuvette.

[0462] Into each cuvette, one magnetic ball was deposited, plus 100 μL Factor VIIa sample/deficient plasma and 100 μL of a diluted rat brain cephalin solution (stored on ice for no greater than 4 hours). Each reagent was added with 5 seconds between each well, and the final mixture was incubated for 300 seconds at 37°C. Diluted rat brain cephalin (RBC) solution was made from 2 mL RBC stock solution (1 vial RBC stock, from Haemachem, plus 10 mL 150mM NaCl) and 4 mL 100mM NaCl, 5mM $CaCl_2$, 0.1% BSA (wt/vol), 50mM Tris, pH 7.4.

[0463] At 300 seconds, the assay was started by the addition of 100 μL of a pre-heated (37°C) solution of soluble tissue factor (2μg/mL; amino acids 1-209) in 100mM NaCl. 12.5mM $CaCl_2$, 0.1% BSA (wt/vol), 50mM Tris, pH 7.4. Again, this next solution was added with a 5 second interval between samples.

[0464] The clotting times from the diluted standards were used to generate a standard curve (log clot time versus log Factor VIIa concentration). The resulting linear regression from the curve was used to determine the relative clotting activities of PEGylated variants. PEGylated Factor VIIa variants were compared against an aliquotted stock of Factor VIIa.

EXAMPLE 7

GlycoPEGylation of Recombinant Factor VIIa produced in BHK cells

[0465] This example sets forth the PEGylation of recombinant Factor VIIa made in BHK cells.

[0466] *Preparation of Asialo-Factor VIIa.* Recombinant Factor VIIa was produced in BHK cells (baby hamster kidney cells). Factor VIIa (14.2 mg) was dissolved at 1 mg/mL in buffer solution (pH 7.4, 0.05 M Tris, 0.15 M NaCl, 0.001 M $CaCl_2$, 0.05% NaN_3) and was incubated with 300 mU/mL sialidase (*Vibrio cholera*)-agarose conjugate for 3 days at 32 °C. To monitor the reaction a small aliquot of the reaction was diluted with the appropriate buffer and an IEF gel performed according to Invitrogen procedures (Figure 157). The mixture was centrifuged at 3,500 rpm and the supernatant was collected. The resin was washed three times (3×2 mL) with the above buffer solution (pH 7.4, 0.05 M Tris, 0.15 M NaCl, 0.05% NaN_3) and the combined washes were concentrated in a Centricon-Plus-20. The remaining solution was buffer exchanged with 0.05 M Tris (pH 7.4), 0.15 M NaCl, 0.05% NaN_3 to a final volume of 14.4 mL.

[0467] *Preparation of Factor VIIa-SA-PEG-1KDa and Factor VIIa-SA-PEG-10KDa.* The desialylation of Factor VIIa solution was split into two equal 7.2 mL samples. To each sample was added either CMP-SA-PEG-1KDa (7.4 mg) or CMP-SA-PEG-10KDa (7.4 mg). ST3Gal3 (1.58U) was added to both tubes and the reaction mixtures were incubated at 32°C for 96 hrs. The reaction was monitored by SDS-PAGE gel using reagents and conditions described by Invitrogen. When the reaction was complete, the reaction mixture was purified using a Toso Haas TSK-Gel-3000 preparative column using PBS buffer (pH 7.1) and collecting fractions based on UV absorption. The combined fractions containing the product were concentrated at 4°C in Centricon-Plus-20 centrifugal filters (Millipore, Bedford, MA) and the concentrated solution reformulated to yield 1.97 mg (bicinchoninic acid protein assay, BCA assay, Sigma-Aldrich, St. Louis MO) of Factor VIIa-SA-PEG. The product of the reaction was analyzed using SDS-PAGE and IEF analysis according to the procedures and reagents supplied by Invitrogen. Samples were dialyzed against water and analyzed by MALDI-TOF. **FIG. 7** shows the MALDI results for native Factor VIIa. **FIG. 8** contains the MALDI results for Factor VIIa-SA-PEG-1KDa. **FIG. 9** contains the MALDI results for Factor VIIa-SA-PEG-10KDa. **FIG. 10** depicts the SDS-PAGE analysis of all of the reaction products, where a band for Factor VIIa-SA-PEG-10KDa is evident.

EXAMPLE 8

Factor VIIa-SA-PEG-10KDa: One Pot Method

[0468] Factor VIIa (5 mg diluted in the product formulation buffer to a final concentration of 1 mg/mL), CMP-SA-PEG-10KDa (10mM, 60 μL) and *A. niger* enzyme ST3Gal3 (33 U/L)

and 10 mM histidine, 50 mM NaCl, 20 mM $CaCl_2$ were combined in a reaction vessel along with either 10 U/L, 1 U/L, 0.5 U/L or 0.1 U/L of sialidase (CalBiochem). The ingredients were mixed and incubated at 32°C. Reaction progress was measured by analyzing aliquots at 30 minute intervals for the first four hours. An aliquot was then removed at the 20 hour timepoint and subjected to SDS-PAGE. Extent of PEGylation was determined by removing 1 mL at 1.5, 2.5 and 3.5 hour timepoint and purifying the sample on a Poros 50HQ column.

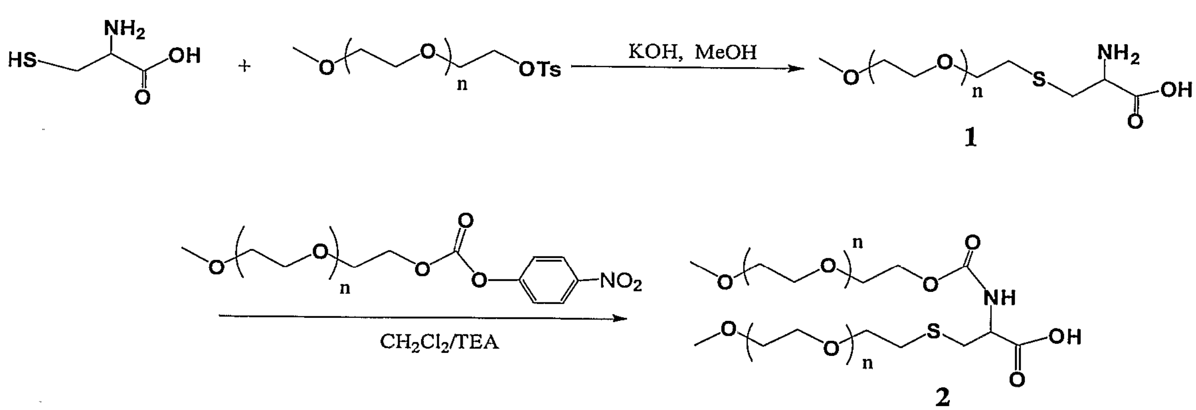
[0469] For the reaction conditions containing 10 U/L of sialidase, no appreciable amount of Factor VIIa-SA-PEG product was formed. For the reaction conditions containing 1 U/L of sialidase, about 17.6 % of the Factor VIIa in the reaction mixture was either mono or diPEGylated after 1.5 hours. This increased to 29% after 2.5 hours, and 40.3% after 3.5 hours. For the reaction conditions containing 0.5 U/L of sialidase, about 44.5 % of the Factor VIIa in the reaction mixture was either mono or diPEGylated after 3 hours, and 0.8% was triPEGylated or greater. After 20 hours, 69.4% was either mono or diPEGylated, and 18.3% was triPEGylated or greater.

[0470] For the reaction conditions containing 0.1 U/L of sialidase, about 29.6% of the Factor VIIa in the reaction mixture was either mono or diPEGylated after 3 hours. After 20 hours, 71.3% was either mono or diPEGylated, and 15.1% was triPEGylated or greater.

[0471] Results are shown in **FIG. 11** and **FIG. 12.**

EXAMPLE 9

Preparation of Cysteine-PEG$_2$ (2)



a. Synthesis of Compound 1

[0472] Potassium hydroxide (84.2 mg, 1.5 mmol, as a powder) was added to a solution of L-cysteine (93.7mg, 0.75 mmol) in anhydrous methanol (20 L) under argon. The mixture was stirred at room temperature for 30 min, and then mPEG-O-tosylate of molecular mass 20

kilodalton (Ts; 1.0 g, 0.05 mmol) was added in several portions over 2 hours. The mixture was stirred at room temperature for 5 days, and concentrated by rotary evaporation. The residue was diluted with water (30 mL), and stirred at room temperature for 2 hours to destroy any excess 20 kilodalton mPEG-O-tosylate. The solution was then neutralized with acetic acid, the pH adjusted to pH 5.0 and loaded onto a reverse phase chromatography (C-18 silica) column. The column was eluted with a gradient of methanol/water (the product elutes at about 70% methanol), product elution monitored by evaporative light scattering, and the appropriate fractions collected and diluted with water (500 mL). This solution was chromatographed (ion exchange, XK 50 Q, BIG Beads, 300 mL, hydroxide form; gradient of water to water/acetic acid-0.75N) and the pH of the appropriate fractions lowered to 6.0 with acetic acid. This solution was then captured on a reversed phase column (C-18 silica) and eluted with a gradient of methanol/water as described above. The product fractions were pooled, concentrated, redissolved in water and freeze-dried to afford 453 mg (44%) of a white solid (**1**).

[0473] Structural data for the compound were as follows: ^{1}H-NMR (500 MHz; D_2O) δ 2.83 (t, 2H, O-C-C$\underline{H}_2$-S), 3.05 (q, 1H, S-C$\underline{H}$H-CHN), 3.18 (q, 1H, (q, 1H, S-C$\underline{H}$H-CHN), 3.38 (s, 3H, C$\underline{H}_3$O), 3.7 (t, OC$\underline{H}_2$C$\underline{H}_2$O), 3.95 (q, 1H, C$\underline{H}$N). The purity of the product was confirmed by SDS PAGE.

b. Synthesis of Cysteine-PEG$_2$ (2)

[0474] Triethylamine (~0.5 mL) was added dropwise to a solution of compound 1 (440 mg, 22 μmol) dissolved in anhydrous CH_2Cl_2 (30 mL) until the solution was basic. A solution of 20 kilodalton mPEG-O-p-nitrophenyl carbonate (660 mg, 33 μmol) and N-hydroxysuccinimide (3.6 mg, 30.8 μmol) in CH_2Cl_2 (20 mL) was added in several portions over 1 hour at room temperature. The reaction mixture was stirred at room temperature for 24 hours. The solvent was then removed by rotary evaporation, the residue was dissolved in water (100 mL), and the pH adjusted to 9.5 with 1.0 N NaOH. The basic solution was stirred at room temperature for 2 hours and was then neutralized with acetic acid to a pH 7.0. The solution was then loaded onto a reversed phase chromatography (C-18 silica) column. The column was eluted with a gradient of methanol/water (the product elutes at about 70% methanol), product elution monitored by evaporative light scattering, and the appropriate fractions collected and diluted with water (500 mL). This solution was chromatographed (ion exchange, XK 50 Q, BIG Beads, 300 mL, hydroxide form; gradient of water to water/acetic acid-0.75N) and the pH of the appropriate fractions lowered to 6.0 with acetic acid. This

solution was then captured on a reversed phase column (C-18 silica) and eluted with a gradient of methanol/water as described above. The product fractions were pooled, concentrated, redissolved in water and freeze-dried to afford 575 mg (70 %) of a white solid (2).

[0475] Structural data for the compound were as follows: ^{1}H-NMR (500 MHz; D_2O) δ 2.83 (t, 2H, O-C-C$\underline{H}_2$-S), 2.95 (t, 2H, O-C-C$\underline{H}_2$-S), 3.12 (q, 1H, S-C$\underline{H}$H-CHN), 3.39 (s, 3H C$\underline{H}_3$O), 3.71 (t, OC$\underline{H}_2$C$\underline{H}_2$O). The purity of the product was confirmed by SDS PAGE.

EXAMPLE 10

Factor VIIa-SA-PEG-40KDa

[0476] *GlycoPEGylation of Factor VIIa (One Pot with Capping).* GlycoPEGylation of Factor VIIa was accomplished in a one-pot reaction where desialation and PEGylation occur simultaneously, followed by capping with sialic acid. The reaction was performed in a jacketed glass vessel controlled at 32°C by a recirculating waterbath. First, the concentrated 0.2μm-filtered Factor VIIa was introduced into the vessel and heated to 32°C by mixing with a stir bar for 20 minutes. A solution of sialidase was made from dry powder in 10mM histidine/50mM NaCl/20mM $CaCl_2$, pH 6.0 at a concentration of 4,000 U/L. Once the Factor VIIa reached 32°C, the sialidase was added to the Factor VIIa, and the reaction was mixed for approximately 5 minutes to ensure a uniform solution after time which the mixing was stopped. The desialation was allowed to proceed for 1.0 h at 32°C. During the desialation reaction, the CMP-SA-PEG-40KDa was dissolved into 10mM histidine/50mM NaCl/20mM $CaCl_2$, pH 6.0 buffer, and the concentration of was determined by UV absorbance at 271nm. After the CMP-SA-PEG-40KDa was dissolved, the CMP-SA-PEG-40KDa was added to the reaction, as well as the ST3Gal3, and the reaction was mixed for approximately 15 minutes with a stir bar to ensure a uniform solution. An additional volume of 85mL of buffer was added to make the reaction 1.0 L. The reaction was allowed to proceed without stirring for 24 hours before CMP-SA was added to a concentration of 4.3 mM to quench the reaction and cap the remaining terminal galactose residues with sialic acid. The quenching was allowed to proceed with mixing for 30 minutes at 32°C. The total volume of the reaction was 1.0 L before quenching. Timepoint samples (1 mL) were taken at 0, 4.5, 7.5, and 24 h, quenched with CMP-SA, and analyzed by RP-HPLC and SDS-PAGE.

[0477] *Purification of Factor VIIa-SA-PEG-40KDa.* After capping, the solution was diluted with 2.0 L of 10mM histidine, pH 6.0 that had been stored overnight at 4 °C and the

sample was filtered through a 0.2μm Millipak 60 filter. The resulting load volume was 3.1 L. The AEX2 chromatography was performed at 20-25°C (ambient room temperature) on an Akta Pilot system. After loading, a 10 column volumes wash with equilibration buffer was performed, and the product was eluted from the column using a 10 column volume gradient of $MgCl_2$ which resulted in resolution of PEGylated-Factor VIIa species from unPEGylated Factor VIIa. The loading for this column was intentionally kept low, targeting < 2 mg Factor VIIa/mL resin. SDS-PAGE gels were run in addition to RP-HPLC analysis of selected fractions and pools of fractions in order to make the pool of bulk product. Pooled fractions were pH adjusted to 6.0 with 1M NaOH and stored in the cold room at 2-8°C overnight.

[0478] *Final Concentration/Diafiltration, aseptic filtration and aliquoting.* The pooled fractions were filtered through a Millipak 20 0.2μm filter and stored overnight at 2-8°C. To perform the concentration/diafiltration, a Millipore 0.1m^2 30 KDa regenerated cellulose membrane was used in a system fitted with a peristaltic pump and silicone tubing. The system was assembled and flushed with water, then sanitized with 0.1M NaOH for at least 1 hour, and then stored in 0.1M NaOH until equilibration with 10 mM histidine/ 5 mM $CaCl_2$/ 100 mM NaCl pH 6.0 diafiltration buffer immediately before use. The product was concentrated to approximately 400 mL and then diafiltered at constant volume with approximately 5 diavolumes of buffer. The product was then concentrated to approximately 300mL and recovered after a low pressure recirculation for 5 minutes, and the membranes were rinsed with 200 mL of diafiltration buffer by a recirculation for 5 minutes. The wash was recovered with product, and another 50mL of buffer was recirculated for another 5 minutes for a final wash. The resulting bulk was approximately 510 mL, and that was filtered through a 1L vacuum filter fitted with a 0.2μm PES membrane (Millipore). The aseptically-filtered bulk was then aliquoted into 25mL aliquots in 50mL sterile falcon tubes and frozen at -80°C.

Analysis of the PEGylation reaction by HPLC (Example 10)

	Conjugation Reaction Time				Purification
	0 hrs	4.5 hrs	7.5 hrs	24 hrs	After Chromatography
% Unpegylated	94.7	76.1	66.6	51.0	0.6
% Monopegylated	0.9	17.9	26.1	39.1	85.6

% Dipegylated	0.1	0.9	1.9	5.1	5.1
% Tripegylated	0.0	0.0	0.0	0.2	0.2

After 24 hours, the bulk product PEG-state distribution was: 0.7% unpegylated, 85.3% mono-pegylated, 11.5% di-pegylated, and 0.3% tri-pegylated. Column chromatography is the main step in the process that generates the product distribution, largely through removing unpegylated material from mono- and di-pegylated species.

EXAMPLE 11

Factor VIIa-SA-PEG-10KDa

[0479] The following example describes a procedure for determining the number of modified sugar attachments to light and heavy chains of Factor VIIa-SA-PEG-10KDa by reverse phase HPLC.

[0480] Factor VIIa-SA-PEG-10KDa was subjected to reducing conditions in order to separate the heavy chain from the light chain. After separation, the heavy and light chains were subjected to separate reverse phase HPLC experiments. Peaks were assigned based on their position relative to the non-modified Factor VIIa peaks in the chromatograms of the native Factor VIIa control.

[0481] The following table describes the HPLC solvent gradient parameters for the light chain. The column temperature was 39°C.

HPLC Light Chain Solvent Gradient Parameters

Time, min	Solvent B, %	Flow rate, mL/min	Comment
0	30	0.5	Initial condition
60	47	0.5	Gradient elution
60.2	90	0.5	Start wash
70	90	0.5	Wash

[0482] The chromatograms of light chain Factor VIIa-SA-PEG-10KDa (top) and native light chain Factor VIIa (bottom) are provided in **FIG. 14A.**

[0483] The following table describes the HPLC solvent gradient parameters for the heavy chain. The column temperature was 52°C.

HPLC Heavy Chain Solvent Gradient Parameters

Time, min	Solvent B, %	Flow rate, ml/min	Comment
0	42.5	0.5	Initial condition
36	52.5	0.5	Gradient elution
36.1	90	0.5	Start wash
41	90	0.5	wash

[0484] The chromatograms of heavy chain Factor VIIa-SA-PEG-10KDa (top) and native heavy chain Factor VIIa (bottom) are provided in **FIG. 14B**.

EXAMPLE 12

Factor VIIa-SA-PEG-40KDa

[0485] The following example describes a procedure for determining the number of modified sugar attachments to light and heavy chains of Factor VIIa-SA-PEG-40KDa by reverse phase HPLC.

[0486] Factor VIIa-SA-PEG-40KDa was subjected to reducing conditions in order to separate the heavy chain from the light chain. After separation, the heavy and light chains were subjected to separate reverse phase HPLC experiments. Peaks were assigned based on their position relative to the non-modified sugar peaks in the chromatograms of the native Factor VIIa control.

[0487] The following table describes the HPLC solvent gradient parameters for the light chain. The column temperature was 25°C.

HPLC Light Chain Solvent Gradient Parameters

Time (min)	**Eluent B (%)**	**Comment**
0	30	Initial conditions
60	47	Gradient elution
60.5	90	Begin wash
65.5	90	End wash
66	42.5	Begin heavy chain method equilibration
70	42.5	End of Run

[0488] The chromatograms of light chain Factor VIIa-SA-PEG-40KDa (bottom) and native light chain Factor VIIa (top) are provided in **FIG. 15A**.

[0489] The following table describes the HPLC solvent gradient parameters for the heavy chain. The column temperature was 40°C.

HPLC Heavy Chain Solvent Gradient Parameters

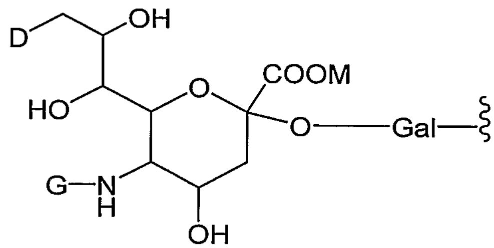
Time (min)	Eluent B (%)	Comment
0	42.5	Initial conditions
36	52.5	Gradient elution
36.5	90	Begin wash
41.5	90	End wash
42	30	Begin light chain method equilibration
47	30	End Run

[0490] The chromatograms of heavy chain Factor VIIa-SA-PEG-40KDa (bottom) and native heavy chain Factor VIIa (top) are provided in **FIG. 15B**.

[0491] It is understood that the examples and embodiments described herein are for illustrative purposes only and that various modifications or changes in light thereof will be suggested to persons skilled in the art and are to be included within the spirit and purview of this application and scope of the appended claims. All publications, patents, and patent applications cited herein are hereby incorporated by reference in their entirety for all purposes.

WHAT IS CLAIMED IS:

1. A method of making a Factor VII/Factor VIIa peptide conjugate comprising a glycosyl linker comprising a modified sialyl residue having the formula:



wherein

D is a member selected from -OH and R^1-L-HN-;

G is a member selected from R^1-L- and -C(O)(C_1-C_6)alkyl-R^1;

R^1 is a moiety comprising a member selected from a straight-chain poly(ethylene glycol) residue and branched poly(ethylene glycol) residue; and

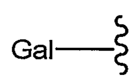
M is a member selected from H, a metal and a single negative charge;

L is a linker which is a member selected from a bond, substituted or unsubstituted alkyl and substituted or unsubstituted heteroalkyl,

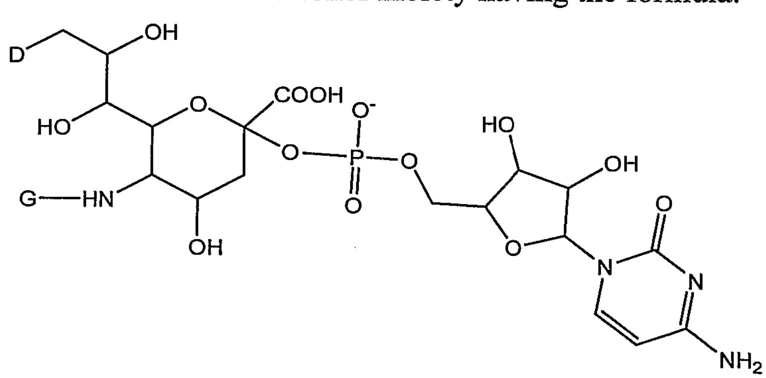
such that when D is OH, G is R^1-L-, and when G is –C(O)(C_1-C_6)alkyl, D is R^1-L-NH-

said method comprising:

(a) contacting a Factor VII/Factor VIIa peptide comprising the glycosyl moiety:

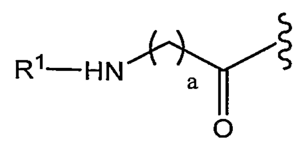


with a PEG-sialic acid donor moiety having the formula:



and an enzyme that transfers said PEG-sialic acid onto the Gal of said glycosyl moiety, under conditions appropriate for said transfer.

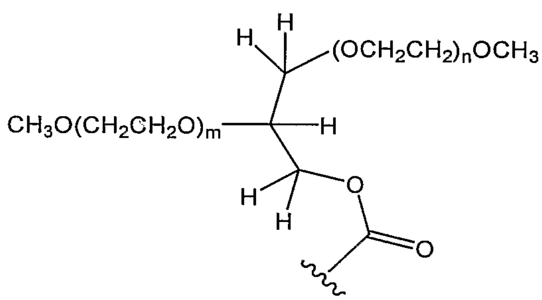
2. The method according to claim **1**, wherein L-R^1 has the formula:



wherein

a is an integer selected from 0 to 20.

3. The method according to claim **1**, wherein R^1 has a structure that is a member selected from:



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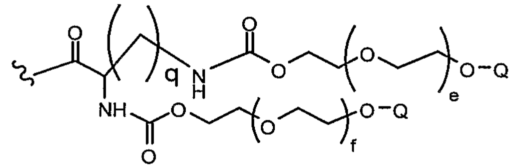

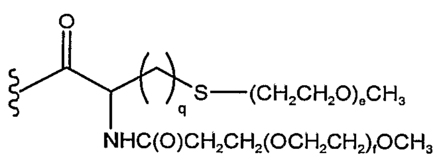

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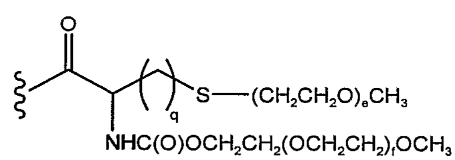
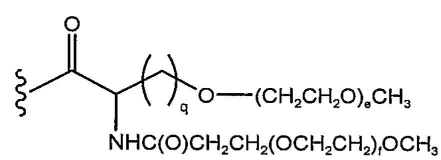
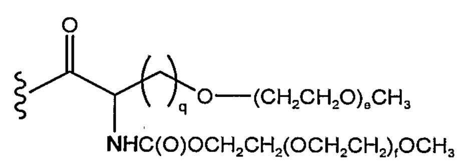




; and

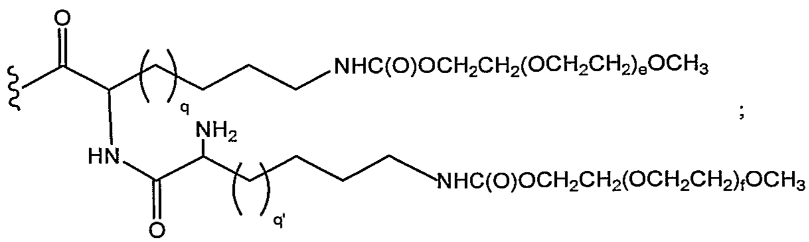


wherein

e, f, m and n are integers independently selected from 1 to 2500; and

q is an integer selected from 0 to 20.

4. The method according to claim **1**, wherein R^1 has a structure that is a member selected from:



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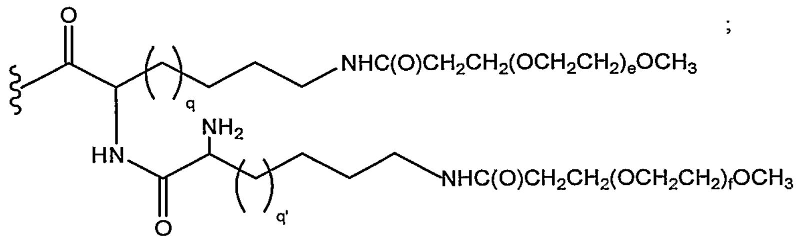

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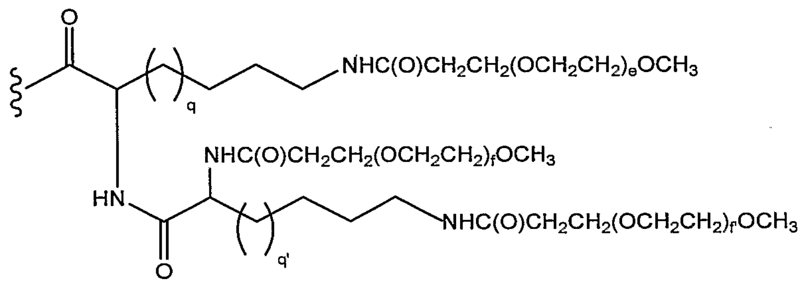
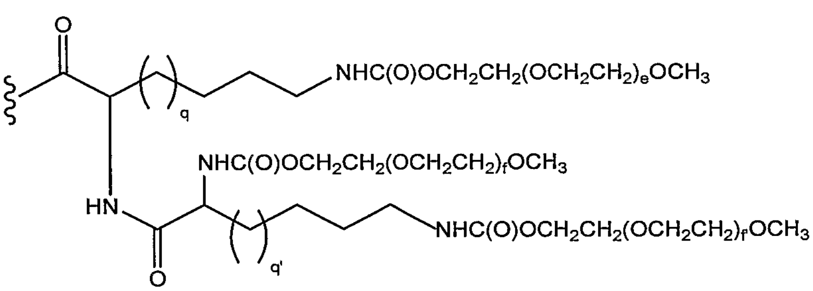


; and



wherein

e, f and f' are integers independently selected from 1 to 2500; and

q and q' are integers independently selected from 1 to 20.

5. The method according to claim **1**, wherein R^1 has a structure that is a member selected from:

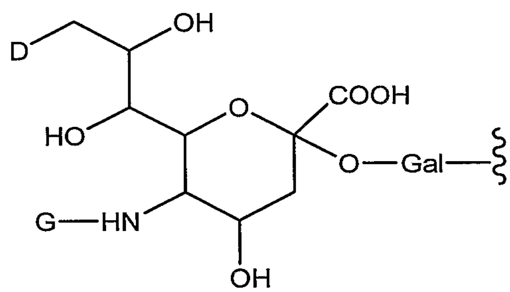
$$\text{—}C(O)CH_2CH_2(OCH_2CH_2)_eOCH_3$$; and

$$\text{—}C(O)OCH_2CH_2(OCH_2CH_2)_fOCH_3$$

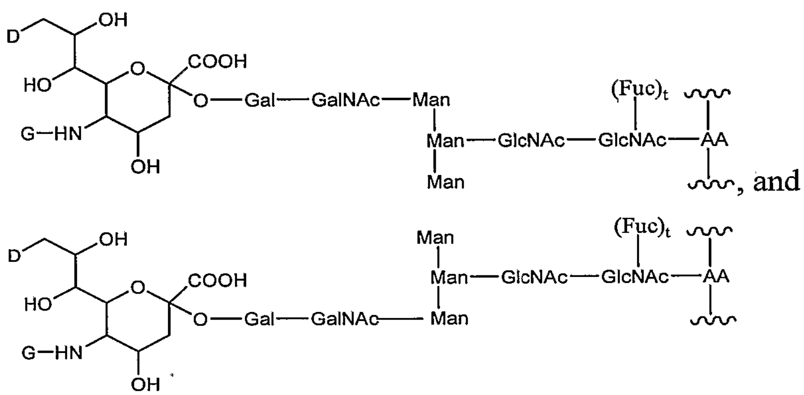
wherein

e and f are integers independently selected from 1 to 2500.

6. The method according to claim **1**, wherein said glycosyl linker has the formula:

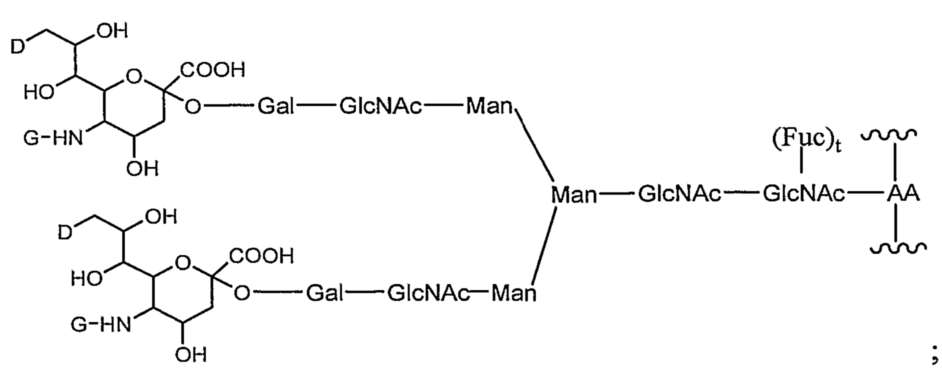


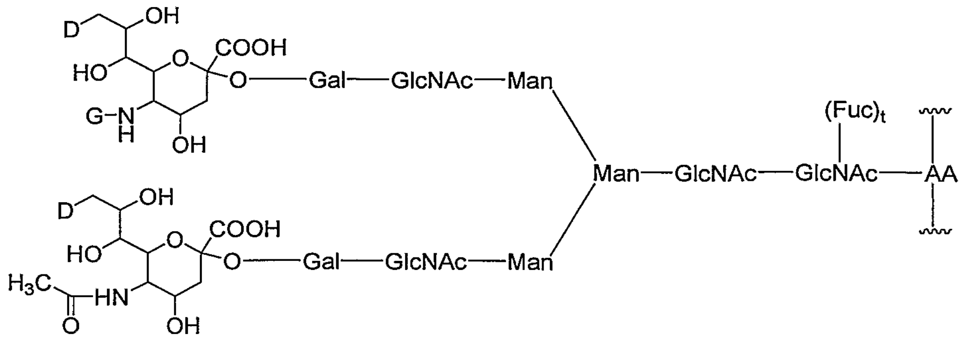
7. The method according to claim **1**, wherein said peptide conjugate comprises at least one of said glycosyl linker according to a formula selected from:



wherein AA is an amino acid residue of said peptide conjugate and t is an integer selected from 0 and 1.

8. The method according to claim **1** wherein said peptide conjugate comprises at least one of said glycosyl linker wherein each of said glycosyl linker has a structure which is a member independently selected from the following formulae:





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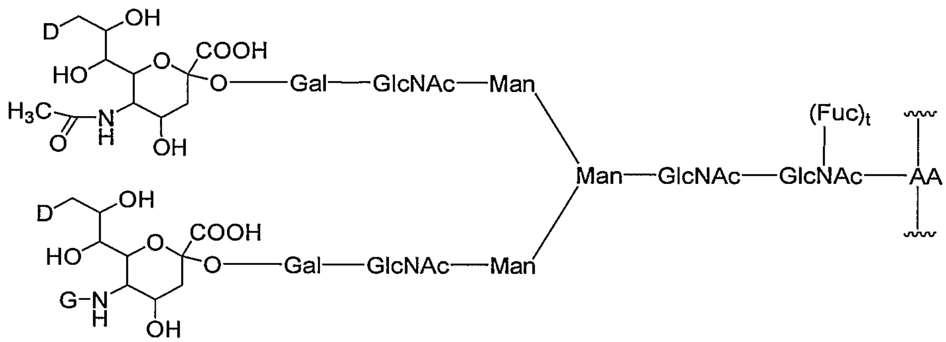

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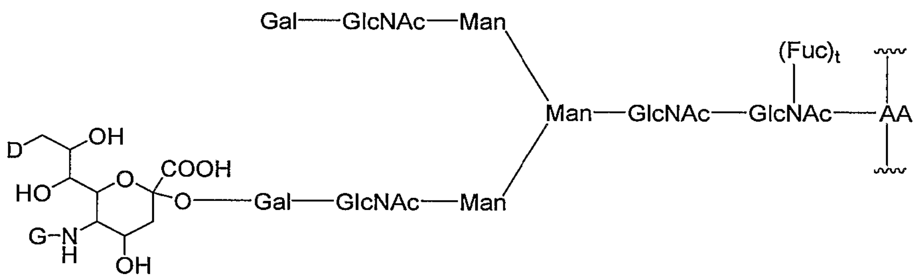

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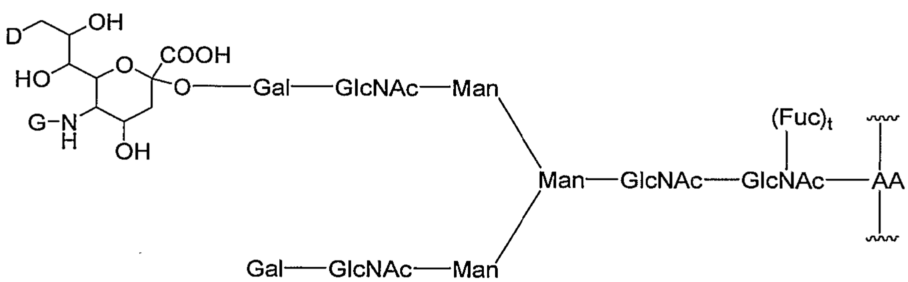
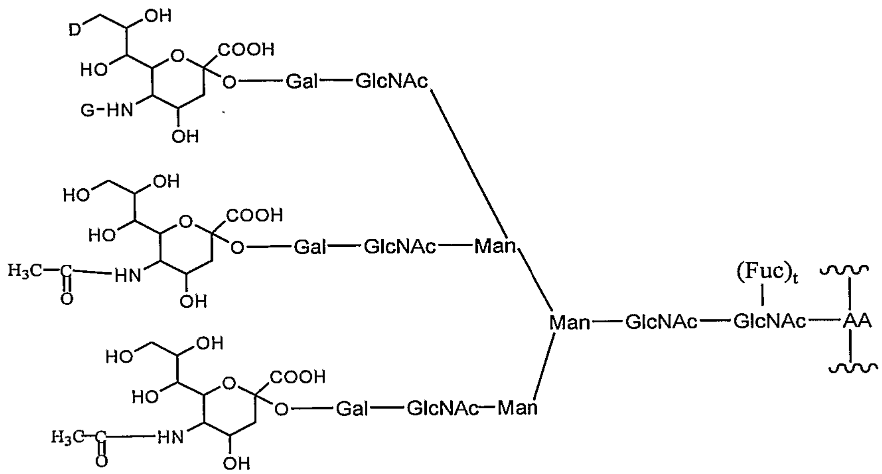
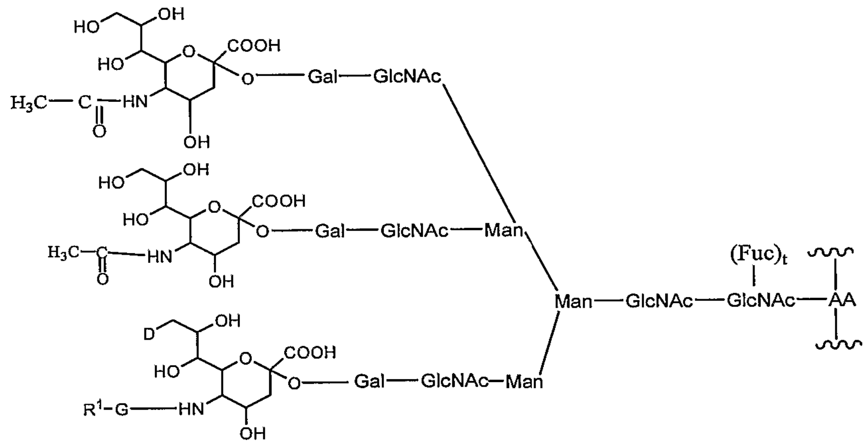


wherein AA is an amino acid residue of said peptide conjugate and t is an integer selected from 0 and 1.

9. The method of claim **1**, wherein said peptide conjugate comprises at least one of said glycosyl linker according to a formula selected from:





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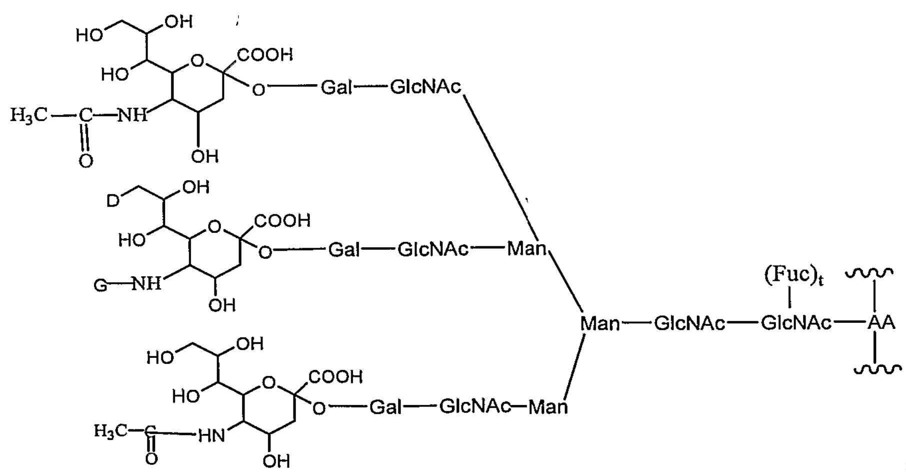
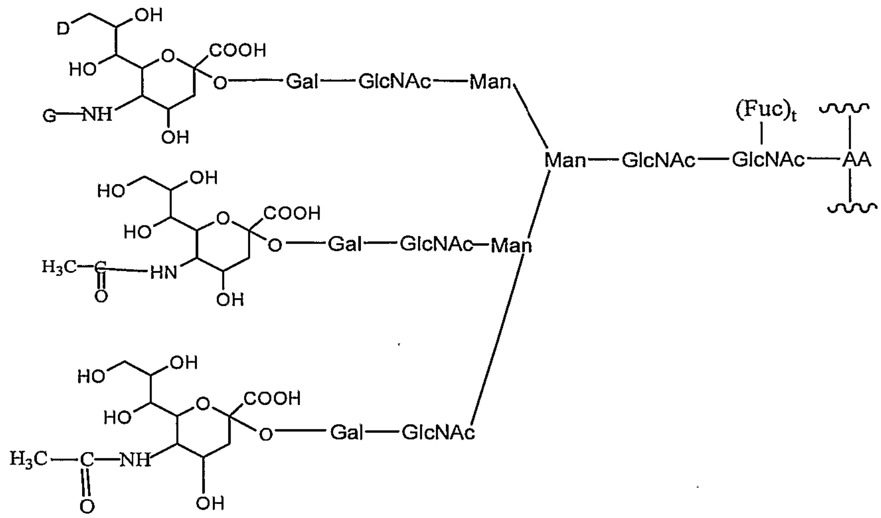


,



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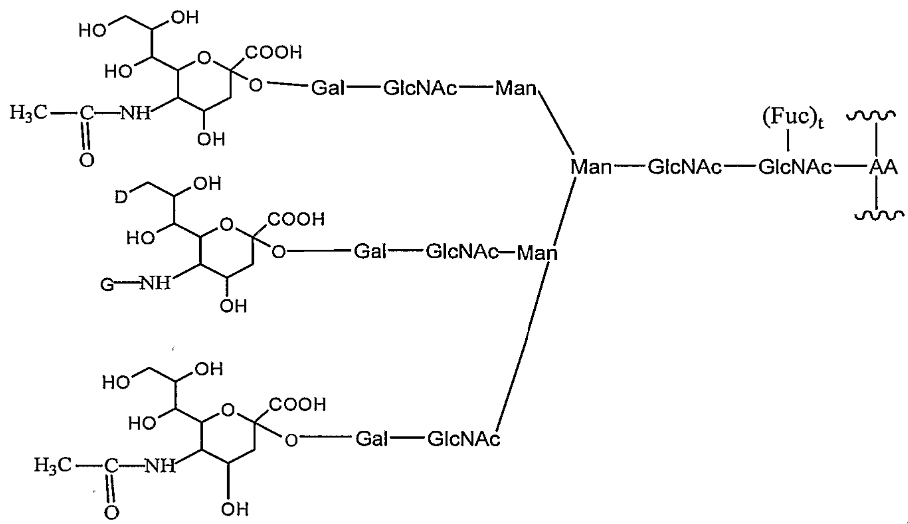
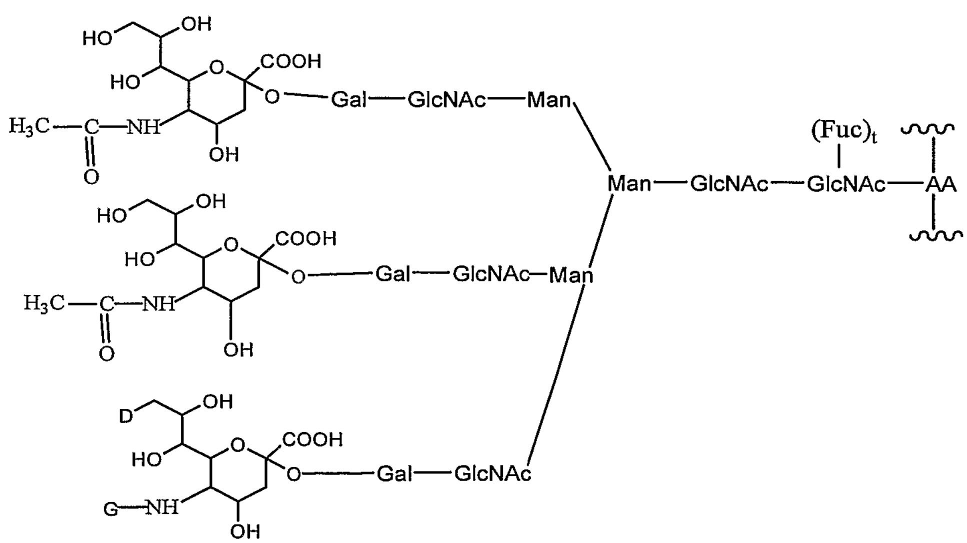


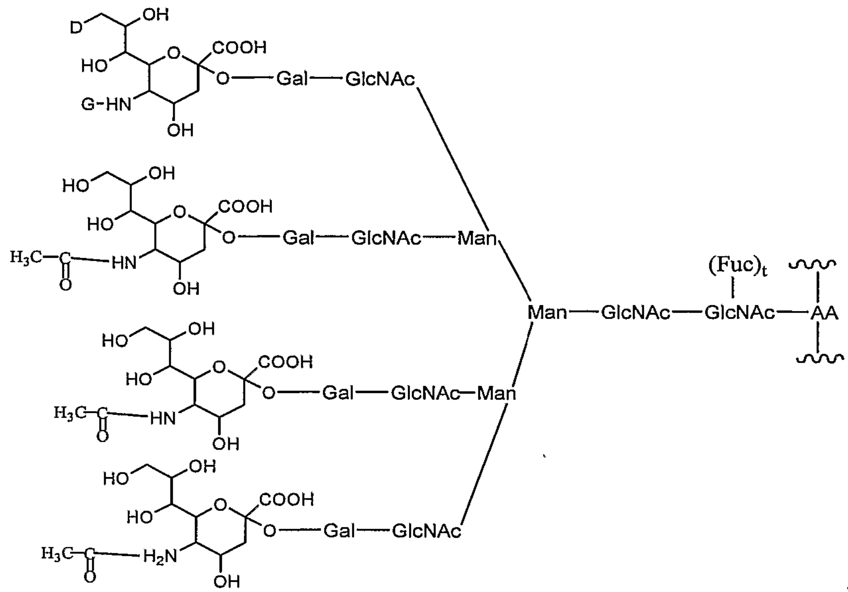
, and



wherein AA is an amino acid residue of said peptide conjugate and t is an integer selected from 0 and 1

and wherein a member selected from 0 and 2 of the sialyl moieties which do not comprise G are absent.

10. The method according to claim **1** wherein said peptide conjugate comprises at least one said glycosyl linker according to a formula selected from:



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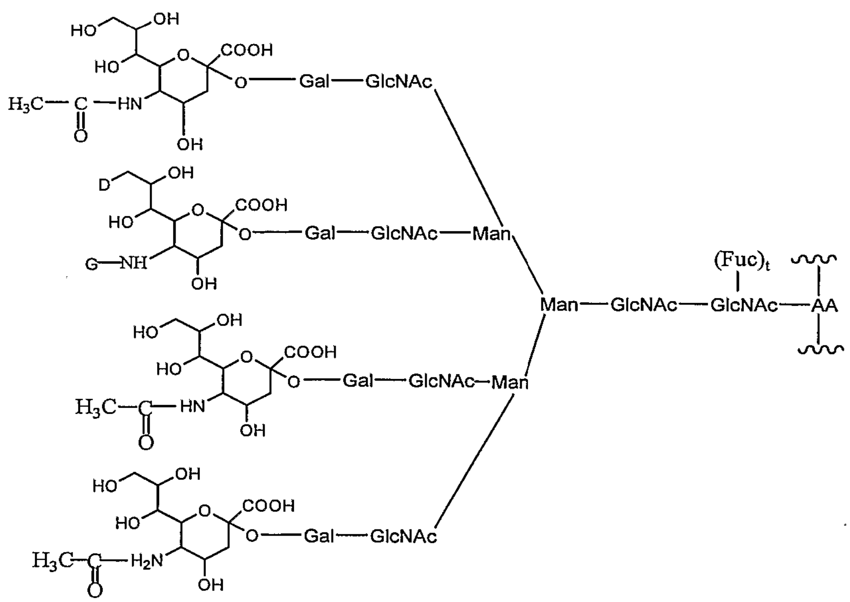

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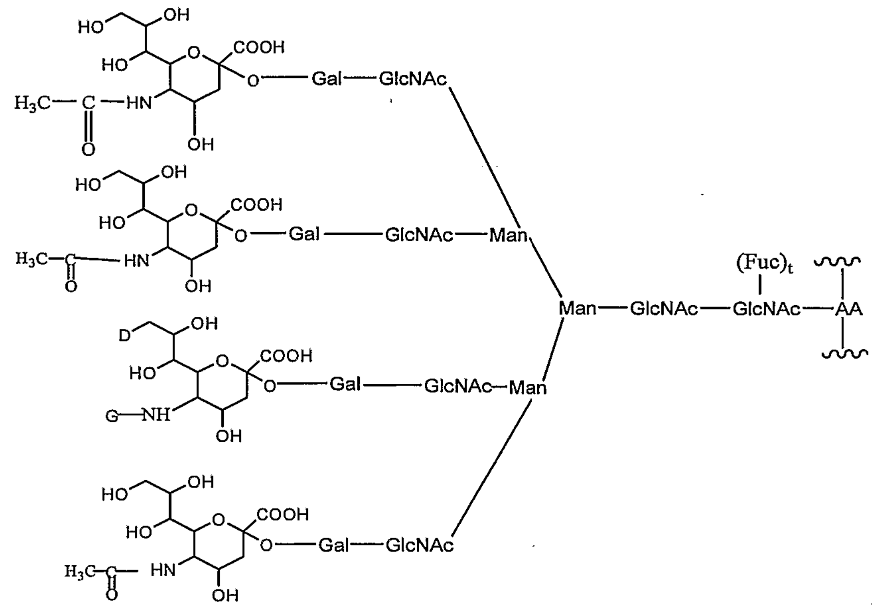
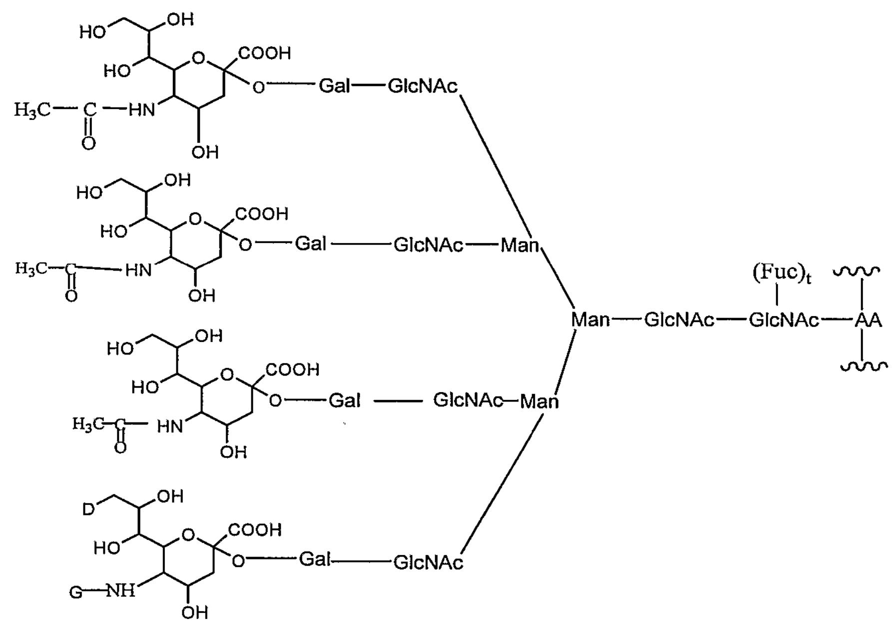


, and



wherein AA is an amino acid residue of said peptide conjugate and t is an integer selected from 0 and 1

and wherein a member selected from 0 and 2 of the sialyl moieties which do not comprise G are absent.

11. The method according to claim **1**, wherein said Factor VII/Factor VIIa peptide has the amino acid sequence of SEQ. ID. NO:1.

12. The method according to claim **1**, wherein said glycosyl linker is attached to said Factor VII/Factor VIIa peptide through an amino acid residue selected from serine and threonine.

13. The method according to claim **1**, wherein said glycosyl linker is attached to said Factor VII/Factor VIIa peptide through an amino acid residue which is an asparagine residue.

14. The method according to claim **13**, wherein said asparagine residue is a member selected from N152, N322 and combinations thereof.

15. The method according to claim **1**, wherein said Factor VIIa peptide is a bioactive Factor VIIa peptide.

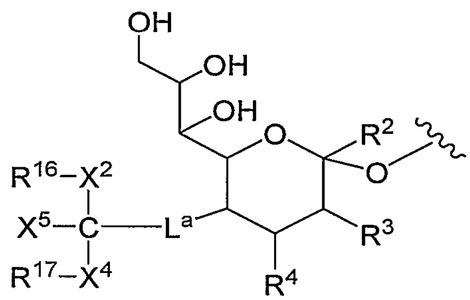
16. The method of claim **1**, further comprising, prior to step (a):
(b) expressing said Factor VII/Factor VIIa peptide in a suitable host.

17. The method of claim **16**, wherein said host is a mammalian expression system.

18. A method of treating a condition in a subject in need thereof, said condition characterized by compromised clotting potency in said subject, said method comprising the step of administering to the subject an amount of the Factor VII/Factor VIIa peptide conjugate produced according to the method of claim **1**, effective to ameliorate said condition in said subject.

19. A method of enhancing clotting potency in a mammal, said method comprising administering to said mammal an amount of the Factor VII/Factor VIIa peptide conjugate produced according to the method of claim **1**.

20. A method of making a Factor VII/Factor VIIa peptide conjugate comprising a glycosyl linker comprising a modified sialyl residue having the formula:



;

wherein

R^2 is H, CH_2OR^7, $COOR^7$ or OR^7

wherein

R^7 represents H, substituted or unsubstituted alkyl or substituted or unsubstituted heteroalkyl;

R^3 and R^4 are members independently selected from H, substituted or unsubstituted alkyl, OR^8, $NHC(O)R^9$

wherein

R^8 and R^9 are independently selected from H, substituted or unsubstituted alkyl, substituted or unsubstituted heteroalkyl or sialic acid;

R^{16} and R^{17} are independently selected polymeric arms;

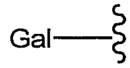
X^2 and X^4 are independently selected linkage fragments joining polymeric moieties R^{16} and R^{17} to C;

X^5 is a non-reactive group; and

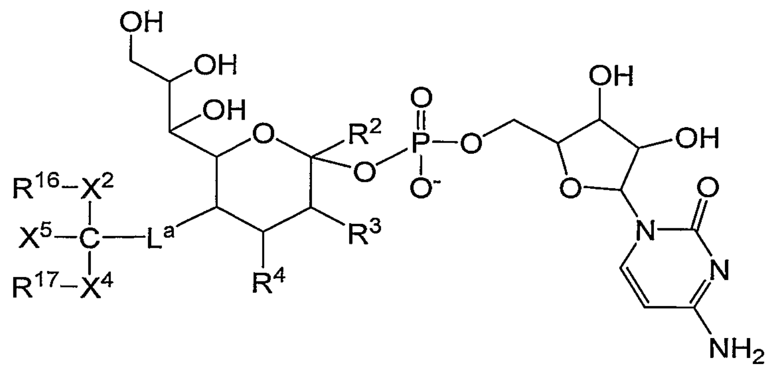
L^a is a linker group

said method comprising:

(a) contacting a Factor VII/FactorVIIa peptide comprising the glycosyl moiety:

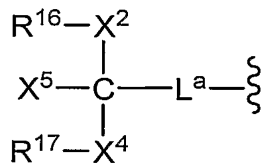


with a PEG-sialic acid donor moiety having the formula:

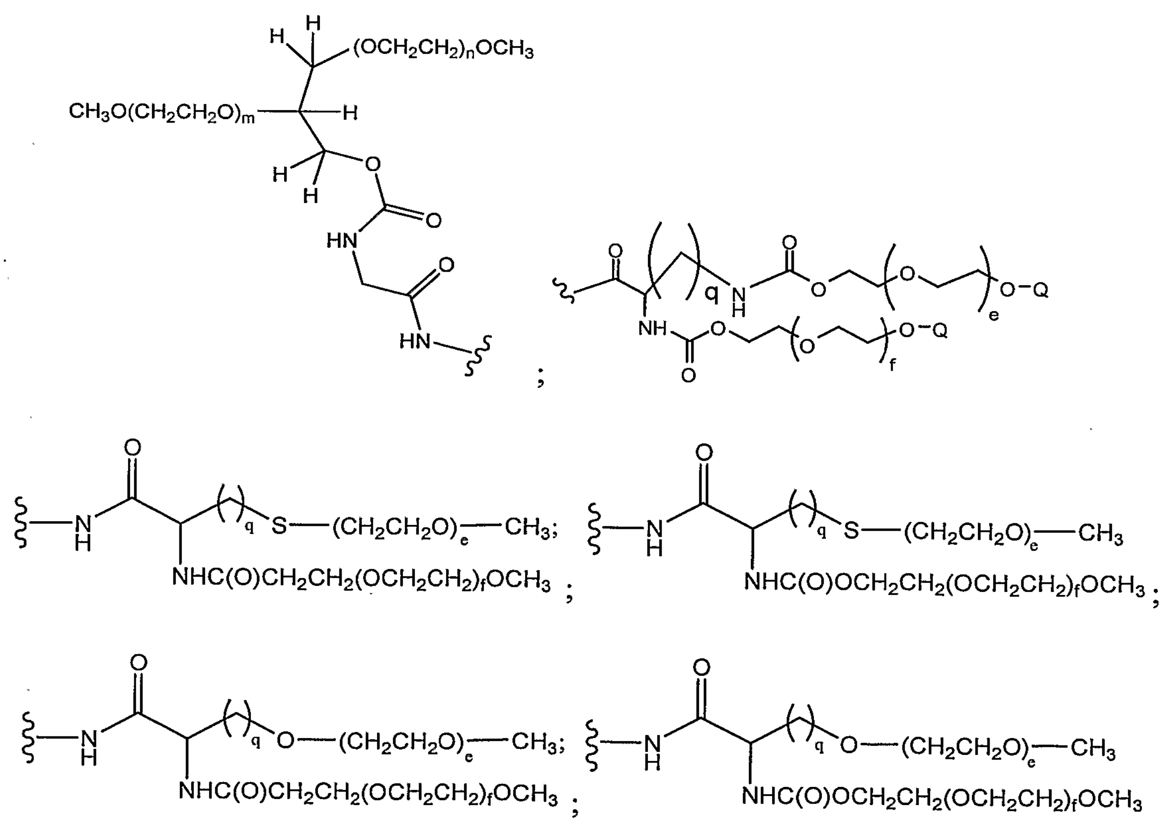


and an enzyme that transfers PEG-sialic acid onto the Gal of said glycosyl moiety, under conditions appropriate for said transfer.

21. The method according to claim **20**, wherein the moiety:



has a formula that is a member selected from:

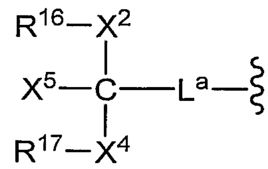


wherein

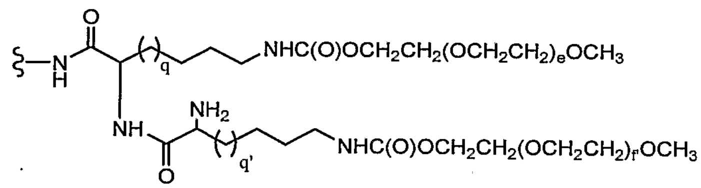
e, f, m and n are integers independently selected from 1 to 2500; and

q is an integer selected from 0 to 20.

22. The method according to claim **20**, wherein the moiety:



has a formula that is a member selected from:



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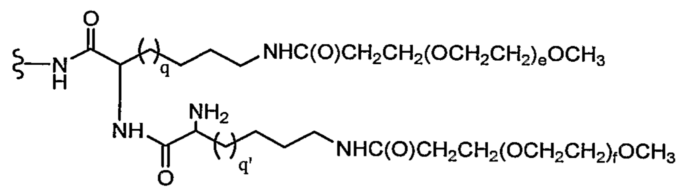

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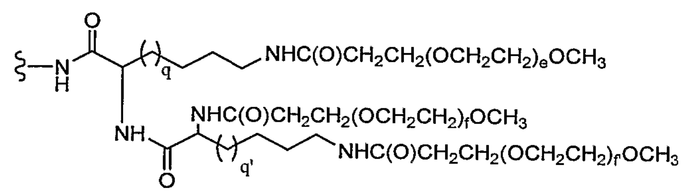
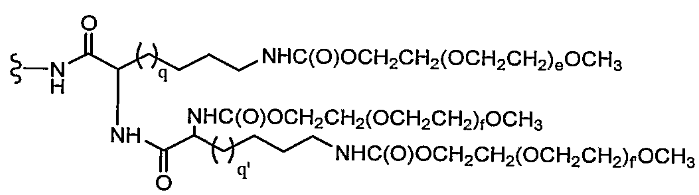


; and

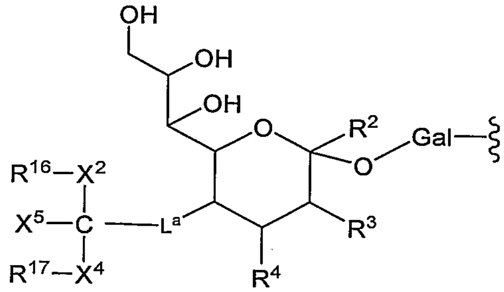


wherein

e, f and f' are integers independently selected from 1 to 2500; and

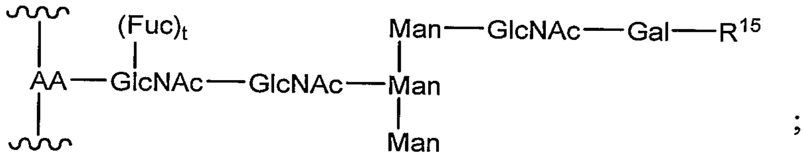
q and q' are integers independently selected from 1 to 20.

23. The method according to claim **20**, wherein said glycosyl linker comprises the formula:



.

24. The method according to claim **20**, wherein the Factor VII/Factor VIIa peptide conjugate comprises at least one glycosyl linker having the formula:



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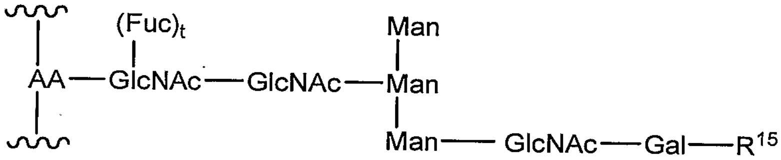

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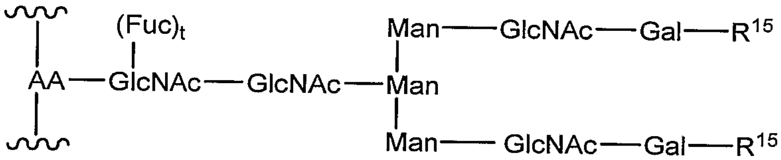

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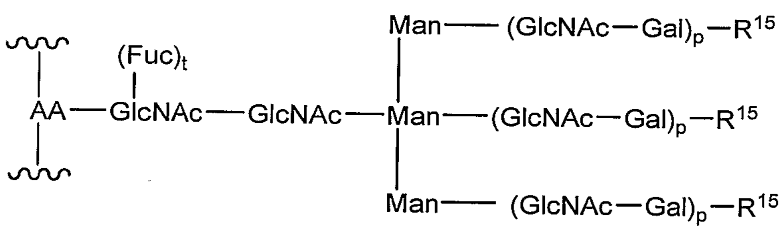
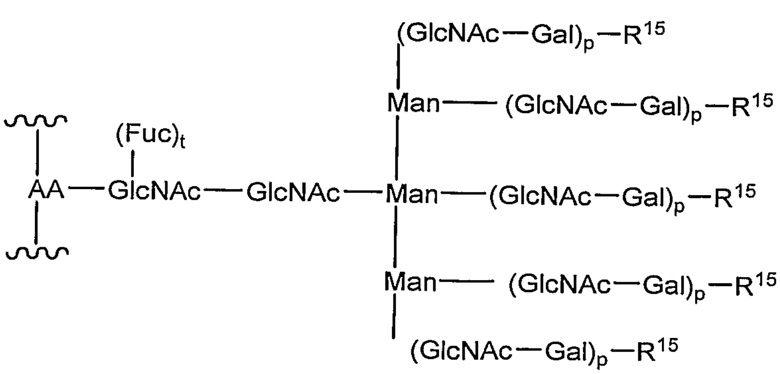


; and



wherein

AA is an amino acid residue of said peptide;

t is an integer selected from 0 and 1; and

R^{15} is the modified sialyl moiety.

25. The method according to claim **20**, wherein said Factor VII/Factor VIIa peptide has the amino acid sequence of SEQ. ID. NO:1.

26. The method according to claim **20**, wherein said glycosyl linker is attached to said Factor VII/Factor VIIa peptide through an amino acid residue which is an asparagine residue.

27. The method according to claim **26**, wherein said asparagine residue is a member selected from N152, N322 and combinations thereof.

28. The method according to claim **20**, wherein said Factor VIIa peptide is a bioactive Factor VIIa peptide.

29. The method of claim **20**, further comprising, prior to step (a): (b) expressing the Factor VII/Factor VIIa peptide in a suitable host.

30. The method of claim **29**, wherein said host is a mammalian expression system.

31. A method of treating a condition in a subject in need thereof, said condition characterized by compromised clotting potency in said subject, said method comprising the step of administering to the subject an amount of the Factor VII/Factor VIIa peptide conjugate produced according to the method of claim **20**, effective to ameliorate said condition in said subject.

32. A method of enhancing clotting potency in a mammal, said method comprising administering to said mammal an amount of the Factor VII/Factor VIIa peptide conjugate produced according to the method of claim **20**.

33. A method of synthesizing a Factor VII or Factor VIIa peptide conjugate, said method comprising combining

a) sialidase;

b) enzyme which is a member selected from glycosyltransferase, exoglycosidase and endoglycosidase

c) modified sugar/modified sialyl residue

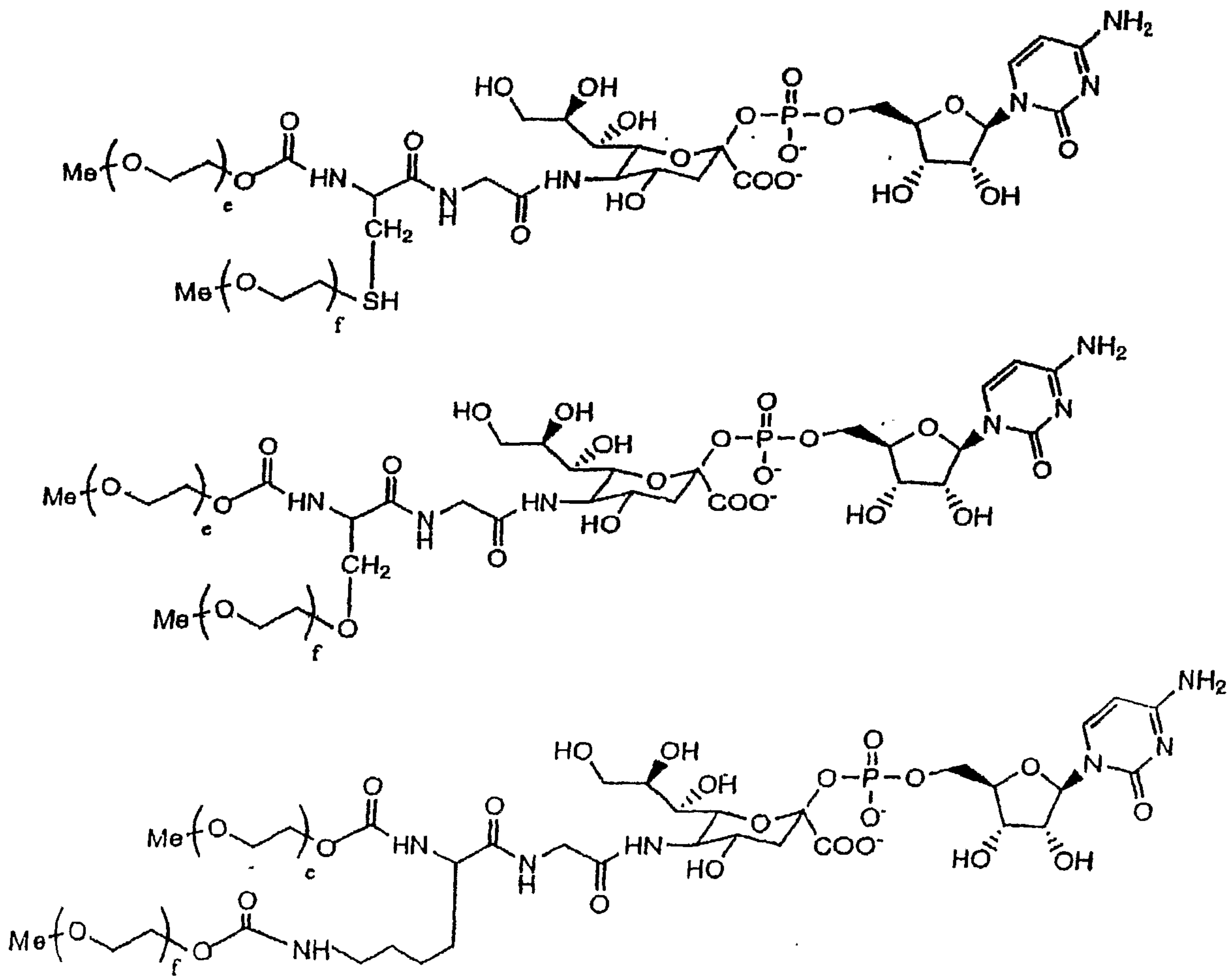
d) Factor VII/Factor VIIa peptide

thus synthesizing said Factor VII or Factor VIIa peptide conjugate.

34. The method of claim **33**, wherein said combining is for a time less than 10 hours.

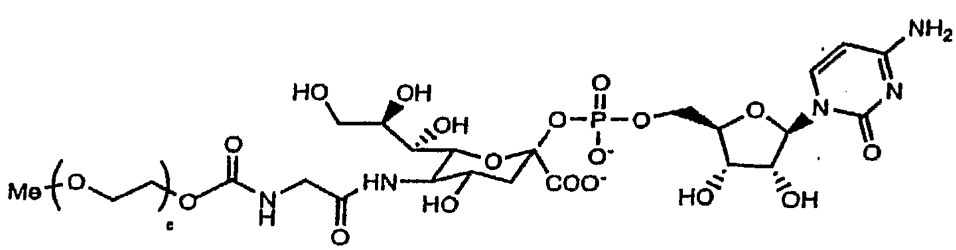
35. The method of claim **33**, further comprising a capping step.

FIGURE 1A



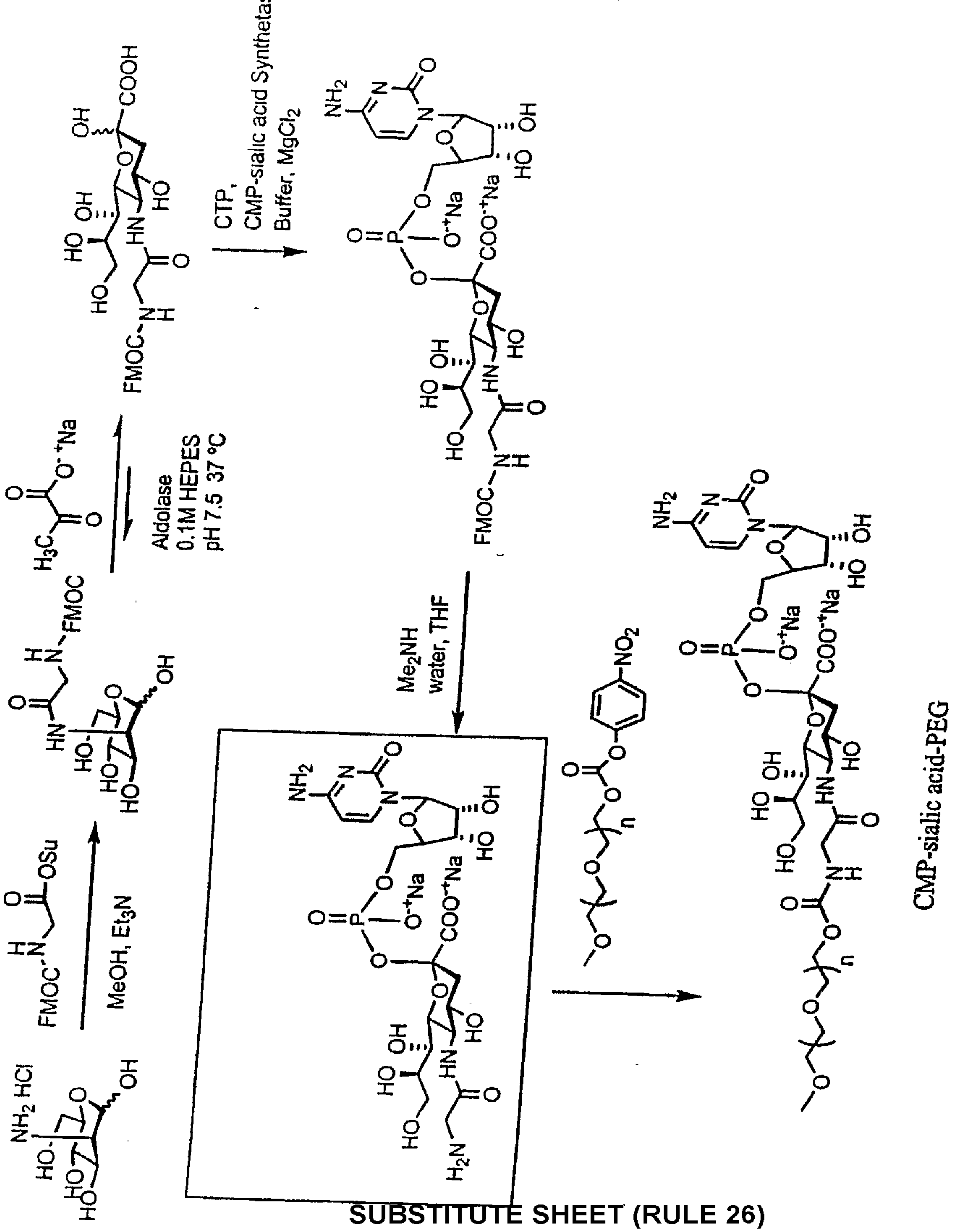
SUBSTITUTE SHEET (RULE 26)

FIGURE 1B



SUBSTITUTE SHEET (RULE 26)

FIGURE 2



SUBSTITUTE SHEET (RULE 26)

FIGURE 3A

Protein	Organism	EC#	GenBank / GenPept		SwissProt	PDB / 3D
At1g08280	*Arabidopsis thaliana*	n.d.	AC011438 BT004583 NC_003070	AAF18241.1 AAO42829.1 **NP_172305.1**	Q84W00 Q9SGD2	
At1g08660/F22O13.14	*Arabidopsis thaliana*	n.d.	AC003981 AY064135 AY124807 NC_003070 NM_180609	AAF99778.1 **AAL36042.1** AAM70516.1 NP_172342.1 NP_850940.1	Q8VZJ0 Q9FRR9	
At3g48820/T21J18_90	*Arabidopsis thaliana*	n.d.	AY080589 AY133816 AL132963 NM_114741	AAL85966.1 AAM91750.1 **CAB87910.1** NP_190451.1	Q8RY00 Q9M301	
α-2,3-sialyltransferase (ST3GAL-IV)	*Bos taurus*	n.d.	AJ584673	**CAE48298.1**		
α-2,3-sialyltransferase (St3Gal-V)	*Bos taurus*	n.d.	AJ585768	**CAE51392.1**		
α-2,6-sialyltransferase (Siat7b)	*Bos taurus*	n.d.	AJ620651	**CAF05850.1**		
α-2,8-sialyltransferase (SIAT8A)	*Bos taurus*	2.4.99.8	AJ699418	**CAG27880.1**		
α-2,8-sialyltransferase (Siat8D)	*Bos taurus*	n.d.	AJ699421 .	**CAG27883.1**		
α-2,8-sialyltransferase ST8Siα-III (Siat8C)	*Bos taurus*	n.d.	AJ704563	**CAG28696.1**		
CMP α-2,6-sialyltransferase (ST6Gal I)	*Bos taurus*	2.4.99.1	Y15111 NM_177517	CAA75385.1 NP_803483.1	O18974	
sialyltransferase 8 (fragment)	*Bos taurus*	n.d.	AF450088	**AAL47018.1**	Q8WN13	
sialyltransferase ST3Gal-II (Siat4B)	*Bos taurus*	n.d.	AJ748841	**CAG44450.1**		
sialyltransferase ST3Gal-III (Siat6)	*Bos taurus*	n.d.	AJ748842	**CAG44451.1**		
sialyltransferase ST3Gal-VI (Siat10)	*Bos taurus*	n.d.	AJ748843	**CAG44452.1**		
ST3Gal I	*Bos taurus*	n.d.	AJ305086	**CAC24698.1**	Q9BEG4	
St6GalNAc-VI	*Bos taurus*	n.d.	AJ620949	**CAF05586.1**		
CDS4	*Branchiostoma floridae*	n.d.	AF391289	**AAM18873.1**	Q8T771	
polysialyltransferase (PST) (fragment) ST8Sia IV	*Cercopithecus aethiops*	2.4.99.-	AF210729	**AAF17105.1**	Q9TT09	
polysialyltransferase (STX) (fragment) ST8Sia II	*Cercopithecus aethiops*	2.4.99.-	AF210318	**AAF17104.1**	Q9TT10	
α-2,3-sialyltransferase ST3Gal I (Siat4)	*Ciona intestinalis*	n.d.	AJ626815	**CAF25173.1**		
α-2,3-sialyltransferase ST3Gal I (Siat4)	*Ciona savignyi*	n.d.	AJ626814	**CAF25172.1**		
α-2,8-polysialyltransferase ST8Sia IV	*Cricetulus griseus*	2.4.99.-	- Z46801	AAE28634 CAA86822.1	**Q64690**	
Gal β-1,3/4-GlcNAc α-2,3-sialyltransferase St3Gal I	*Cricetulus griseus*	n.d.	AY266675	**AAP22942.1**	Q80WL0	
Gal β-1,3/4-GlcNAc α-2,3-sialyltransferase St3Gal II (fragment)	*Cricetulus griseus*	n.d.	AY266676	**AAP22943.1**	Q80WK9	
α-2,3-sialyltransferase ST3Gal I (Siat4)	*Danio rerio*	n.d.	AJ783740	**CAH04017.1**		
α-2,3-sialyltransferase ST3Gal II (Siat5)	*Danio rerio*	n.d.	AJ783741	**CAH04018.1**		

SUBSTITUTE SHEET (RULE 26)

FIGURE 3B

Protein	Organism	EC#	GenBank / GenPept		SwissProt	PDB / 3D
α-2,3-sialyltransferase ST3Gal III (Siat6)	*Danio rerio*	n.d.	AJ626821	**CAF25179.1**		
α-2,3-sialyltransferase ST3Gal IV (Siat4c)	*Danio rerio*	n.d.	AJ744809	**CAG32845.1**		
α-2,3-sialyltransferase ST3Gal V-r (Siat5-related)	*Danio rerio*	n.d.	AJ783742	**CAH04019.1**		
α-2,6-sialyltransferase ST6Gal I (Siat1)	*Danio rerio*	n.d.	AJ744801	**CAG32837.1**		
α-2,6-sialyltransferase ST6GalNAc II (Siat7B)	*Danio rerio*	n.d.	AJ634459	**CAG25680.1**		
α-2,6-sialyltransferase ST6GalNAc V (Siat7E) (fragment)	*Danio rerio*	n.d.	AJ646874	**CAG26703.1**		
α-2,6-sialyltransferase ST6GalNAc VI (Siat7F) (fragment)	*Danio rerio*	n.d.	AJ646883	**CAG26712.1**		
α-2,8-sialyltransferase ST8Sia I (Siat 8A) (fragment)	*Danio rerio*	n.d.	AJ715535	**CAG29374.1**		
α-2,8-sialyltransferase ST8Sia III (Siat 8C) (fragment)	*Danio rerio*	n.d.	AJ715543	**CAG29382.1**		
α-2,8-sialyltransferase ST8Sia IV (Siat 8D) (fragment)	*Danio rerio*	n.d.	AJ715545	**CAG29384.1**		
α-2,8-sialyltransferase ST8Sia V (Siat 8E) (fragment)	*Danio rerio*	n.d.	AJ715546	**CAG29385.1**		
α-2,8-sialyltransferase ST8Sia VI (Siat 8F) (fragment)	*Danio rerio*	n.d.	AJ715551	**CAG29390.1**		
β-galactosamide α-2,6-sialyltransferase II (ST6Gal II)	*Danio rerio*	n.d.	AJ627627	**CAF29495.1**		
N-glycan α-2,8-sialyltransferase	*Danio rerio*	n.d.	BC050483 AY055462 NM_153662	AAH50483.1 **AAL17875.1** NP_705948.1	Q7ZU51 Q8QH83	
ST3Gal III-related (siat6r)	*Danio rerio*	n.d.	BC053179 AJ626820 NM_200355	AAH53179.1 **CAF25178.1** NP_956649.1	Q7T3B9	
St3Gal-V	*Danio rerio*	n.d.	AJ619960	**CAF04061.1**		
st6GalNAc-VI	*Danio rerio*	n.d.	BC060932 AJ620947	**AAH60932.1** CAF06584.1		
α-2,6-sialyltransferase (CG4871) ST6Gal I	*Drosophila melanogaster*	2.4.99.1	AE003465 AF218237 AF397532 AE003465 NM_079129 NM_166684	AAF47256.1 **AAG13185.1** AAK92126.1 AAM70791.1 NP_523853.1 NP_726474.1	Q9GU23 Q9W121	
α-2,3-sialyltransferase (ST3Gal-VI)	*Gallus gallus*	n.d.	AJ585767 AJ627204	**CAE51391.1** CAF25503.1		
α-2,3-sialyltransferase ST3Gal I	*Gallus gallus*	2.4.99.4	X80503 NM_205217	**CAA56666.1** NP_990548.1	Q11200	
α-2,3-sialyltransferase ST3Gal IV (fragment)	*Gallus gallus*	2.4.99.-	AF035250	AAC14163.1	**O73724**	
α-2,3-sialytransferase (ST3GAL-II)	*Gallus gallus*	n.d.	AJ585761	**CAE51385.2**		
α-2,6-sialyltransferase (Siat7b)	*Gallus gallus*	n.d.	AJ620653	**CAF05852.1**		
α-2,6-sialyltransferase ST6Gal I	*Gallus gallus*	2.4.99.1	X75558 NM_205241	**CAA53235.1** NP_990572.1	Q92182	
α-2,6-sialyltransferase	*Gallus gallus*	2.4.99.3	-	AAE68028.1	Q92183	

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FIGURE 3C

Protein	Organism	EC#	GenBank / GenPept		SwissProt	PDB / 3D
ST6GalNAc I			- X74946 NM_205240	AAE68029.1 CAA52902.1 NP_990571.1		
α-2,6-sialyltransferase ST6GalNAc II	*Gallus gallus*	2.4.99.-	X77775 NM_205233	AAE68030.1 CAA54813.1 NP_990564.1	Q92184	
α-2,6-sialyltransferase ST6GalNAc III (SIAT7C) (fragment)	*Gallus gallus*	n.d.	AJ634455	CAG25677.1		
α-2,6-sialyltransferase ST6GalNAc V (SIAT7E) (fragment)	*Gallus gallus*	n.d.	AJ646877	CAG26706.1		
α-2,8-sialyltransferase (GD3 Synthase) ST8Sia I	*Gallus gallus*	2.4.99.-	U73176	AAC28888.1	P79783	
α-2,8-sialyltransferase (SIAT8B)	*Gallus gallus*	n.d.	AJ699419	CAG27881.1		
α-2,8-sialyltransferase (SIAT8C)	*Gallus gallus*	n.d.	AJ699420	CAG27882.1		
α-2,8-sialyltransferase (SIAT8F)	*Gallus gallus*	n.d.	AJ699424	CAG27886.1		
α-2,8-syalyltransferase ST8Siα-V (SIAT8C)	*Gallus gallus*	n.d.	AJ704564	CAG28697.1		
β-galactosamide α-2,6-sialyltransferase II (ST6Gal II)	*Gallus gallus*	n.d.	AJ627629	CAF29497.1		
GM3 synthase (SIAT9)	*Gallus gallus*	2.4.99.9	AY515255	AAS83519.1		
polysialyltransferase ST8Sia IV	*Gallus gallus*	2.4.99.-	AF008194	AAB95120.1	O42399	
α-2,3-sialyltransferase ST3Gal I	*Homo sapiens*	2.4.99.4	L29555 AF059321 L13972 AF155238 AF186191 BC018357 NM_003033 NM_173344	AAA36612.1 AAC17874.1 AAC37574.1 AAD39238.1 AAG29876.1 AAH18357.1 NP_003024.1 NP_775479.1	Q11201 O60677 Q9UN51	
α-2,3-sialyltransferase ST3Gal II	*Homo sapiens*	2.4.99.4	U63090 BC036777 X96667 NM_006927	AAB40389.1 AAH36777.1 CAA65447.1 NP_008858.1	Q16842 O00654	
α-2,3-sialyltransferase ST3Gal III (SiaT6)	*Homo sapiens*	2.4.99.6	L23768 BC050380 AF425851 AF425852 AF425853 AF425854 AF425855 AF425856 AF425857 AF425858 AF425859 AF425860 AF425861 AF425862 AF425863 AF425864 AF425865 AF425866 AF425867 AY167992 AY167993 AY167994	AAA35778.1 AAH50380.1 AAO13859.1 AAO13860.1 AAO13861.1 AAO13862.1 AAO13863.1 AAO13864.1 AAO13865.1 AAO13866.1 AAO13867.1 AAO13868.1 AAO13869.1 AAO13870.1 AAO13871.1 AAO13872.1 AAO13873.1 AAO13874.1 AAO13875.1 AAO38806.1 AAO38807.1 AAO38808.1	Q11203 Q86UR6 Q86UR7 Q86UR8 Q86UR9 Q86US0 Q86US1 Q86US2 Q8IX43 Q8IX44 Q8IX45 Q8IX46 Q8IX47 Q8IX48 Q8IX49 Q8IX50 Q8IX51 Q8IX52 Q8IX53 Q8IX54 Q8IX55 Q8IX56	

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FIGURE 3D

Protein	Organism		EC#	GenBank / GenPept		SwissProt	PDB / 3D
				AY167995	AAO38809.1	Q8IX57	
				AY167996	AAO38810.1	Q8IX58	
				AY167997	AAO38811.1		
				AY167998	AAO38812.1		
				NM_006279	NP_006270.1		
				NM_174964	NP_777624.1		
				NM_174965	NP_777625.1		
				NM_174966	NP_777626.1		
				NM_174967	NP_777627.1		
				NM_174969	NP_777629.1		
				NM_174970	NP_777630.1		
				NM_174972	NP_777632.1		
α-2,3-sialyltransferase ST3Gal IV		*Homo sapiens*	2.4.99.-	L23767	AAA16460.1	Q11206	
				AF035249	AAC14162.1	O60497	
				BC010645	AAH10645.1	Q96QQ9	
				AY040826	AAK93790.1	Q8N6A6	
				AF516602	AAM66431.1	Q8N6A7	
				AF516603	AAM66432.1	Q8NFD3	
				AF516604	AAM66433.1	Q8NFG7	
				AF525084	AAM81378.1		
				X74570	CAA52662.1		
				CR456858	CAG33139.1		
				NM_006278	NP_006269.1		
α-2,3-sialyltransferase ST3Gal VI		*Homo sapiens*	2.4.99.4	AF119391	AAD39131.1	Q9Y274	
				BC023312	AAH23312.1		
				AB022918	BAA77609.1		
				AX877828	CAE89895.1		
				AX886023	CAF00161.1		
				NM_006100	NP_006091.1		
α-2,6-sialyltransferase (ST6Gal II ; KIAA1877)		*Homo sapiens*	n.d.	BC008680	AAH08680.1	Q86Y44	
				AB058780	BAB47506.1	Q8IUG7	
				AB059555	BAC24793.1	Q96HE4	
				AJ512141	CAD54408.1	Q96JF0	
				AX795193	CAE48260.1		
				AX795193	CAE48261.1		
				NM_032528	NP_115917.1		
α-2,6-sialyltransferase (ST6GALNAC III)		*Homo sapiens*	n.d.	BC059363	AAH59363.1	Q8N259	
				AY358540	AAQ88904.1	Q8NDV1	
				AK091215	BAC03611.1		
				AJ507291	CAD45371.1		
				NM_152996	NP_694541.1		
α-2,6-sialyltransferase (ST6GalNAc V)		*Homo sapiens*	n.d.	BC001201	AAH01201.1	Q9BVH7	
				AK056241	BAB71127.1		
				AL035409	CAB72344.1		
				AJ507292	CAD45372.1		
				NM_030965	NP_112227.1		
α-2,6-sialyltransferase (SThM) ST6GalNAc II		*Homo sapiens*	2.4.99.-	U14550	AAA52228.1	Q9UJ37	
				BC040455	AAH40455.1	Q12971	
				AJ251053	CAB61434.1		
				NM_006456	NP_006447.1		
α-2,6-sialyltransferase ST6Gal I		*Homo sapiens*	2.4.99.1	BC031476	AAH31476.1	P15907	
				BC040009	AAH40009.1		
				A17362	CAA01327.1		
				A23699	CAA01686.1		
				X17247	CAA35111.1		
				X54363	CAA38246.1		
				X62822	CAA44634.1		
				NM_003032	NP_003023.1		
				NM_173216	NP_775323.1		
α-2,6-sialyltransferase ST6GalNAc I		*Homo sapiens*	2.4.99.3	BC022462	AAH22462.1	Q8TBJ6	
				AY096001	AAM22800.1	Q9NSC7	
				AY358918	AAQ89277.1	Q9NXQ7	
				AK000113	BAA90953.1		
				Y11339	CAA72179.2		

FIGURE 3E

Protein		Organism	EC#	GenBank / GenPept		SwissProt	PDB / 3D
				NM_018414	NP_060884.1		
α-2,8-polysialyltransferase ST8Sia IV		*Homo sapiens*	2.4.99.-	L41680 BC027866 BC053657 NM_005668	AAC41775.1 AAH27866.1 **AAH53657.1** NP_005659.1	Q8N1F4 Q92187 Q92693	
α-2,8-sialyltransferase (GD3 synthase) ST8Sia I		*Homo sapiens*	2.4.99.8	L32867 L43494 BC046158 - AY569975 D26360 X77922 NM_003034	AAA62366.1 **AAC37586.1** AAH46158.1 AAQ53140.1 AAS75783.1 BAA05391.1 CAA54891.1 NP_003025.1	Q86X71 Q92185 Q93064	
α-2,8-sialyltransferase ST8Sia II	-	*Homo sapiens*	2.4.99.-	L29556 U82762 U33551 BC069584 NM_006011	AAA36613.1 AAB51242.1 **AAC24458.1** AAH69584.1 NP_006002.1	Q92186 Q92470 Q92746	
α-2,8-sialyltransferase ST8Sia III		*Homo sapiens*	2.4.99.-	AF004668 AF003092 NM_015879	**AAB87642.1** AAC15901.2 NP_056963.1	O43173 Q9NS41	
α-2,8-sialyltransferase ST8Sia V		*Homo sapiens*	2.4.99.-	U91641 CR457037 NM_013305	**AAC51727.1** CAG33318.1 NP_037437.1	O15466	
ENSP00000020221 (fragment)			n.d.	AC023295	-		
lactosylceramide α-2,3-sialyltransferase (ST3Gal V)		*Homo sapiens*	2.4.99.9	AF105026 AF119415 BC065936 AY152815 AAP65066 AY359105 AB018356 AX876536 NM_003896	**AAD14634.1** AAF66146.1 AAH65936.1 AAO16866.1 AAP65066.1 AAQ89463.1 BAA33950.1 CAE89320.1 NP_003887.2	Q9UNP4 O94902	
N-acetylgalactosaminide α-2,6-sialyltransferase (ST6GalNAc VI)		*Homo sapiens*	2.4.99.-	BC006564 BC007802 BC016299 AY358672 AB035173 AK023900 AJ507293 AX880950 CR457318 NM_013443	AAH06564.1 AAH07802.1 AAH16299.1 AAQ89035.1 **BAA87035.1** BAB14715.1 CAD45373.1 CAE91145.1 CAG33599.1 NP_038471.2	Q969X2 Q9H8A2 Q9ULB8	
N-acetylgalactosaminide α-2,6-sialyltransferase IV (ST6GalNAc IV)		*Homo sapiens*	2.4.99.-	AF127142 BC036705 - AB035172 AK000600 Y17461 AJ271734 AX061620 AX068265 AX969252 NM_014403 NM_175039	AAF00102.1 AAH36705.1 AAP63349.1 **BAA87034.1** BAA91281.1 CAB44354.1 CAC07404.1 CAC24981.1 CAC27250.1 CAF14360.1 NP_055218.3 NP_778204.1	Q9H4F1 Q9NWU6 Q9UKU1 Q9ULB9 Q9Y3G3 Q9Y3G4	
ST8SIA-VI (fragment)		*Homo sapiens*	n.d.	AJ621583 XM_291725	**CAF21722.1** XP_291725.2		
unnamed protein product		*Homo sapiens*	n.d.	AK021929 AX881696	**BAB13940.1** CAE91353.1	Q9HAA9	
Gal β-1,3/4-GlcNAc α-		*Mesocricetus*	2.4.99.6	AJ245699	**CAB53394.1**	Q9QXF6	

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FIGURE 3F

Protein	Organism		EC#	GenBank / GenPept		SwissProt	PDB / 3D
2,3-sialyltransferase (ST3Gal III)		auratus					
Gal β-1,3/4-GlcNAc α-2,3-sialyltransferase (ST3Gal IV)		*Mesocricetus auratus*	2.4.99.6	AJ245700	CAB53395.1	Q9QXF5	
GD3 synthase (fragment) ST8Sia I		*Mesocricetus auratus*	n.d.	AF141657	AAD33879.1	Q9WUL1	
polysialyltransferase (ST8Sia IV)		*Mesocricetus auratus*	2.4.99.-	AJ245701	CAB53396.1	Q9QXF4	
α-2,3-sialyltransferase ST3Gal I	*St3gal1*	*Mus musculus*	2.4.99.4	AF214028 AK031344 AK078469 X73523 NM_009177	AAF60973.1 BAC27356.1 BAC37290.1 CAA51919.1 NP_033203.1	P54751 Q11202 Q9JL30	
α-2,3-sialyltransferase ST3Gal II	*St3gal2*	*Mus musculus*	2.4.99.4	BC015264 BC066064 AK034554 AK034863 AK053827 X76989 NM_009179 NM_178048	AAH15264.1 AAH66064.1 BAC28752.1 BAC28859.1 BAC35543.1 CAA54294.1 NP_033205.1 NP_835149.1	Q11204 Q8BPL0 Q8BSA0 Q8BSE9 Q91WH6	
α-2,3-sialyltransferase ST3Gal III	*St3gal3*	*Mus musculus*	2.4.99.-	BC006710 AK005053 AK013016 X84234 NM_009176	AAH06710.1 BAB23779.1 BAB28598.1 CAA59013.1 NP_033202.2	P97325 Q922X5 Q9CZ48 Q9DBB6	
α-2,3-sialyltransferase ST3Gal IV	*St3gal4*	*Mus musculus*	2.4.99.4	BC011121 BC050773 D28941 AK008543 AB061305 X95809 NM_009178	AAH11121.1 AAH50773.1 BAA06068.1 BAB25732.1 BAB47508.1 CAA65076.1 NP_033204.2	P97354 Q61325 Q91Y74 Q921R5 Q9CVE8	
α-2,3-sialyltransferase ST3Gal VI	*St3gal6*	*Mus musculus*	2.4.99.4	AF119390 BC052338 AB063326 AK033562 AK041173 NM_018784	AAD39130.1 AAH52338.1 BAB79494.1 BAC28360.1 BAC30851.1 NP_061254	Q80UR7 Q8BLV1 Q8VIB3 Q9WVG2	
α-2,6-sialyltransferase ST6GalNAc II	*St6galnac2*	*Mus musculus*	2.4.99.-	NM_009180 BC010208 AB027198 AK004613 X93999 X94000 NM_009180	6677963 AAH10208.1 BAB00637.1 BAB23410.1 CAA63821.1 CAA63822.1 NP_033206.2	P70277 Q9DC24 Q9JJM5	
α-2,6-sialyltransferase ST6Gal I	*St6gal1*	*Mus musculus*	2.4.99.1	- BC027833 D16106 AK034768 AK084124 NM_145933	AAE68031.1 AAH27833.1 BAA03680.1 BAC28828.1 BAC39120.1 NP_666045.1	Q64685 Q8BM62 Q8K1L1	
α-2,6-sialyltransferase ST6Gal II	*St6gal2*	*Mus musculus*	n.d.	AK082566 AB095093 AK129462 NM_172829	BAC38534.1 BAC87752.1 BAC98272.1 NP_766417.1	Q8BUU4	
α-2,6-sialyltransferase ST6GalNAc I	*St6galnac1*	*Mus musculus*	2.4.99.3	Y11274 NM_011371	CAA72137.1 NP_035501.1	Q9QZ39 Q9JJP5	
α-2,6-sialyltransferase ST6GalNAc III	*St6galnac3*	*Mus musculus*	n.d.	BC058387 AK034804 Y11342 Y11343	AAH58387.1 BAC28836.1 CAA72181.2 CAB95031.1	Q9WUV2 Q9JHP5	

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FIGURE 3G

Protein	Organism		EC#	GenBank / GenPept		SwissProt	PDB / 3D
				NM_011372	NP_035502		
α-2,6-sialyltransferase ST6GalNAc IV	*St6galnac4*	*Mus musculus*	2.4.99.7	BC056451 AK085730 AJ007310 Y15779 Y15780 Y19055 Y19057 NM_011373	AAH56451.1 BAC39523.1 CAA07446.1 CAB43507.1 **CAB43514.1** CAB93946.1 CAB93948.1 NP_035503.1	Q8C3J2 Q9JHP2 Q9R2B6 O88725 Q9JHP0 Q9QUP9 Q9R2B5	
α-2,8-sialyltransferase (GD3 synthase) ST8Sia I	*St8sia1*	*Mus musculus*	2.4.99.8	L38677 BC024821 AK046188 AK052444 X84235 AJ401102 NM_011374	AAA91869.1 **AAH24821.1** BAC32625.1 BAC34994.1 CAA59014.1 CAC20706.1 NP_035504.1	Q64468 Q64687 Q8BL76 Q8BWI0 Q8K1C1 Q9EPK0	
α-2,8-sialyltransferase (ST8Sia VI)	*St8sia6*	*Mus musculus*	n.d.	AB059554 AK085105 NM_145838	**BAC01265.1** BAC39367.1 NP_665837.1	Q8BI43 Q8K4T1	
α-2,8-sialyltransferase ST8Sia II	*St8sia2*	*Mus musculus*	2.4.99.-	X83562 X99646 X99647 X99648 X99649 X99650 X99651 NM_009181	CAA58548.1 CAA67965.1 CAA67965.1 CAA67965.1 CAA67965.1 CAA67965.1 CAA67965.1 NP_033207.1	O35696	
α-2,8-sialyltransferase ST8Sia IV	*St8sia4*	*Mus musculus*	2.4.99.8	BC060112 AK003690 AK041723 AJ223956 X86000 Y09484 NM_009183	AAH60112.1 BAB22941.1 BAC31044.1 **CAA11685.1** CAA59992.1 CAA70692.1 NP_033209.1	Q64692 Q8BY70	
α-2,8-sialyltransferase ST8Sia V	*St8sia5*	*Mus musculus*	2.4.99.-	BC034855 AK078670 X98014 X98014 X98014 NM_013666 NM_153124 NM_177416	AAH34855.1 BAC37354.1 **CAA66642.1** CAA66643.1 CAA66644.1 NP_038694.1 NP_694764.1 NP_803135.1	P70126 P70127 P70128 Q8BJW0 Q8JZQ3	
α-2,8-sialytransferase ST8Sia III	*St8sia3*	*Mus musculus*	2.4.99.-	BC075645 AK015874 X80502 NM_009182	AAH75645.1 BAB30012.1 CAA56665.1 NP_033208.1	**Q64689** Q9CUJ6	
GD1 synthase (ST6GalNAc V)	*St6galnac5*	*Mus musculus*	n.d.	BC055737 AB030836 AB028840 AK034387 AK038434 AK042683 NM_012028	**AAH55737.1** BAA85747.1 BAA89292.1 BAC28693.1 BAC29997.1 BAC31331.1 NP_036158.2	Q8CAM7 Q8CBX1 Q9QYJ1 Q9R0K6	
GM3 synthase (α-2,3-sialyltransferase) ST3Gal V	*St3gal5*	*Mus musculus*	2.4.99.9	AF119416 - AB018048 AB013302 AK012961 Y15003 NM_011375	**AAF66147.1** AAP65063.1 BAA33491.1 BAA76467.1 BAB28571.1 CAA75235.1 NP_035505.1	O88829 Q9CZ65 Q9QWF9	
N-acetylgalactosaminide α-2,6-sialyltransferase (ST6GalNAc VI)	*St6galnac6*	*Mus musculus*	2.4.99.-	BC036985 AB035174 AB035123 AK030648	**AAH36985.1** BAA87036.1 BAA95940.1 BAC27064.1	Q8CDC3 Q8JZW3 Q9JM95 Q9R0G9	

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FIGURE 3H

Protein	Organism		EC#	GenBank / GenPept		SwissProt	PDB / 3D
				NM_016973	NP_058669.1		
M138L		*Myxoma virus*	n.d.	U46578 AF170726 NC_001132	AAD00069.1 AAE61323.1 AAE61326.1 **AAF15026.1** NP_051852.1		
α-2,3-sialyltransferase (St3Gal-I)		*Oncorhynchus mykiss*	n.d.	AJ585760	**CAE51384.1**		
α-2,6-sialyltransferase (Siat1)		*Oncorhynchus mykiss*	n.d.	AJ620649	**CAF05848.1**		
α-2,8-polysialyltransferase IV (ST8Sia IV)		*Oncorhynchus mykiss*	n.d.	AB094402	**BAC77411.1**	Q7T2X5	
GalNAc α-2,6-sialyltransferase (RtST6GalNAc)	-	*Oncorhynchus mykiss*	n.d.	AB097943	**BAC77520.1**	Q7T2X4	
α-2,3-sialyltransferase ST3Gal IV		*Oryctolagus cuniculus*	2.4.99.-	AF121967	**AAF28871.1**	Q9N257	
OJ1217_F02.7		*Oryza sativa (japonica cultivar-group)*	n.d.	AP004084	**BAD07616.1**		
OSJNBa0043L24.2 or OSJNBb0002J11.9		*Oryza sativa (japonica cultivar-group)*	n.d.	AL731626 AL662969	**CAD41185.1** CAE04714.1		
P0683f02.18 or P0489B03.1		*Oryza sativa (japonica cultivar-group)*	n.d.	AP003289 AP003794	**BAB63715.1** BAB90552.1		
α-2,6-sialyltransferase ST6GalNAc V (Siat7E) (fragment)		*Oryzias latipes*	n.d.	AJ646876	**CAG26705.1**		
α-2,3-sialyltransferase ST3Gal I (Siat4)		*Pan troglodytes*	n.d.	AJ744803	**CAG32839.1**		
α-2,3-sialyltransferase ST3Gal II (Siat5)		*Pan troglodytes*	n.d.	AJ744804	**CAG32840.1**		
α-2,3-sialyltransferase ST3Gal III (Siat6)		*Pan troglodytes*	n.d.	AJ626819	**CAF25177.1**		
α-2,3-sialyltransferase ST3Gal IV (Siat4c)		*Pan troglodytes*	n.d.	AJ626824	**CAF25182.1**		
α-2,3-sialyltransferase ST3Gal VI (Siat10)		*Pan troglodytes*	n.d.	AJ744808	**CAG32844.1**		
α-2,6-sialyltransferase (Sia7A)		*Pan troglodytes*	n.d.	AJ748740	**CAG38615.1**		
α-2,6-sialyltransferase (Sia7B)		*Pan troglodytes*	n.d.	AJ748741	**CAG38616.1**		
α-2,6-sialyltransferase ST6GalNAc III (Siat7C)		*Pan troglodytes*	n.d.	AJ634454	**CAG25676.1**		
α-2,6-sialyltransferase ST6GalNAc IV (Siat7D) (fragment)		*Pan troglodytes*	n.d.	AJ646870	**CAG26699.1**		
α-2,6-sialyltransferase ST6GalNAc V (Siat7E)		*Pan troglodytes*	n.d.	AJ646875	**CAG26704.1**		
α-2,6-sialyltransferase ST6GalNAc VI (Siat7F) (fragment)		*Pan troglodytes*	n.d.	AJ646882	**CAG26711.1**		
α-2,8-sialyltransferase 8A (Siat8A)		*Pan troglodytes*	2.4.99.8	AJ697658	**CAG26896.1**		
α-2,8-sialyltransferase 8B (Siat8B)		*Pan troglodytes*	n.d.	AJ697659	**CAG26897.1**		
α-2,8-sialyltransferase 8C (Siat8C)		*Pan troglodytes*	n.d.	AJ697660	**CAG26898.1**		
α-2,8-sialyltransferase 8D (Siat8D)		*Pan troglodytes*	n.d.	AJ697661	**CAG26899.1**		
α-2,8-sialyltransferase		*Pan troglodytes*	n.d.	AJ697662	**CAG26900.1**		

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FIGURE 3I

Protein	Organism	EC#	GenBank / GenPept		SwissProt	PDB / 3D
8E (Siat8E)						
α-2,8-sialyltransferase 8F (Siat8F)	*Pan troglodytes*	n.d.	AJ697663	**CAG26901.1**		
β-galactosamide α-2,6-sialyltransferase I (ST6Gal I; Siat1)	*Pan troglodytes*	2.4.99.1	AJ627624	**CAF29492.1**		
β-galactosamide α-2,6-sialyltransferase II (ST6Gal II)	*Pan troglodytes*	n.d.	AJ627625	**CAF29493.1**		
GM3 synthase ST3Gal V (Siat9)	*Pan troglodytes*	n.d.	AJ744807	**CAG32843.1**		
S138L	*Rabbit fibroma virus Kasza*	n.d.	NC_001266	**NP_052025**		
α-2,3-sialyltransferase ST3Gal III	*Rattus norvegicus*	2.4.99.6	M97754 NM_031697	AAA42146.1 NP_113885.1	**Q02734**	
α-2,3-sialyltransferase ST3Gal IV (Siat4c)	*Rattus norvegicus*	n.d.	AJ626825	**CAF25183.1**		
α-2,3-sialyltransferase ST3Gal VI	*Rattus norvegicus*	n.d.	AJ626743	**CAF25053.1**		
α-2,6-sialyltransferase ST3Gal II	*Rattus norvegicus*	2.4.99.-	X76988 NM_031695	CAA54293.1 NP_113883.1	**Q11205**	
α-2,6-sialyltransferase ST6Gal I	*Rattus norvegicus*	2.4.99.1	M18769 M83143	AAA41196.1 AAB07233.1	**P13721**	
α-2,6-sialyltransferase ST6GalNAc I (Siat7A)	*Rattus norvegicus*	n.d.	AJ634458	**CAG25684.1**		
α-2,6-sialyltransferase ST6GalNAc II (Siat7B)	*Rattus norvegicus*	n.d.	AJ634457	**CAG25679.1**		
α-2,6-sialyltransferase ST6GalNAc III	*Rattus norvegicus*	2.4.99.-	L29554 BC072501 NM_019123	AAC42086.1 AAH72501.1 NP_061996.1	**Q64686**	
α-2,6-sialyltransferase ST6GalNAc IV (Siat7D) (fragment)	*Rattus norvegicus*	n.d.	AJ646871	**CAG26700.1**		
α-2,6-sialyltransferase ST6GalNAc V (Siat7E)	*Rattus norvegicus*	n.d.	AJ646872	**CAG26701.1**		
α-2,6-sialyltransferase ST6GalNAc VI (Siat7F) (fragment)	*Rattus norvegicus*	n.d.	AJ646881	**CAG26710.1**		
α-2,8-sialyltransferase (GD3 synthase) ST8Sia I	*Rattus norvegicus*	2.4.99.-	U53883 D45255	AAC27541.1 BAA08213.1	**P70554** **P97713**	
α-2,8-sialyltransferase (SIAT8E)	*Rattus norvegicus*	n.d.	AJ699422	**CAG27884.1**		
α-2,8-sialyltransferase (SIAT8F)	*Rattus norvegicus*	n.d.	AJ699423	**CAG27885.1**		
α-2,8-sialyltransferase ST8Sia II	*Rattus norvegicus*	2.4.99.-	L13445 NM_057156	AAA42147.1 NP_476497.1	**Q07977** **Q64688**	
α-2,8-sialyltransferase ST8Sia III	*Rattus norvegicus*	2.4.99.-	U55938 NM_013029	AAB50061.1 NP_037161.1	**P97877**	
α-2,8-sialyltransferase ST8Sia IV	*Rattus norvegicus*	2.4.99.-	U90215	AAB49989.1	**O08563**	
β-galactosamide α-2,6-sialyltransferase II (ST6Gal II)	*Rattus norvegicus*	n.d.	AJ627626	**CAF29494.1**		
GM3 synthase ST3Gal V	*Rattus norvegicus*	n.d.	AB018049 NM_031337	BAA33492.1 NP_112627.1	**O88830**	

FIGURE 3J

Protein	Organism	EC#	GenBank / GenPept		SwissProt	PDB / 3D
sialyltransferase ST3Gal-I (Siat4A)	*Rattus norvegicus*	n.d.	AJ748840	CAG44449.1		
α-2,3-sialyltransferase (St3Gal-II)	*Silurana tropicalis*	n.d.	AJ585763	CAE51387.1		
α-2,6-sialyltransferase (Siat7b)	*Silurana tropicalis*	n.d.	AJ620650	CAF05849.1		
α-2,6-sialyltransferase (St6galnac)	*Strongylocentrotus purpuratus*	n.d.	AJ699425	CAG27887.1		
α-2,3-sialyltransferase (ST3GAL-III)	*Sus scrofa*	n.d.	AJ585765	CAE51389.1		
α-2,3-sialyltransferase (ST3GAL-IV)	*Sus scrofa*	n.d.	AJ584674	CAE48299.1		
α-2,3-sialyltransferase ST3Gal I	*Sus scrofa*	2.4.99.4	M97753	AAA31125.1	Q02745	
α-2,6-sialyltransferase (fragment) ST6Gal I	*Sus scrofa*	2.4.99.1	AF136746	AAD33059.1	Q9XSG8	
β-galactosamide α-2,6-sialyltransferase (ST6GalNAc-V)	*Sus scrofa*	n.d.	AJ620948	CAF06585.2		
sialyltransferase (fragment) ST6Gal I	*sus scrofa*	n.d.	AF041031	AAC15633.1	O62717	
ST6GALNAC-V	*Sus scrofa*	n.d.	AJ620948	CAF06585.1		
α-2,3-sialyltransferase (Siat5-r)	*Takifugu rubripes*	n.d.	AJ744805	CAG32841.1		
α-2,3-sialyltransferase ST3Gal I (Siat4)	*Takifugu rubripes*	n.d.	AJ626816	CAF25174.1		
α-2,3-sialyltransferase ST3Gal II (Siat5) (fragment)	*Takifugu rubripes*	n.d.	AJ626817	CAF25175.1		
α-2,3-sialyltransferase ST3Gal III (Siat6)	*Takifugu rubripes*	n.d.	AJ626818	CAF25176.1		
α-2,6-sialyltransferase ST6Gal I (Siat1)	*Takifugu rubripes*	n.d.	AJ744800	CAG32836.1		
α-2,6-sialyltransferase ST6GalNAc II (Siat7B)	*Takifugu rubripes*	n.d.	AJ634460	CAG25681.1		
α-2,6-sialyltransferase ST6GalNAc II B (Siat7B-related)	*Takifugu rubripes*	n.d.	AJ634461	CAG25682.1		
α-2,6-sialyltransferase ST6GalNAc III (Siat7C) (fragment)	*Takifugu rubripes*	n.d.	AJ634456	CAG25678.1		
α-2,6-sialyltransferase ST6GalNAc IV (siat7D) (fragment)	*Takifugu rubripes*	2.4.99.3	Y17466 AJ646869	CAB44338.1 CAG26698.1	Q9W6U6	
α-2,6-sialyltransferase ST6GalNAc V (Siat7E) (fragment)	*Takifugu rubripes*	n.d.	AJ646873	CAG26702.1		
α-2,6-sialyltransferase ST6GalNAc VI (Siat7F) (fragment)	*Takifugu rubripes*	n.d.	AJ646880	CAG26709.1		
α-2,8-sialyltransferase ST8Sia I (Siat 8A) (fragment)	*Takifugu rubripes*	n.d.	AJ715534	CAG29373.1		
α-2,8-sialyltransferase ST8Sia II (Siat 8B) (fragment)	*Takifugu rubripes*	n.d.	AJ715538	CAG29377.1		
α-2,8-sialyltransferase ST8Sia III (Siat 8C) (fragment)	*Takifugu rubripes*	n.d.	AJ715541	CAG29380.1		
α-2,8-sialyltransferase ST8Sia IIIr (Siat 8Cr)	*Takifugu rubripes*	n.d.	AJ715542	CAG29381.1		
α-2,8-sialyltransferase ST8Sia V (Siat 8E)	*Takifugu rubripes*	n.d.	AJ715547	CAG29386.1		

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FIGURE 3K

Protein	Organism	EC#	GenBank / GenPept		SwissProt	PDB / 3D
(fragment)						
α-2,8-sialyltransferase ST8Sia VI (Siat 8F) (fragment)	*Takifugu rubripes*	n.d.	AJ715549	CAG29388.1		
α-2,8-sialyltransferase ST8Sia VIr (Siat 8Fr)	*Takifugu rubripes*	n.d.	AJ715550	CAG29389.1		
α-2,3-sialyltransferase (Siat5-r)	*Tetraodon nigroviridis*	n.d.	AJ744806	CAG32842.1		
α-2,3-sialyltransferase ST3Gal I (Siat4)	*Tetraodon nigroviridis*	n.d.	AJ744802	CAG32838.1		
α-2,3-sialyltransferase ST3Gal III (Siat6)	*Tetraodon nigroviridis*	n.d.	AJ626822	CAF25180.1		
α-2,6-sialyltransferase ST6GalNAc II (Siat7B)	*Tetraodon nigroviridis*	n.d.	AJ634462	CAG25683.1		
α-2,6-sialyltransferase ST6GalNAc V (Siat7E) (fragment)	*Tetraodon nigroviridis*	n.d.	AJ646879	CAG26708.1		
α-2,8-sialyltransferase ST8Sia I (Siat 8A) (fragment)	*Tetraodon nigroviridis*	n.d.	AJ715536	CAG29375.1		
α-2,8-sialyltransferase ST8Sia II (Siat 8B) (fragment)	*Tetraodon nigroviridis*	n.d.	AJ715537	CAG29376.1		
α-2,8-sialyltransferase ST8Sia III (Siat 8C) (fragment)	*Tetraodon nigroviridis*	n.d.	AJ715539	CAG29378.1		
α-2,8-sialyltransferase ST8Sia IIIr (Siat 8Cr) (fragment)	*Tetraodon nigroviridis*	n.d.	AJ715540	CAG29379.1		
α-2,8-sialyltransferase ST8Sia V (Siat 8E) (fragment)	*Tetraodon nigroviridis*	n.d.	AJ715548	CAG29387.1		
α-2,3-sialyltransferase (St3Gal-II)	*Xenopus laevis*	n.d.	AJ585762	CAE51386.1		
α-2,3-sialyltransferase (St3Gal-VI)	*Xenopus laevis*	n.d.	AJ585766	CAE51390.1		
α-2,3-sialyltransferase St3Gal-III (Siat6)	*Xenopus laevis*	n.d.	AJ585764 AJ626823	CAE51388.1 CAF25181.1		
α-2,8-polysialyltransferase	*Xenopus laevis*	2.4.99.-	AB007468	BAA32617.1	O93234	
α-2,8-sialyltransferase ST8Siα-I (Siat8A;GD3 synthase)	*Xenopus laevis*	n.d.	AY272056 AY272057 AJ704562	AAQ16162.1 AAQ16163.1 CAG28695.1		
Unknown (protein for MGC:81265)	*Xenopus laevis*	n.d.	BC068760	AAH68760.1		
α-2,3-sialyltransferase (3Gal-VI)	*Xenopus tropicalis*	n.d.	AJ626744	CAF25054.1		
α-2,3-sialyltransferase (Siat4c)	*Xenopus tropicalis*	n.d.	AJ622908	CAF22058.1		
α-2,6-sialyltransferase ST6GalNAc V (Siat7E) (fragment)	*Xenopus tropicalis*	n.d.	AJ646878	CAG26707.1		
α-2,8-sialyltransferase ST8Sia III (Siat 8C) (fragment)	*Xenopus tropicalis*	n.d.	AJ715544	CAG29383.1		
β-galactosamide α-2,6-sialyltransferase II (ST6Gal II)	*Xenopus tropicalis*	n.d.	AJ627628	CAF29496.1		
sialytransferase St8Sial	*Xenopus tropicalis*	n.d.	AY652775	AAT67042		
poly-α-2,8-sialosyl sialyltransferase (NeuS)	*Escherichia coli K1*	2.4.-.-	M76370 X60598	AAA24213.1 CAA43053.1	Q57269	
polysialyltransferase	*Escherichia coli K92*	2.4.-.-	M88479	AAA24215.1	Q47404	

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FIGURE 3L

Protein	Organism	EC#	GenBank / GenPept		SwissProt	PDB / 3D
α-2,8 polysialyltransferase SiaD	*Neisseria meningitidis B1940*	2.4.-.-	M95053 X78068	AAA20478.1 CAA54985.1	**Q51281** **Q51145**	
SynE	*Neisseria meningitidis FAM18*	n.d.	U75650	**AAB53842.1**	O06435	
polysialyltransferase (SiaD)(fragment)	*Neisseria meningitidis M1019*	n.d.	AY234192	**AAO85290.1**		
SiaD (fragment)	*Neisseria meningitidis M209*	n.d.	AY281046	**AAP34769.1**		
SiaD (fragment)	*Neisseria meningitidis M3045*	n.d.	AY281044	**AAP34767.1**		
polysialyltransferase (SiaD)(fragment)	*Neisseria meningitidis M3315*	n.d.	AY234191	**AAO85289.1**		
SiaD (fragment)	*Neisseria meningitidis M3515*	n.d.	AY281047	**AAP34770.1**		
polysialyltransferase (SiaD)(fragment)	*Neisseria meningitidis M4211*	n.d.	AY234190	**AAO85288.1**		
SiaD (fragment)	*Neisseria meningitidis M4642*	n.d.	AY281048	**AAP34771.1**		
polysialyltransferase (SiaD)(fragment)	*Neisseria meningitidis M5177*	n.d.	AY234193	**AAO85291.1**		
SiaD	*Neisseria meningitidis M5178*	n.d.	AY281043	**AAP34766.1**		
SiaD (fragment)	*Neisseria meningitidis M980*	n.d.	AY281045	**AAP34768.1**		
NMB0067	*Neisseria meningitidis MC58*	n.d.	NC_003112	**NP_273131**		
Lst	*Aeromonas punctata Sch3*	n.d.	AF126256	**AAS66624.1**		
ORF2	*Haemophilus influenzae A2*	n.d.	M94855	**AAA24979.1**		
HI1699	*Haemophilus influenzae Rd*	n.d.	U32842 NC_000907	**AAC23345.1** NP_439841.1	Q48211	
α-2,3-sialyltransferase	*Neisseria gonorrhoeae F62*	2.4.99.4	U60664	**AAC44539.1** AAE67205.1	P72074	
α-2,3-sialyltransferase	*Neisseria meningitidis 126E, NRCC 4010*	2.4.99.4	U60662	**AAC44544.2**		
α-2,3-sialyltransferase	*Neisseria meningitidis 406Y, NRCC 4030*	2.4.99.4	U60661	**AAC44543.1**		
α-2,3-sialyltransferase (NMB0922)	*Neisseria meningitidis MC58*	2.4.99.4	U60660 AE002443 NC_003112	AAC44541.1 AAF41330.1 **NP_273962.1**	P72097	
NMA1118	*Neisseria meningitidis Z2491*	n.d.	AL162755 NC_003116	CAB84380.1 NP_283887.1	**Q9JUV5**	
PM0508	*Pasteurella multocida PM70*	n.d.	AE006086 NC_002663	**AAK02592.1** NP_245445.1	Q9CNC4	
WaaH	*Salmonella enterica SARB25*	n.d.	AF519787	**AAM82550.1**	Q8KS93	
WaaH	*Salmonella enterica SARB3*	n.d.	AF519788	**AAM82551.1**	Q8KS92	
WaaH	*Salmonella enterica SARB39*	n.d.	AF519789	**AAM82552.1**		
WaaH	*Salmonella enterica SARB53*	n.d.	AF519790	**AAM82553.1**		
WaaH	*Salmonella enterica SARB57*	n.d.	AF519791	**AAM82554.1**	Q8KS91	
WaaH	*Salmonella enterica SARB71*	n.d.	AF519793	**AAM82556.1**	Q8KS89	
WaaH	*Salmonella enterica*	n.d.	AF519792	**AAM82555.1**	Q8KS90	

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FIGURE 3M

Protein	Organism	EC#	GenBank / GenPept		SwissProt	PDB / 3D
	SARB8					
WaaH	*Salmonella enterica SARC10V*	n.d.	AF519779	**AAM88840.1**	Q8KS99	
WaaH (fragment)	*Salmonella enterica SARC12*	n.d.	AF519781	**AAM88842.1**		
WaaH (fragment)	*Salmonella enterica SARC13I*	n.d.	AF519782	**AAM88843.1**	Q8KS98	
WaaH (fragment)	*Salmonella enterica SARC14I*	n.d.	AF519783	**AAM88844.1**	Q8KS97	
WaaH	*Salmonella enterica SARC15II*	n.d.	AF519784	**AAM88845.1**	Q8KS96	
WaaH	*Salmonella enterica SARC16II*	n.d.	AF519785	**AAM88846.1**	Q8KS95	
WaaH (fragment)	*Salmonella enterica SARC3I*	n.d.	AF519772	**AAM88834.1**	Q8KSA4	
WaaH (fragment)	*Salmonella enterica SARC4I*	n.d.	AF519773	**AAM88835.1**	Q8KSA3	
WaaH	*Salmonella enterica SARC5IIa*	n.d.	AF519774	**AAM88836.1**		
WaaH	*Salmonella enterica SARC6IIa*	n.d.	AF519775	**AAM88837.1**	Q8KSA2	
WaaH	*Salmonella enterica SARC8*	n.d.	AF519777	**AAM88838.1**	Q8KSA1	
WaaH	*Salmonella enterica SARC9V*	n.d.	AF519778	**AAM88839.1**	Q8KSA0	
UDP-glucose : α-1,2-glucosyltransferase (WaaH)	*Salmonella enterica subsp. arizonae SARC 5*	2.4.1.-	AF511116	**AAM48166.1**		
bifunctional α-2,3/-2,8-sialyltransferase (Cst-II)	*Campylobacter jejuni ATCC 43449*	n.d.	AF401529	**AAL06004.1**	Q93CZ5	
Cst	*Campylobacter jejuni 81-176*	n.d.	AF305571	**AAL09368.1**		
α-2,3-sialyltransferase (Cst-III)	*Campylobacter jejuni ATCC 43429*	2.4.99.-	AY044156	**AAK73183.1**		
α-2,3-sialyltransferase (Cst-III)	*Campylobacter jejuni ATCC 43430*	2.4.99.-	AF400047	**AAK85419.1**		
α-2,3-sialyltransferase (Cst-II)	*Campylobacter jejuni ATCC 43432*	2.4.99.-	AF215659	**AAG43979.1**	Q9F0M9	
α-2,3/8-sialyltransferase (CstII)	*Campylobacter jejuni ATCC 43438*	n.d.	AF400048	**AAK91725.1**	Q93MQ0	
α-2,3-sialyltransferase cst-II	*Campylobacter jejuni ATCC 43446*	2.4.99.-	AF167344	**AAF34137.1**		
α-2,3-sialyltransferase (Cst-II)	*Campylobacter jejuni ATCC 43456*	2.4.99.-	AF401528	**AAL05990.1**	Q93D05	
α-2,3-/α-2,8-sialyltransferase (CstII)	*Campylobacter jejuni ATCC 43460*	2.4.99.-	AY044868	**AAK96001.1**	Q938X6	
α-2,3/8-sialyltransferase (Cst-II)	*Campylobacter jejuni ATCC 700297*	n.d.	AF216647	**AAL36462.1**		
ORF	*Campylobacter jejuni GB11*	n.d.	AY422197	**AAR82875.1**		
α-2,3-sialyltransferase cstIII	*Campylobacter jejuni MSC57360*	2.4.99.-	AF195055	**AAG29922.1**		
α-2,3-sialyltransferase cstIII Cj1140	*Campylobacter jejuni NCTC 11168*	2.4.99.-	AL139077 NC_002163	**CAB73395.1** NP_282288.1	Q9PNF4	
α-2,3/α-2,8-sialyltransferase II (cstII)	*Campylobacter jejuni O:10*	n.d.	- AX934427	AAO96669.1 **CAF04167.1**		
α-2,3/α-2,8-sialyltransferase II (CstII)	*Campylobacter jejuni O:19*	n.d.	AX934431	**CAF04169.1**		
α-2,3/α-2,8-sialyltransferase II (CstII)	*Campylobacter jejuni O:36*	n.d.	AX934436	**CAF04171.1**		
α-2,3/α-2,8-	*Campylobacter*	n.d.	AX934434	**CAF04170.1**		

FIGURE 3N

Protein	Organism	EC#	GenBank	GenPept	SwissProt	PDB / 3D
sialyltransferase II (CstII)	*jejuni O:4*					
α-2,3/α-2,8-sialyltransferase II (CstII)	*Campylobacter jejuni O:41*	n.d.	- - AX934429	AAO96670.1 AAT17967.1 **CAF04168.1**		
α-2,3-sialyltransferase cst-I	*Campylobacter jejuni OH4384*	2.4.99.-	AF130466 -	**AAF13495.1** AAS36261.1	Q9RGF1	
bifunctional α-2,3/-2,8-sialyltransferase (Cst-II)	*Campylobacter jejuni OH4384*	2.4.99.-	AF130984 AX934425	**AAF31771.1** CAF04166.1	1RO7 1RO8	C A
HI0352 (fragment)	*Haemophilus influenzae Rd*	n.d.	U32720 X57315 NC_000907	AAC22013.1 CAA40567.1 NP_438516.1	**P24324**	
PM1174	*Pasteurella multocida PM70*	n.d.	AE006157 NC_002663	**AAK03258.1** NP_246111.1	Q9CLP3	
Sequence 10 from patent US 6503744	*Unknown.*	n.d.	-	**AAO96672.1**		
Sequence 10 from patent US 6699705	*Unknown.*	n.d.	-	**AAT17969.1**		
Sequence 12 from patent US 6699705	*Unknown.*	n.d.	-	**AAT17970.1**		
Sequence 2 from patent US 6709834	*Unknown.*	n.d.	-	**AAT23232.1**		
Sequence 3 from patent US 6503744	*Unknown.*	n.d.	-	**AAO96668.1**		
Sequence 3 from patent US 6699705	*Unknown.*	n.d.	-	**AAT17965.1**		
Sequence 34 from patent US 6503744	*Unknown.*	n.d.	-	**AAO96684.1**		
Sequence 35 from patent US 6503744 (fragment)	*Unknown.*	n.d.	- -	**AAO96685.1** AAS36262.1		
Sequence 48 from patent US 6699705	*Unknown.*	n.d.	-	**AAT17988.1**		
Sequence 5 from patent US 6699705	*Unknown.*	n.d.	-	**AAT17966.1**		
Sequence 9 from patent US 6503744	*Unknown.*	n.d.	-	**AAO96671.1**		

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FIGURE 4A

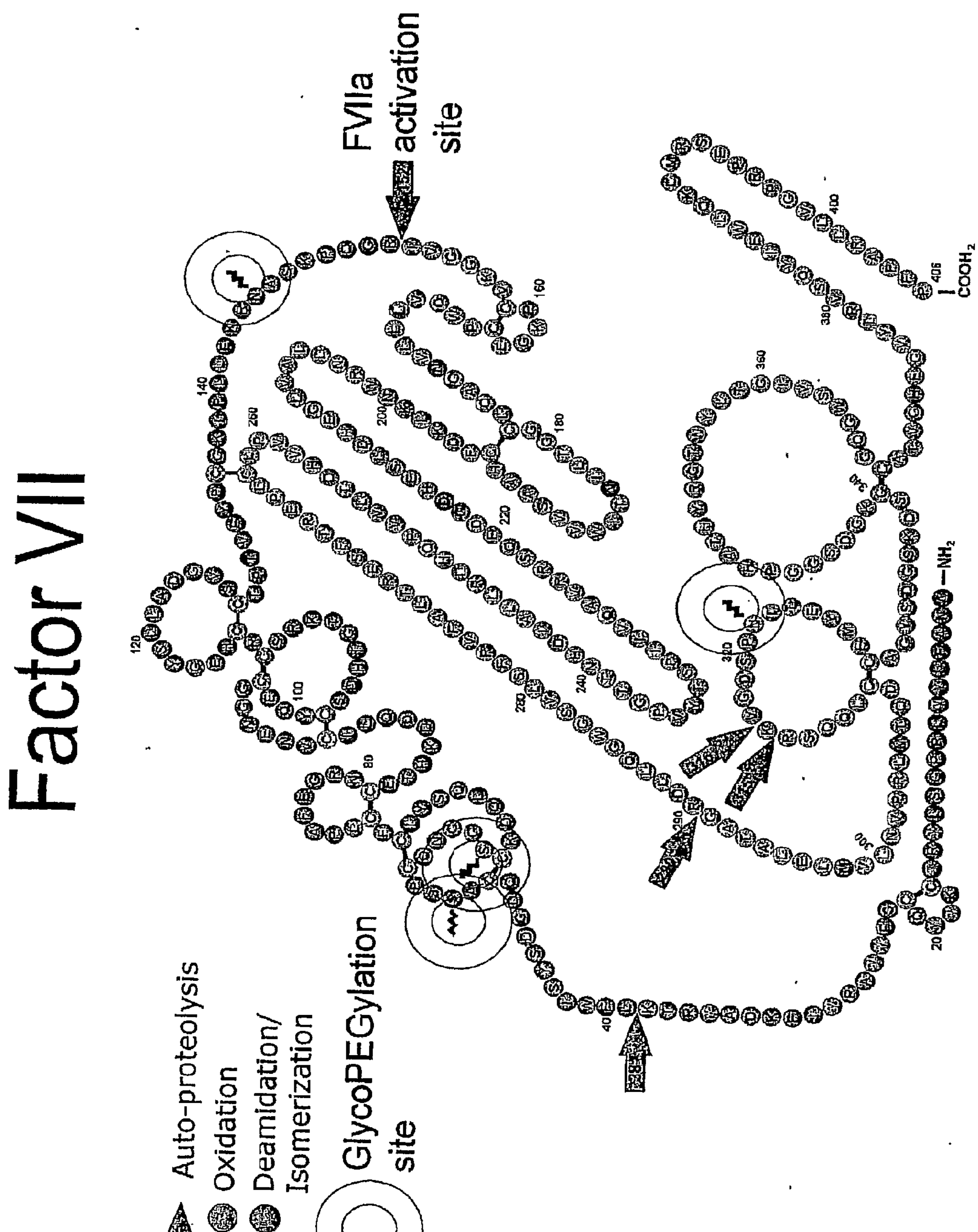

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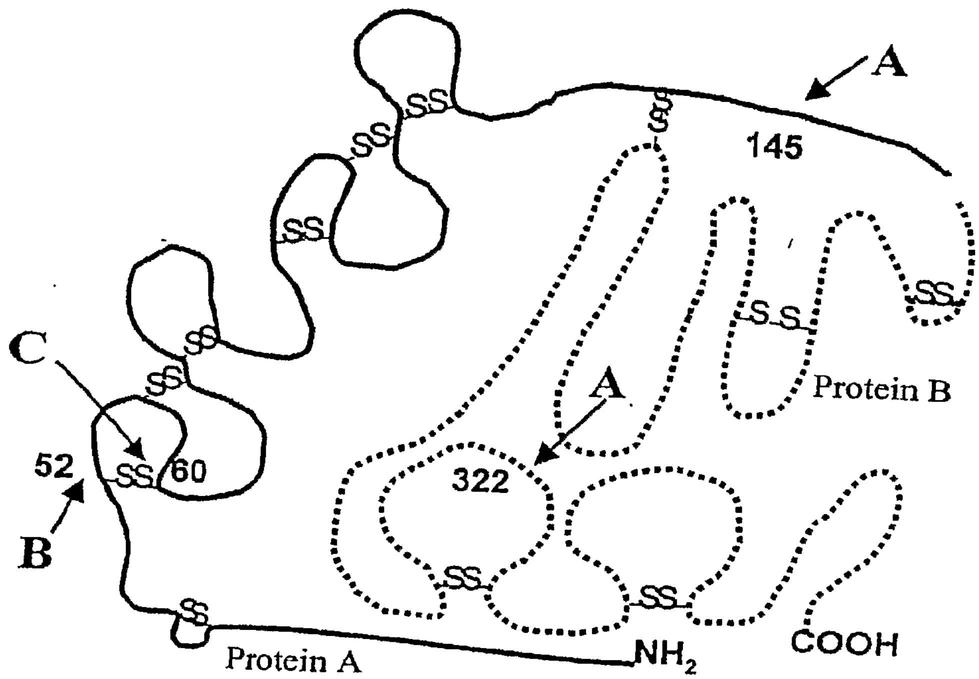
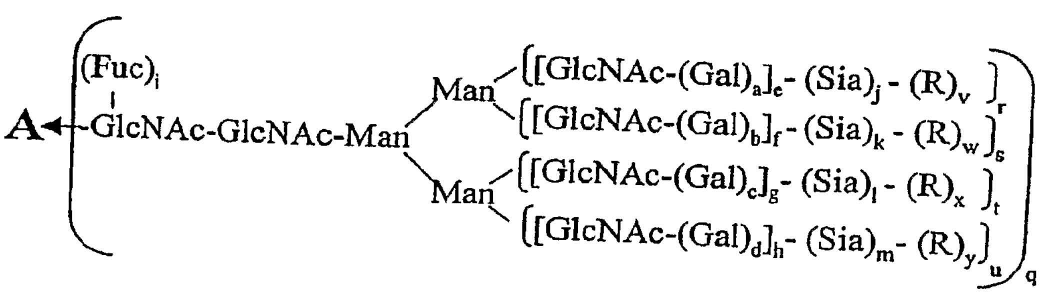
FIGURE 4B





$\mathbf{B} \leftarrow \{\text{Glc-(Xyl)}_n\}_o$

$\mathbf{C} \leftarrow \{\text{-Fuc}\}_p$

a-d, i, q-u (independently selected) = 0 or 1.
o, p (independently selected) = 0 or 1.
e-h, n (independently selected) = 0 to 6.
j-m (independently selected) = 0 to 20.
v-y = 0;
R = modifying group, mannose, oligo-
saccharide, Sia-Lewis X, Sia-Lewis A..

FIGURE 4C

BHK expressed Factor VII or VIIa
a-d, e, i, g, q, j, l, o, p (independently selected) = 0 or 1;
r, t = 1; f, h, k, m, s, u, v-y = 0; n = 0-4.

↓
1. Sialidase
2. CMP-SA-PEG (16 mole eq), ST3Gal3

a-d, e, g, i, q, j, l, o, p (independently selected) = 0 or 1;
r, t = 1; f, h, k, m, s, u, w, y = 0; n = 0-4;
v, x, (independently selected) = 1,
when j, l (respectively, independently selected) is 1;
R = PEG.

FIGURE 4D

CHO, BHK, 293 cells, Vero expressed Factor VII or VIIa
a-d, e, i, g, q, j, l, o, p (independently selected) = 0 or 1;
r, t = 1; f, h, k, m, s, u, v-y = 0; n = 0-4.

↓
1. Sialidase
2. CMP-SA-PEG (1.2 mole eq), ST3Gal3
3. CMP-SA (8 mol eq), ST3Gal3

a-d, e, g, i, q, j, l, o, p (independently selected) = 0 or 1;
r, t = 1; f, h, k, m, s, u, w, y = 0; n = 0-4;
v or x, (independently selected) = 1,
when j or l, (respectively, independently selected) is 1;
R = PEG.

FIGURE 4E

NSO expressed Factor VII or VIIa
a--u (independently selected) = 0 or 1;
v-y = 0; n = 0-4;
Sia (independently selected) = Sia or Gal.

1. Sialidase and α-galactosidase
2. Galactosyltransferase, UDP-Gal
3. CMP-SA-PEG, ST3Gal3

a-m, o-u (independently selected) = 0 or 1;
n = 0-4; v-y (independently selected) = 1,
when j-m (independently selected) is 1;
Sia = Sia; R = PEG.

FIGURE 5A

ATGGTCTCCCAGGCCCTCAGGCTCCTCTGCCTTCTGCTTGGGCTTCAG
GGCTGCCTGGCTGCAGTCTTCGTAACCCAGGAGGAAGCCCACGGCGT
CCTGCACCGGCGCCGGCGCGCCAACGCGTTCCTGGAGGAGCTGCGGC
CGGGCTCCCTGGAGAGGGAGTGCAAGGAGGAGCAGTGCTCCTTCGA
GGAGGCCCGGGAGATCTTCAAGGACGCGGAGAGGACGAAGCTGTTC
TGGATTTCTTACAGTGATGGGGACCAGTGTGCCTCAAGTCCATGCCA
GAATGGGGGCTCCTGCAAGGACCAGCTCCAGTCCTATATCTGCTTCT
GCCTCCCTGCCTTCGAGGGCCGGAACTGTGAGACGCACAAGGATGAC
CAGCTGATCTGTGTGAACGAGAACGGCGGCTGTGAGCAGTACTGCAG
TGACCACACGGGCACCAAGCGCTCCTGTCGGTGCCACGAGGGGTACT
CTCTGCTGGCAGACGGGGTGTCCTGCACACCCACAGTTGAATATCCA
TGTGGAAAAATACCTATTCTAGAAAAAAGAAATGCCAGCAAACCCCA
AGGCCGAATTGTGGGGGGCAAGGTGTGCCCCAAAGGGGAGTGTCCA
TGGCAGGTCCTGTTGTTGGTGAATGGAGCTCAGTTGTGTGGGGGGAC
CCTGATCAACACCATCTGGGTGGTCTCCGCGGCCCACTGTTTCGACAA
AATCAAGAACTGGAGGAACCTGATCGCGGTGCTGGGCGAGCACGAC
CTCAGCGAGCACGACGGGGATGAGCAGAGCCGGCGGGTGGCGCAGG
TCATCATCCCCAGCACGTACGTCCCGGGCACCACCAACCACGACATC
GCGCTGCTCCGCCTGCACCAGCCCGTGGTCCTCACTGACCATGTGGTG
CCCCTCTGCCTGCCCGAACGGACGTTCTCTGAGAGGACGCTGGCCTTC
GTGCGCTTCTCATTGGTCAGCGGCTGGGGCCAGCTGCTGGACCGTGG
CGCCACGGCCCTGGAGCTCATGGTGCTCAACGTGCCCCGGCTGATGA
CCCAGGACTGCCTGCAGCAGTCACGGAAGGTGGGAGACTCCCCAAAT
ATCACGGAGTACATGTTCTGTGCCGGCTACTCGGATGGCAGCAAGGA
CTCCTGCAAGGGGGACAGTGGAGGCCCACATGCCACCCACTACCGGG
GCACGTGGTACCTGACGGGCATCGTCAGCTGGGGCCAGGGCTGCGCA
ACCGTGGGCCACTTTGGGGTGTACACCAGGGTCTCCCAGTACATCGA
GTGGCTGCAAAAGCTCATGCGCTCAGAGCCACGCCCAGGAGTCCTCC
TGCGAGCCCCATTTCCC

FIGURE 5B

Met Val Ser Gln Ala Leu Arg Leu Leu Cys Leu Leu Leu Gly Leu Gln Gly Cys
Leu Ala Ala Val Phe Val Thr Gln Glu Glu Ala His Gly Val Leu His Arg Arg Arg
Arg Ala Asn Ala Phe Leu Glu Glu Leu Arg Pro Gly Ser Leu Glu Arg Glu Cys
Lys Glu Glu Gln Cys Ser Phe Glu Glu Ala Arg Glu Ile Phe Lys Asp Ala Glu Arg
Thr Lys Leu Phe Trp Ile Ser Tyr Ser Asp Gly Asp Gln Cys Ala Ser Ser Pro Cys
Gln Asn Gly Gly Ser Cys Lys Asp Gln Leu Gln Ser Tyr Ile Cys Phe Cys Leu Pro
Ala Phe Glu Gly Arg Asn Cys Glu Thr His Lys Asp Asp Gln Leu Ile Cys Val
Asn Glu Asn Gly Gly Cys Glu Gln Tyr Cys Ser Asp His Thr Gly Thr Lys Arg
Ser Cys Arg Cys His Glu Gly Tyr Ser Leu Leu Ala Asp Gly Val Ser Cys Thr Pro
Thr Val Glu Tyr Pro Cys Gly Lys Ile Pro Ile Leu Glu Lys Arg Asn Ala Ser Lys
Pro Gln Gly Arg Ile Val Gly Gly Lys Val Cys Pro Lys Gly Glu Cys Pro Trp Gln
Val Leu Leu Leu Val Asn Gly Ala Gln Leu Cys Gly Gly Thr Leu Ile Asn Thr Ile
Trp Val Val Ser Ala Ala His Cys Phe Asp Lys Ile Lys Asn Trp Arg Asn Leu Ile
Ala Val Leu Gly Glu His Asp Leu Ser Glu His Asp Gly Asp Glu Gln Ser Arg
Arg Val Ala Gln Val Ile Ile Pro Ser Thr Tyr Val Pro Gly Thr Thr Asn His Asp
Ile Ala Leu Leu Arg Leu His Gln Pro Val Val Leu Thr Asp His Val Val Pro Leu
Cys Leu Pro Glu Arg Thr Phe Ser Glu Arg Thr Leu Ala Phe Val Arg Phe Ser
Leu Val Ser Gly Trp Gly Gln Leu Leu Asp Arg Gly Ala Thr Ala Leu Glu Leu
Met Val Leu Asn Val Pro Arg Leu Met Thr Gln Asp Cys Leu Gln Gln Ser Arg
Lys Val Gly Asp Ser Pro Asn Ile Thr Glu Tyr Met Phe Cys Ala Gly Tyr Ser Asp
Gly Ser Lys Asp Ser Cys Lys Gly Asp Ser Gly Gly Pro His Ala Thr His Tyr Arg
Gly Thr Trp Tyr Leu Thr Gly Ile Val Ser Trp Gly Gln Gly Cys Ala Thr Val Gly
His Phe Gly Val Tyr Thr Arg Val Ser Gln Tyr Ile Glu Trp Leu Gln Lys Leu Met
Arg Ser Glu Pro Arg Pro Gly Val Leu Leu Arg Ala Pro Phe Pro

FIGURE 6

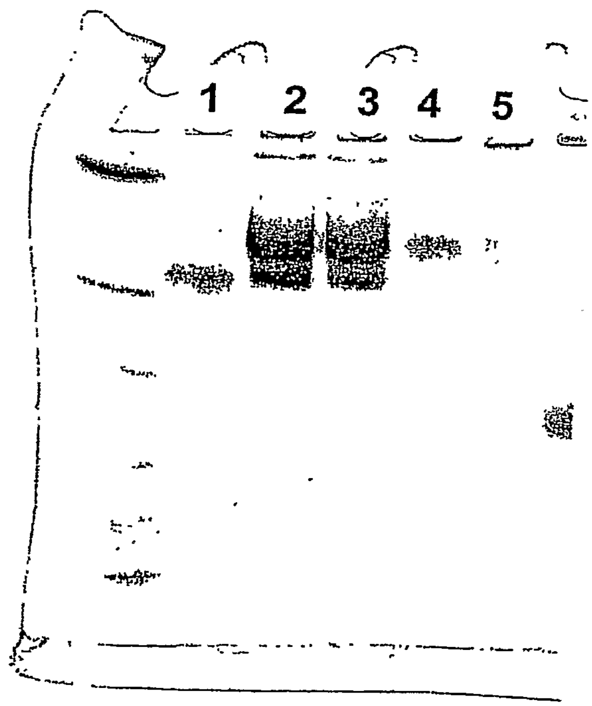


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FIGURE 7

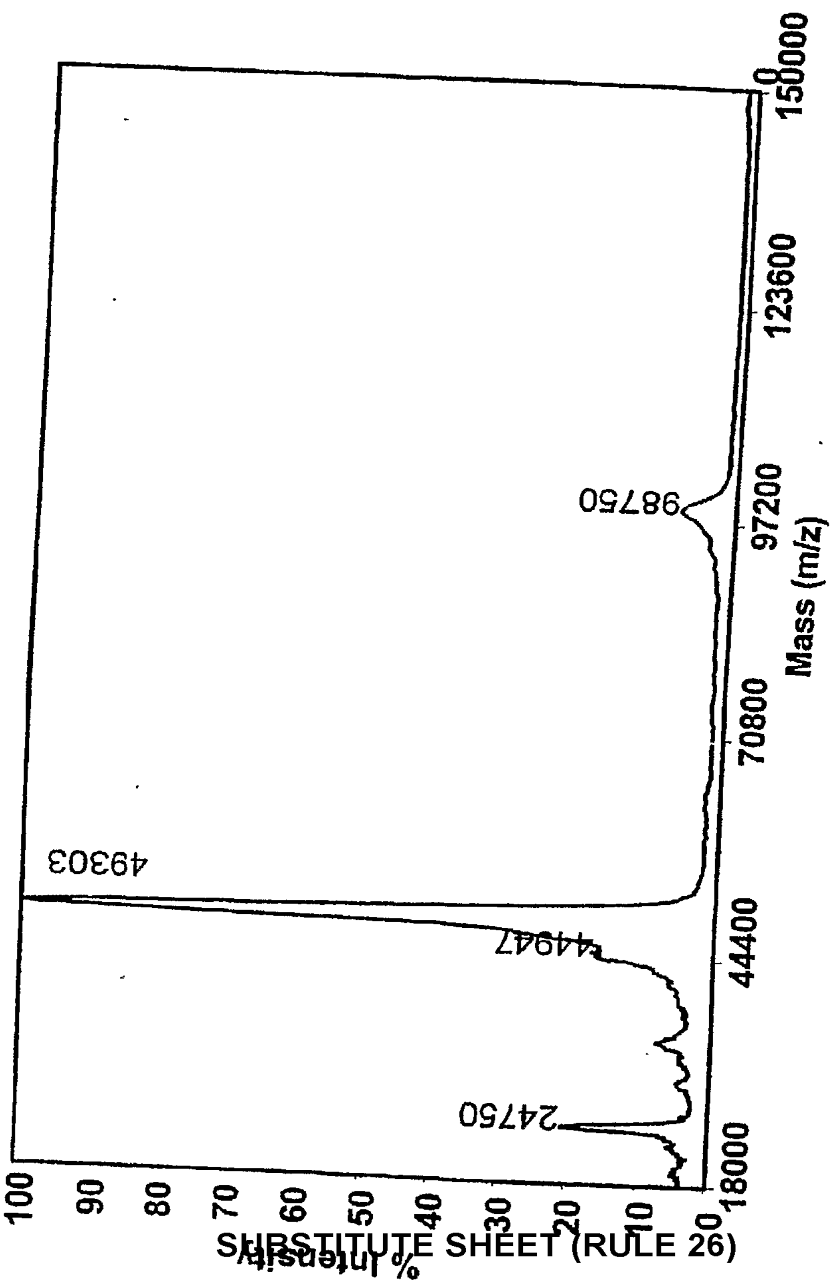

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FIGURE 8

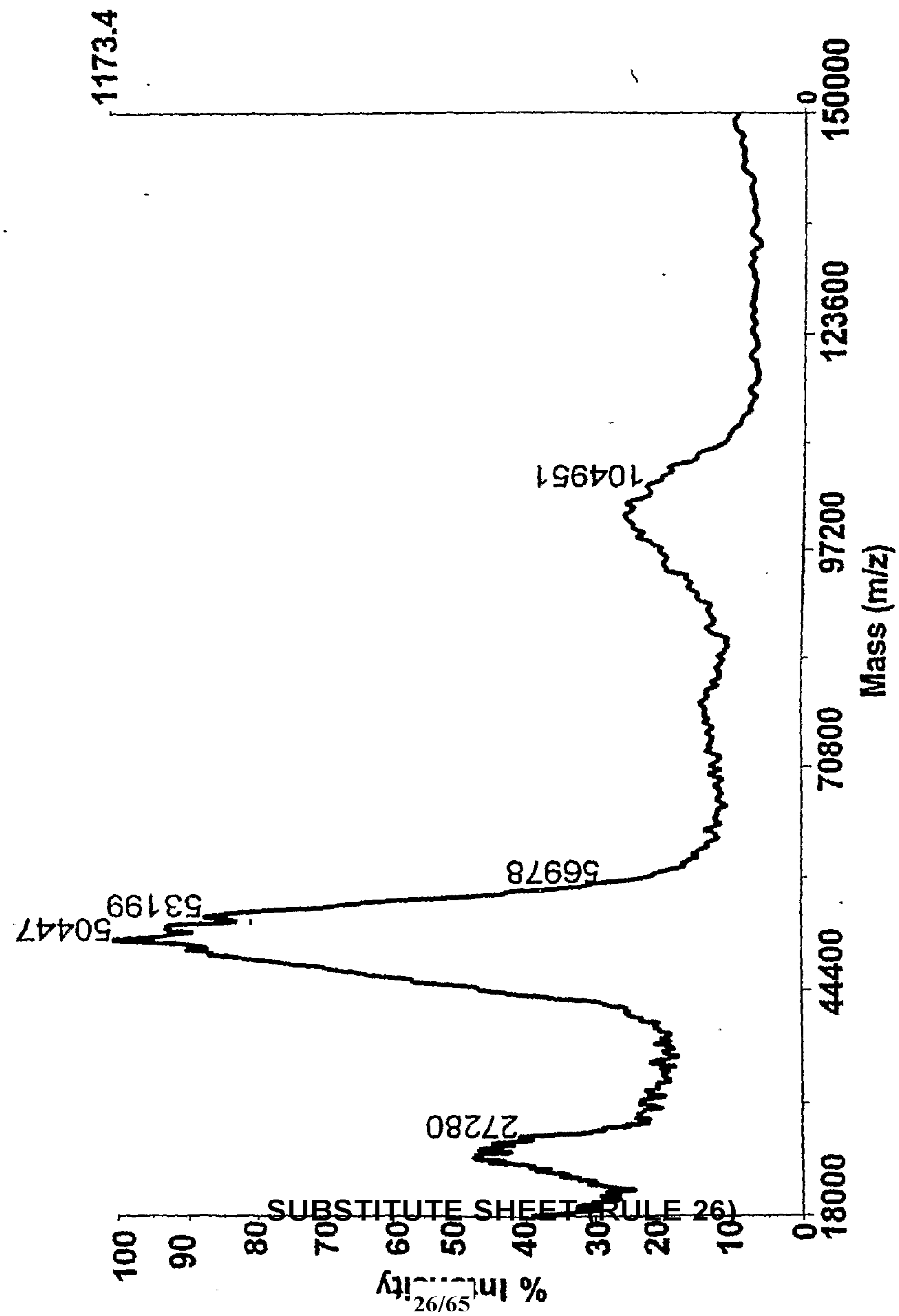

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FIGURE 9

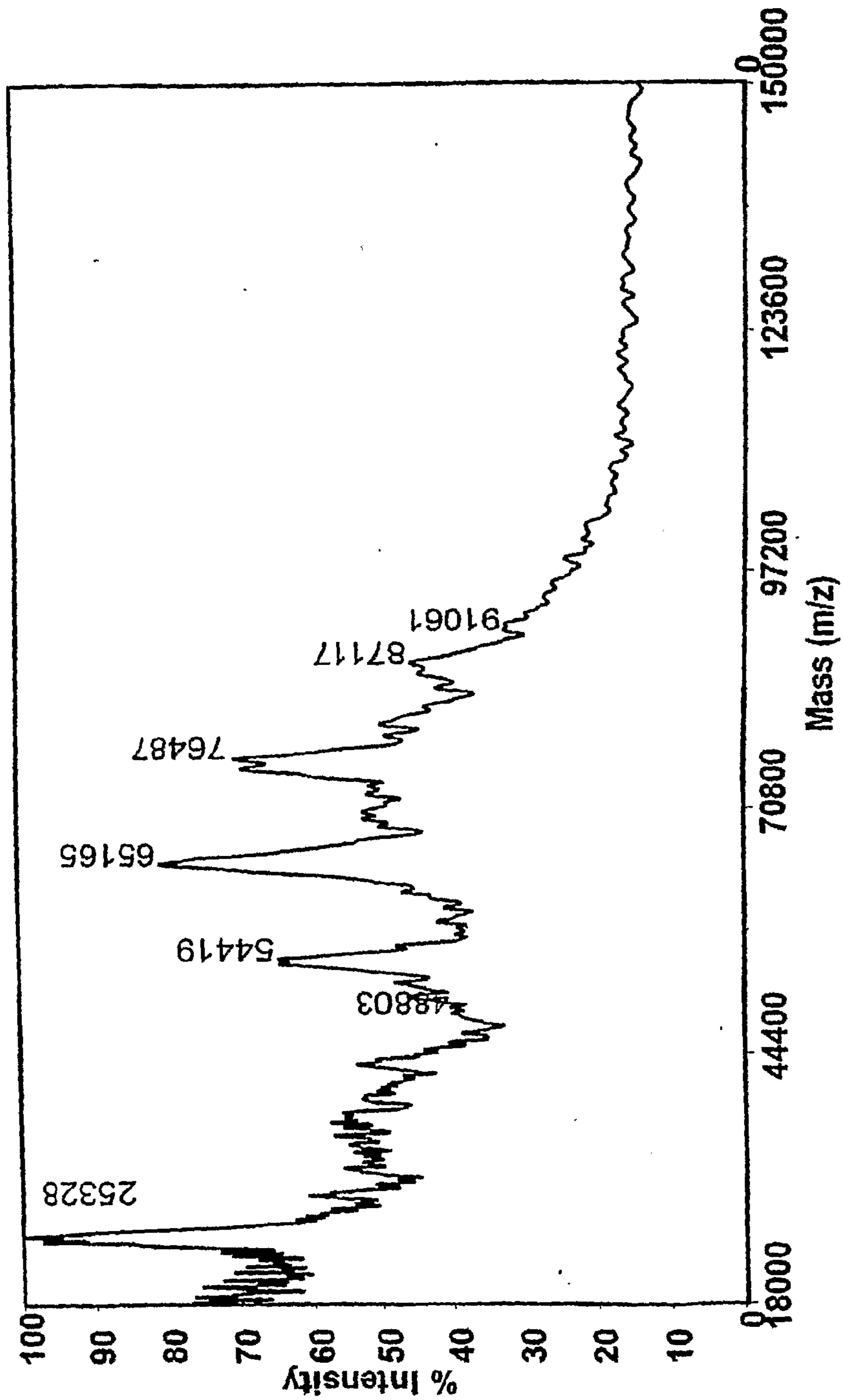

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FIGURE 10

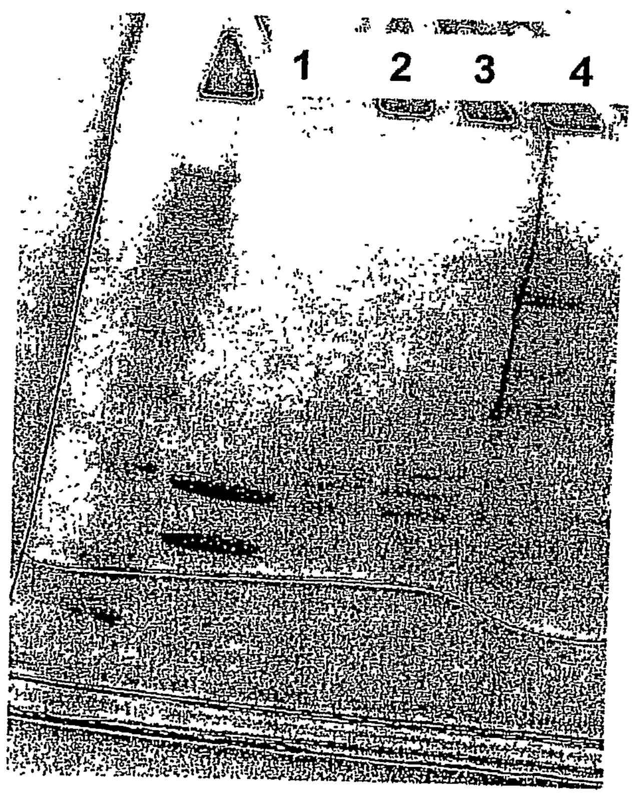

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FIGURE 11A

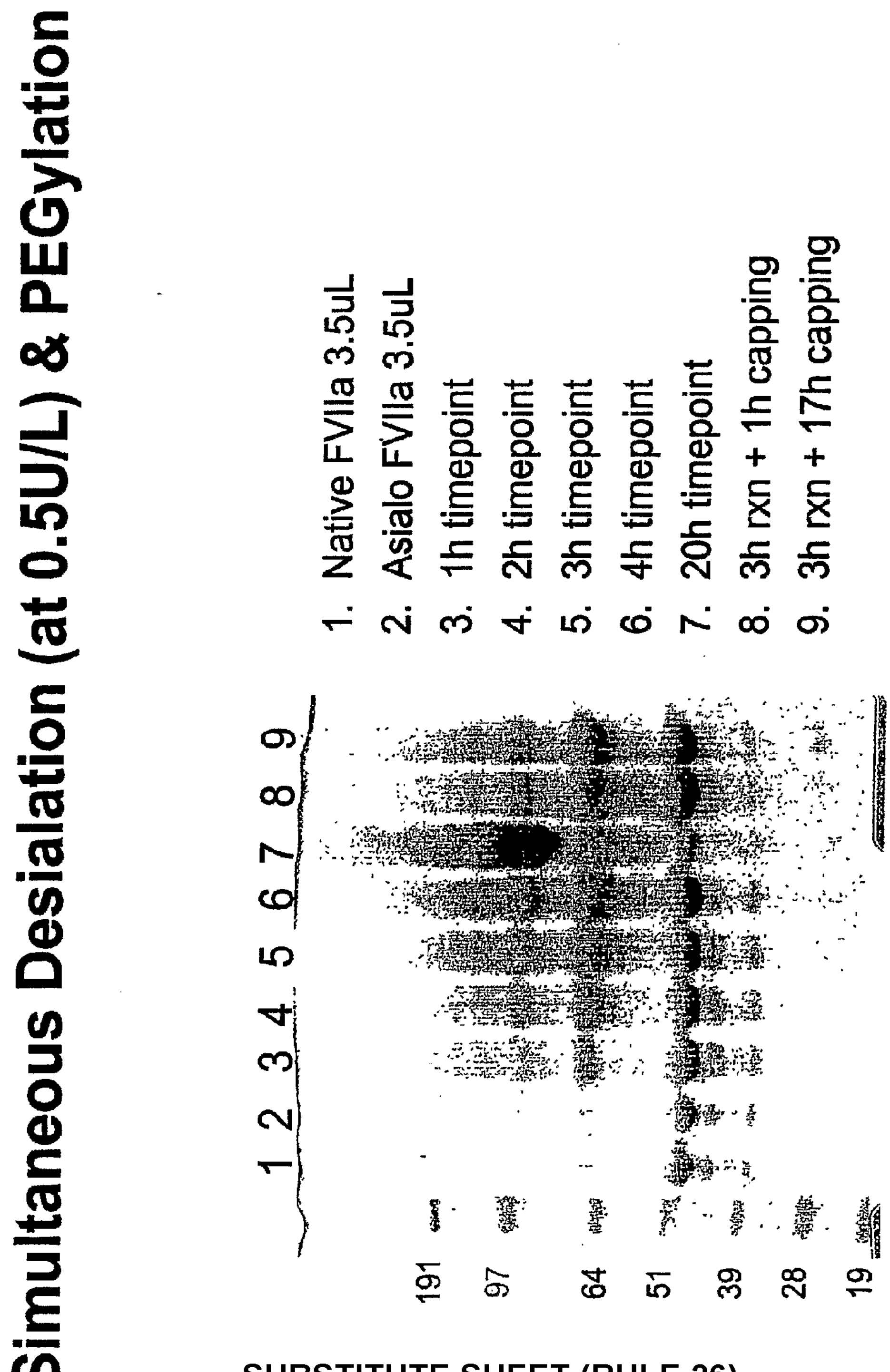

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FIGURE 11B

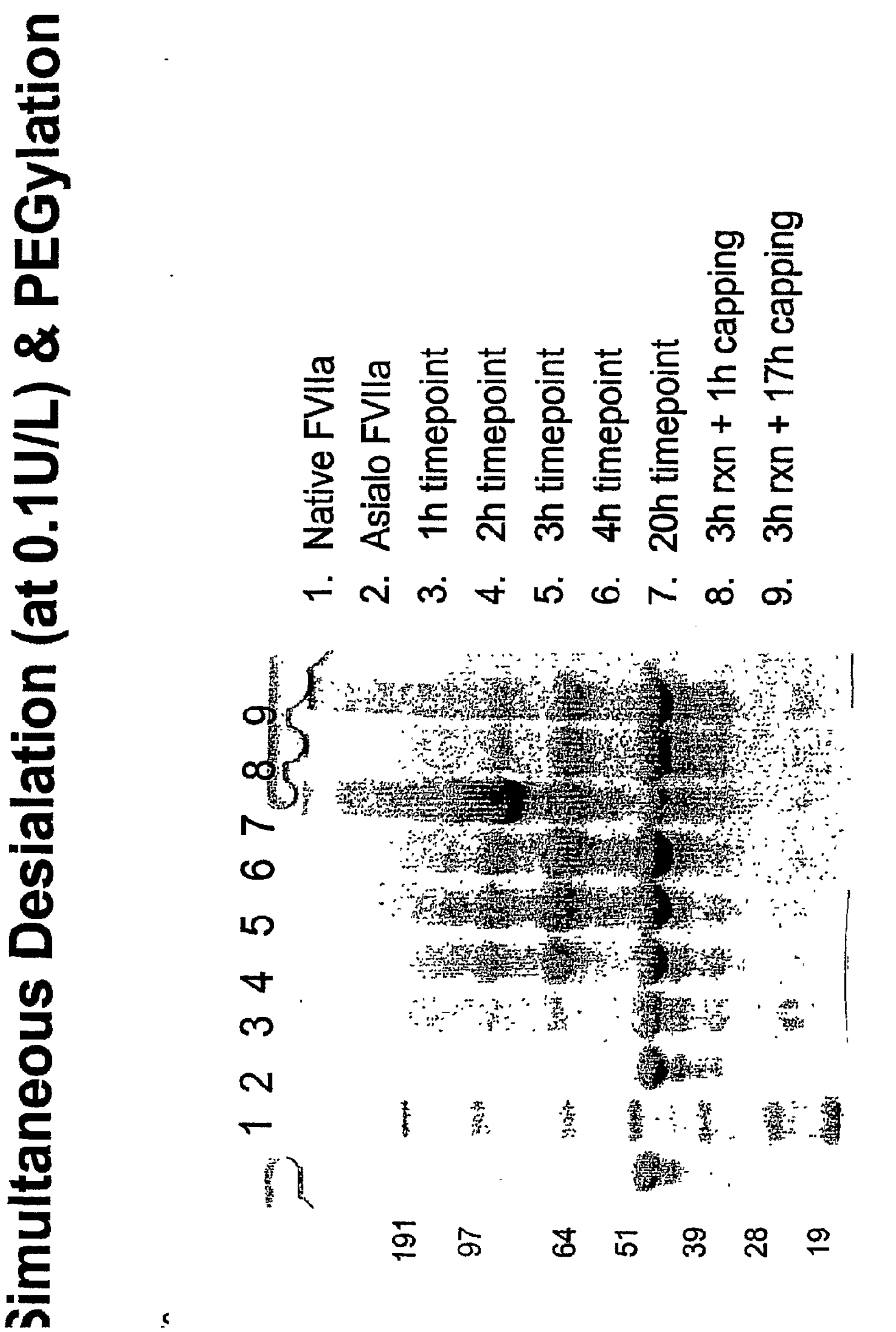

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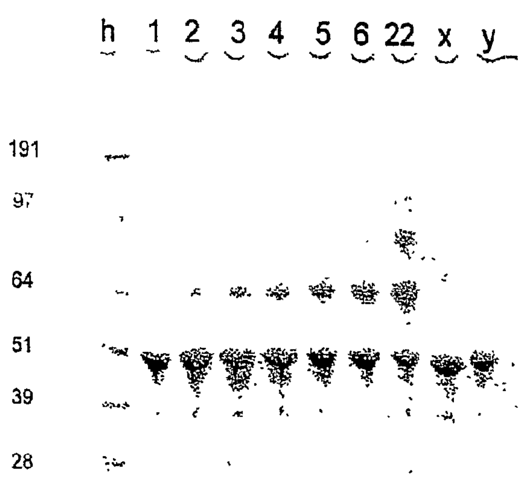
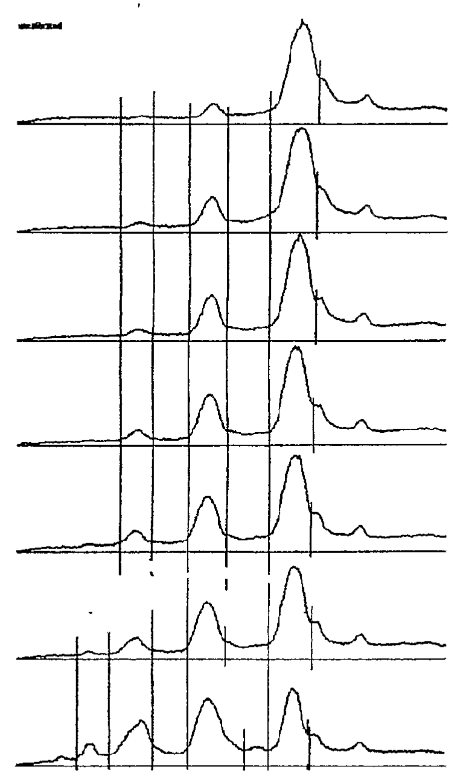
FIGURE 12A

Sample1A

33U/L ST3Gal3 A. niger / 0.2mM 10K CMP-SA-PEG



x – asialo FVIIa
y – native FVIIa

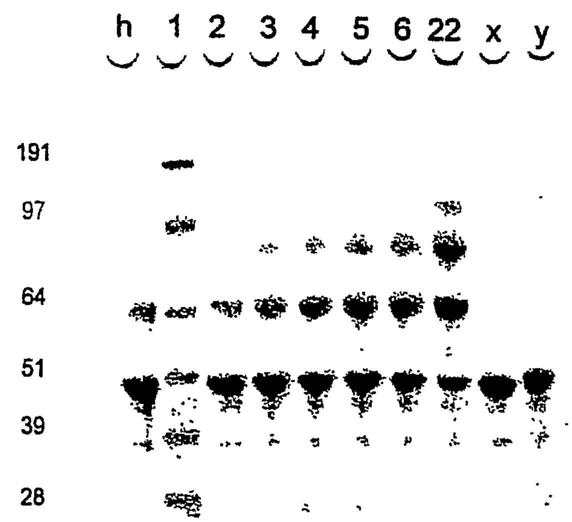


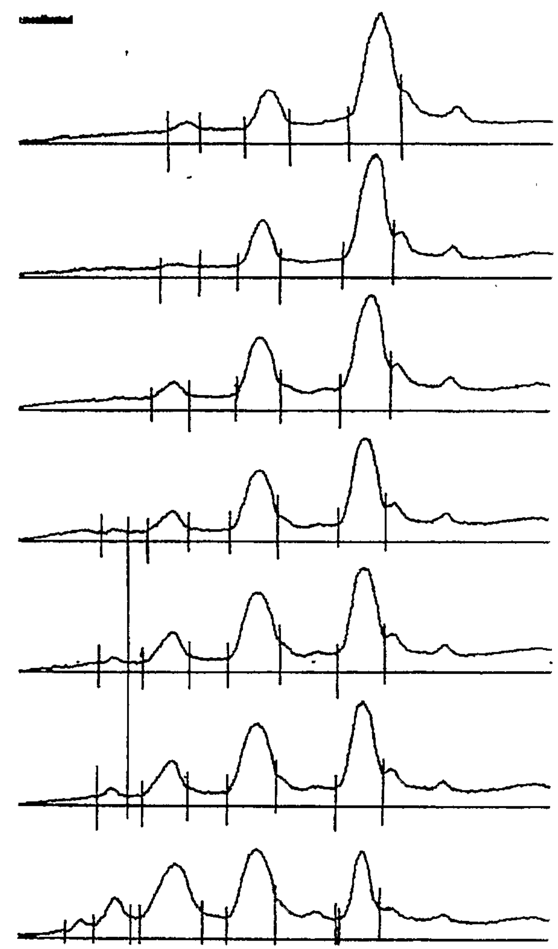
1A	% Area						
	1h	2h	3h	4h	5h	6h	22h
native FVIIa	81.0	74.2	68.8	63.0	57.6	53.1	31.9
1xPEG	13.0	19.5	24.4	28.7	31.5	30.4	38.1
2xPEG	6.0	6.4	6.8	8.3	10.9	12.3	21.8
3xPEG	0.0	0.0	0.0	0.0	0.0	4.2	8.1
4xPEG	0.0	0.0	0.0	0.0	0.0	0.0	0.0
5xPEG	0.0	0.0	0.0	0.0	0.0	0.0	0.0

FIGURE 12B

Sample1B

33U/L ST3Gal3 A. niger / 0.2mM 10K CMP-SA-PEG
30min delayed PEGylation





1B	% Area						
	1h	2h	3h	4h	5h	6h	22h
native FVIIa	66.6	65.4	55.2	47.6	42.5	39.9	22.0
1xPEG	24.9	26.7	32.9	34.5	36.8	36.6	31.5
2xPEG	8.5	7.9	11.9	13.6	16.1	17.9	32.0
3xPEG	0.0	0.0	0.0	4.4	4.7	5.5	10.6
4xPEG	0.0	0.0	0.0	0.0	0.0	0.0	3.9
5xPEG	0.0	0.0	0.0	0.0	0.0	0.0	0.0

FIGURE 13A

12AP1/E5 -- Viventia Biotech
1964 -- Aventis
20K growth hormone -- AMUR
28P6/E6 -- Viventia Biotech
3-Hydroxyphthaloyl-beta-lactoglobulin –
4-IBB ligand gene therapy –
64-Cu MAb conjugate TETA-1A3 -- Mallinckrodt Institute of Radiology
64-Cu MAb conjugate TETA-cT84.66
64-Cu Trastuzumab TETA conjugate – Genentech
A 200 -- Amgen
A10255 – Eli Liliy
A1PDX – Hedral Therapeutics
A6 -- Angstrom
aaAT-III -- Genzyme
Abciximab -- Centocor
ABI.001 – Atlantic BioPharmaceuticals
ABT-828 – Abbott
Accutin
Actinohivin
activin -- Biotech Australia, Human Therapeutics, Curis
AD 439 – Tanox
AD 519 – Tanox
Adalimumab -- Cambridge Antibody Tech.
Adenocarcinoma vaccine – Biomira -- NIS
Adenosine deanimase -- Enzond
Adenosine A2B receptor antagonists -- Adenosine Therapeutics
ADP-001 – Axis Genetics
AF 13948 – Affymax
Afelimomab – Knoll
AFP-SCAN – Immunomedics
AG 2195 – Corixa
agalsidase alfa -- Transkaryotic Therapies
agalsidase beta -- Genzyme
AGENT– Antisoma
AI 300 – AutoImmune
AI-101 – Teva
AI-102 – Teva
AI-201 – AutoImmune
AI-301 – AutoImmune
AIDS vaccine – ANRS, CIBG, Hesed Biomed, Hollis-Eden, Rome, United Biomedical, American Home Products, Maxygen
airway receptor ligand -- IC Innovations
AJvW 2 -- Ajinomoto
AK 30 NGF -- Alkermes
Albuferon -- Human Genome Sciences
albumin – Biogen, DSM Anti-Infectives, Genzyme Transgenics, PPL Therapeutics, TranXenoGen, Welfide Corp.
aldesleukin -- Chiron
alefacept -- Biogen
Alemtuzumab
Allergy therapy -- ALK-Abello/Maxygen, ALK-Abello/RP Scherer
allergy vaccines -- Allergy Therapeutics
Alnidofibatide -- Aventis Pasteur
Alnorine -- SRC VB VECTOR
ALP 242 -- Gruenenthal
Alpha antitrypsin -- Arriva/Hyland Immuno/ProMetic/Protease Sciences
Alpha-1 antitrypsin – Cutter, Bayer, PPL Therapeutics, Profile, ZymoGenetics, Arriva
Alpha-1 protease inhibitor -- Genzyme Transgenics, Welfide Corp.
Alpha-galactose fusion protein – Immunomedics
Alpha-galactosidase A -- Research Corporation Technologies, Genzyme
Alpha-glucosidase – Genzyme, Novazyme
Alpha-lactalbumin
Alpha-L-iduronidase -- Transkaryotic Therapies, BioMarin
alteplase -- Genentech
alvircept sudotox – NIH
ALX-0600, a GLP-2 agonist -- NPS Allelix Corp.

FIGURE 13B

ALX1-11 –sNPS Pharmaceuticals
Alzheimer's disease gene therapy
AM-133 -- AMRAD
Amb a 1 immunostim conj. -- Dynavax
AMD 3100 – AnorMED -- NIS
AMD 3465 – AnorMED -- NIS
AMD 3465 – AnorMED -- NIS
AMD Fab -- Genentech
Amediplase – Menarini, Novartis
AM-F9
Amoebiasis vaccine
Amphiregulin -- Octagene
anakinra -- Amgen
analgesic -- Nobex
ancestim -- Amgen
AnergiX.RA – Corixa, Organon
Angiocidin -- InKine
angiogenesis inhibitors -- ILEX
AngioMab – Antisoma
Angiopoietins -- Regeneron/Procter & Gamble
angiostatin -- EntreMed
Angiostatin/endostatin gene therapy -- Genetix Pharmaceuticals
angiotensin-II, topical -- Maret
Anthrax -- EluSys Therapeutics/US Army Medical Research Institute
Anthrax vaccine
Anti platelet-derived growth factor D human monoclonal antibodies -- CuraGen
Anti-17-1A MAb 3622W94 -- GlaxoSmithKline
Anti-2C4 MAb -- Genentech
anti-4-1BB monoclonal antibodies -- Bristol-Myers Squibb
Anti-Adhesion Platform Tech. -- Cytovax
Anti-adipocyte MAb -- Cambridge Antibody Tech./ObeSys
antiallergics -- Maxygen
antiallergy vaccine -- Acambis
Anti-alpha-4-integrin MAb
Anti-alphavβ3 integrin MAb – Applied Molecular Evolution
Anti-angiogenesis monoclonal antibodies -- KS Biomedix/Schering AG
Anti-B4 MAb-DC1 conjugate -- ImmunoGen
Anti-B7 antibody PRIMATIZED -- IDEC
Anti-B7-1 MAb 16-10A1
Anti-B7-1 MAb 1G10
Anti-B7-2 MAb GL-1
Anti-B7-2-gelonin immunotoxin –
Antibacterials/antifungals -- Diversa/IntraBiotics
Anti-beta-amyloid monoclonal antibodies -- Cambridge Antibody Tech., Wyeth-Ayerst
Anti-BLyS antibodies -- Cambridge Antibody Tech. /Human Genome Sciences
Antibody-drug conjugates -- Seattle Genetics/Eos
Anti-C5 MAb BB5-1 -- Alexion
Anti-C5 MAb N19-8 -- Alexion
Anti-C8 MAb
anticancer cytokines -- BioPulse
anticancer matrix – Telios Integra
Anticancer monoclonal antibodies – ARIUS, Immunex
anticancer peptides – Maxygen, Micrologix
Anticancer prodrug Tech. -- Alexion Antibody Technologies
anticancer Troy-Bodies -- Affite -- Affitech
anticancer vaccine -- NIH
anticancers -- Epimmune
Anti-CCR5/CXCR4 sheep MAb -- KS Biomedix Holdings
Anti-CD11a MAb KBA –
Anti-CD11a MAb M17
Anti-CD11a MAb TA-3 –
Anti-CD11a MAb WT.1 –
Anti-CD11a MAb -- Pharmacia

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FIGURE 13C

Anti-CD11b MAb LM2
Anti-CD154 MAb -- Biogen
Anti-CD16-anti-CD30 MAb -- Biotest
Anti-CD18 MAb -- Pharmacia
Anti-CD19 MAb B43 –
Anti-CD19 MAb -liposomal sodium butyrate conjugate –
Anti-CD147
Anti-CD19 MAb-saporin conjugate –
Anti-CD19-dsFv-PE38-immunotoxin –
Anti-CD2 MAb 12-15 –
Anti-CD2 MAb B-E2 -- Diaclone
Anti-CD2 MAb OX34 –
Anti-CD2 MAb OX54 –
Anti-CD2 MAb OX55 –
Anti-CD2 MAb RM2-1
Anti-CD2 MAb RM2-2
Anti-CD2 MAb RM2-4
Anti-CD20 MAb BCA B20
Anti-CD20-anti-Fc alpha RI bispecific MAb – Medarex, Tenovus
Anti-CD22 MAb-saporin-6 complex –
Anti-CD3 immunotoxin –
Anti-CD3 MAb 145-2C11 -- Pharming
Anti-CD3 MAb CD4IgG conjugate -- Genentech
Anti-CD3 MAb humanised – Protein Design, RW Johnson
Anti-CD3 MAb WT32
Anti-CD3 MAb-ricin-chain-A conjugate –
Anti-CD3 MAb-xanthine-oxidase conjugate –
Anti-CD30 MAb BerH2 -- Medac
Anti-CD30 MAb-saporin conjugate
Anti-CD30-scFv-ETA'-immunotoxin
Anti-CD38 MAb AT13/5
Anti-CD38 MAb-saporin conjugate
Anti-CD3-anti-CD19 bispecific MAb
Anti-CD3-anti-EGFR MAb
Anti-CD3-anti-interleukin-2-receptor MAb
Anti-CD3-anti-MOv18 MAb -- Centocor
Anti-CD3-anti-SCLC bispecific MAb
Anti-CD4 idiotype vaccine
Anti-CD4 MAb – Centocor, IDEC Pharmaceuticals, Xenova Group
Anti-CD4 MAb 16H5
Anti-CD4 MAb 4162W94 -- GlaxoSmithKline
Anti-CD4 MAb B-F5 -- Diaclone
Anti-CD4 MAb GK1-5
Anti-CD4 MAb KT6
Anti-CD4 MAb OX38
Anti-CD4 MAb PAP conjugate -- Bristol-Myers Squibb
Anti-CD4 MAb RIB 5-2
Anti-CD4 MAb W3/25
Anti-CD4 MAb YTA 3.1.2
Anti-CD4 MAb YTS 177-9
Anti-CD40 ligand MAb 5c8 -- Biogen
Anti-CD40 MAb
Anti-CD40 MAb 5D12 – Tanox
Anti-CD44 MAb A3D8
Anti-CD44 MAb GKWA3
Anti-CD44 MAb IM7
Anti-CD44 MAb KM81
Anti-CD44 variant monoclonal antibodies -- Corixa/Hebrew University
Anti-CD45 MAb BC8-I-131
Anti-CD45RB MAb
Anti-CD48 MAb HuLy-m3
Anti-CD48 MAb WM-63
Anti-CD5 MAb -- Becton Dickinson
Anti-CD5 MAb OX19
Anti-CD6 MAb
Anti-CD7 MAb-PAP conjugate
Anti-CD7 MAb-ricin-chain-A conjugate
Anti-CD8 MAb – Amerimmune, Cytodyn, Becton Dickinson
Anti-CD8 MAb 2-43
Anti-CD8 MAb OX8
Anti-CD80 MAb P16C10 -- IDEC
Anti-CD80 MAb P7C10 -- ID Vaccine
Anti-CD8-idarubicin conjugate
Anti-CEA MAb CE-25

FIGURE 13D

Anti-CEA MAb MN 14 – Immunomedics
Anti-CEA MAb MN14-PE40 conjugate – Immunomedics
Anti-CEA MAb T84.66-interleukin-2 conjugate
Anti-CEA sheep MAb -- KS Biomedix Holdings
Anti-cell surface monoclonal antibodies -- Cambridge Antibody Tech. /Pharmacia
Anti-c-erbB2-anti-CD3 bifunctional MAb -- Otsuka
Anti-CMV MAb -- Scotgen
Anti-complement
Anti-CTLA-4 MAb
Anti-EGFR catalytic antibody -- Hesed Biomed
anti-EGFR immunotoxin -- IVAX
Anti-EGFR MAb -- Abgenix
Anti-EGFR MAb 528
Anti-EGFR MAb KSB 107 -- KS Biomedix
Anti-EGFR MAb-DM1 conjugate -- ImmunoGen
Anti-EGFR MAb-LA1 –
Anti-EGFR sheep MAb -- KS Biomedix
Anti-FAP MAb F19-I-131
Anti-Fas IgM MAb CH11
Anti-Fas MAb Jo2
Anti-Fas MAb RK-8
Anti-Flt-1 monoclonal antibodies -- ImClone
Anti-fungal peptides -- State University of New York
antifungal tripeptides -- BTG
Anti-ganglioside GD2 antibody-interleukin-2 fusion protein -- Lexigen
Anti-GM2 MAb -- Kyowa
Anti-GM-CSF receptor monoclonal antibodies -- AMRAD
Anti-gp130 MAb -- Tosoh
Anti-HCA monoclonal antibodies -- AltaRex/Epigen
Anti-hCG antibodies -- Abgenix/AVI BioPharma
Anti-heparanase human monoclonal antibodies -- Oxford Glycosciences/Medarex
Anti-hepatitis C virus human monoclonal antibodies -- XTL Biopharmaceuticals
Anti-HER-2 antibody gene therapy
Anti-herpes antibody -- Epicyte
Anti-HIV antibody -- Epicyte
anti-HIV catalytic antibody -- Hesed Biomed
anti-HIV fusion protein -- Idun
anti-HIV proteins -- Cangene
Anti-HM1-24 MAb -- Chugai
Anti-hR3 MAb
Anti-Human-Carcinoma-Antigen MAb -- Epicyte
Anti-ICAM-1 MAb -- Boehringer Ingelheim
Anti-ICAM-1 MAb 1A-29 -- Pharmacia
Anti-ICAM-1 MAb HA58
Anti-ICAM-1 MAb YN1/1.7.4
Anti-ICAM-3 MAb ICM3 -- ICOS
Anti-idiotype breast cancer vaccine 11D10
Anti-idiotype breast cancer vaccine ACA14C5 –
Anti-idiotype cancer vaccine -- ImClone Systems/Merck KGaA ImClone, Viventia Biotech
Anti-idiotype cancer vaccine 1A7 -- Titan
Anti-idiotype cancer vaccine 3H1 -- Titan
Anti-idiotype cancer vaccine TriAb -- Titan
Anti-idiotype Chlamydia trachomatis vaccine
Anti-idiotype colorectal cancer vaccine -- Novartis
Anti-idiotype colorectal cancer vaccine -- Onyvax
Anti-idiotype melanoma vaccine -- IDEC Pharmaceuticals
Anti-idiotype ovarian cancer vaccine ACA

FIGURE 13E

Anti-idiotype ovarian cancer vaccine AR54 -- AltaRex
Anti-idiotype ovarian cancer vaccine CA-125 – AltaRex, Biomira
Anti-IgE catalytic antibody -- Hesed Biomed
Anti-IgE MAb E26 -- Genentech
Anti-IGF-1 MAb
anti-inflammatory -- GeneMax
anti-inflammatory peptide -- BTG
anti-integrin peptides -- Burnha
Anti-interferon-alpha-receptor MAb 64G12 – Pharma Pacific Management
Anti-interferon-gamma MAb -- Protein Design Labs
Anti-interferon-gamma polyclonal antibody -- Advanced Biotherapy
Anti-interleukin-10 MAb –
Anti-interleukin-12 MAb –
Anti-interleukin-1-beta polyclonal antibody -- R&D Systems
Anti-interleukin-2 receptor MAb 2A3
Anti-interleukin-2 receptor MAb 33B3-1 -- Immunotech
Anti-interleukin-2 receptor MAb ART-18
Anti-interleukin-2 receptor MAb LO-Tact-1
Anti-interleukin-2 receptor MAb Mikbeta1
Anti-interleukin-2 receptor MAb NDS61
Anti-interleukin-4 MAb 11B11
Anti-interleukin-5 MAb -- Wallace Laboratories
Anti-interleukin-6 MAb – Centocor, Diaclone, Pharmadigm
Anti-interleukin-8 MAb -- Abgenix
Anti-interleukin-8 MAb – Xenotech
Anti-JL1 MAb
Anti-Klebsiella sheep MAb -- KS Biomedix Holdings
Anti-Laminin receptor MAb-liposomal doxorubicin conjugate
Anti-LCG MAb -- Cytoclonal
Anti-lipopolysaccharide MAb -- VitaResc
Anti-L-selectin monoclonal antibodies -- Protein Design Labs, Abgenix, Stanford University
Anti-MBL monoclonal antibodies -- Alexion/Brigham and Women's Hospital
Anti-MHC monoclonal antibodies
Anti-MIF antibody humanised – IDEC, Cytokine PharmaSciences
Anti-MRSA/VRSA sheep MAb -- KS Biomedix Holdings
Anti-mu MAb -- Novartis
Anti-MUC-1 MAb
Anti-MUC 18
Anti-Nogo-A MAb IN1
Anti-nuclear autoantibodies -- Procyon
Anti-ovarian cancer monoclonal antibodies -- Dompe
Anti-p185 monoclonal antibodies
Anti-p43 MAb
Antiparasitic vaccines
Anti-PDGF/bFGF sheep MAb -- KS Biomedix
Anti-properdin monoclonal antibodies -- Abgenix/Gliatech
Anti-PSMA (prostrate specific membrane antigen)
Anti-PSMA MAb J591 -- BZL Biologics
Anti-Rev MAb gene therapy –
Anti-RSV antibodies – Epicyte, Intracell
Anti-RSV monoclonal antibodies -- Medarex/MedImmune, Applied Molecular Evolution/MedImmune
Anti-RSV MAb, inhalation -- Alkermes/MedImmune
Anti-RT gene therapy
Antisense K-ras RNA gene therapy
Anti-SF-25 MAb
Anti-sperm antibody -- Epicyte
Anti-Tac(Fv)-PE38 conjugate
Anti-TAPA/CD81 MAb AMP1
Anti-tat gene therapy
Anti-TCR-alphabeta MAb H57-597

FIGURE 13F

Anti-TCR-alphabeta MAb R73
Anti-tenascin MAb BC-4-I-131
Anti-TGF-beta human monoclonal antibodies -- Cambridge Antibody Tech., Genzyme
Anti-TGF-beta MAb 2G7 -- Genentech
Antithrombin III -- Genzyme Transgenics, Aventis, Bayer, Behringwerke, CSL, Myriad
Anti-Thy1 MAb
Anti-Thy1.1 MAb
Anti-tissue factor/factor VIIA sheep MAb -- KS Biomedix
Anti-TNF monoclonal antibodies – Centocor, Chiron, Peptech, Pharacia, Serono
Anti-TNF sheep MAb -- KS Biomedix Holdings
Anti-TNFalpha MAb -- Genzyme
Anti-TNFalpha MAb B-C7 -- Diaclone
Anti-tooth decay MAb -- Planet BioTech.
Anti-TRAIL receptor-1 MAb -- Takeda
Antitumour RNases -- NIH
Anti-VCAM MAb 2A2 -- Alexion
Anti-VCAM MAb 3F4 -- Alexion
Anti-VCAM-1 MAb
Anti-VEC MAb -- ImClone
Anti-VEGF MAb -- Genentech
Anti-VEGF MAb 2C3
Anti-VEGF sheep MAb -- KS Biomedix Holdings
Anti-VLA-4 MAb HP1/2 -- Biogen
Anti-VLA-4 MAb PS/2
Anti-VLA-4 MAb R1-2
Anti-VLA-4 MAb TA-2
Anti-VAP-1 human MAb
Anti-VRE sheep MAb -- KS Biomedix Holdings
ANUP -- TranXenoGen
ANUP-1 -- Pharis
AOP-RANTES -- Senetek
Apan-CH -- Praecis Pharmaceuticals
APC-8024 -- Demegen
ApoA-1 -- Milano, Pharmacia
Apogen -- Alexion
apolipoprotein A1 -- Avanir
Apolipoprotein E -- Bio-Tech. General
Applaggin -- Biogen
aprotinin -- ProdiGene
APT-070C -- AdProTech
AR 177 -- Aronex Pharmaceuticals
AR 209 -- Aronex Pharmaceuticals, Antigenics
AR545C
ARGENT gene delivery systems -- ARIAD
Arresten
ART-123 -- Asahi Kasei
arylsulfatase B -- BioMarin
Arylsulfatase B, Recombinant human -- BioMarin
AS 1051 -- Ajinomoto
ASI-BCL -- Intracell
Asparaginase - Merck
ATL-101 -- Alizyme
Atrial natriuretic peptide -- Pharis
Aurintricarboxylic acid-high molecular weight
Autoimmune disorders -- GPC Biotech/MorphoSys
Autoimmune disorders and transplant rejection -- Bristol-Myers Squibb/Genzyme Tra
Autoimmune disorders/cancer -- Abgenix/Chiron, CuraGen
Autotaxin
Avicidin -- NeoRx
axogenesis factor-1 -- Boston Life Sciences
Axokine -- Regeneron
B cell lymphoma vaccine -- Biomira
B7-1 gene therapy –
BABS proteins -- Chiron
BAM-002 -- Novelos Therapeutics
Basiliximab (anti-CD25 MAb) -- Novartis

FIGURE 13G

Bay-16-9996 -- Bayer
Bay-39-9437 -- Bayer
Bay-50-4798 -- Bayer
BB-10153 -- British Biotech
BBT-001 -- Bolder BioTech.
BBT-002 -- Bolder BioTech.
BBT-003 -- Bolder BioTech.
BBT-004 -- Bolder BioTech.
BBT-005 -- Bolder BioTech.
BBT-006 -- Bolder BioTech.
BBT-007 -- Bolder BioTech.
BCH-2763 -- Shire
BCSF -- Millenium Biologix
BDNF -- Regeneron – Amgen
Becaplermin -- Johnson & Johnson, Chiron
Bectumomab – Immunomedics
Beriplast -- Aventis
Beta-adrenergic receptor gene therapy -- University of Arkansas
bFGF -- Scios
BI 51013 -- Behringwerke AG
BIBH 1 -- Boehringer Ingelheim
BIM-23190 -- Beaufour-Ipsen
birch pollen immunotherapy -- Pharmacia
bispecific fusion proteins -- NIH
Bispecific MAb 2B1 -- Chiron
Bitistatin
BIWA 4 -- Boehringer Ingelheim
blood substitute – Northfield, Baxter Intl.
BLP-25 -- Biomira
BLS-0597 -- Boston Life Sciences
BLyS -- Human Genome Sciences
BLyS radiolabelled -- Human Genome Sciences
BM 06021 -- Boehringer Mannheim
BM-202 -- BioMarin
BM-301 -- BioMarin
BM-301 -- BioMarin
BM-302 -- BioMarin
BMP 2 -- Genetics Institute/Medtronic-Sofamor Danek, Genetics Institute/ Collagenesis, Genetics Institute/Yamanouch
BMP 2 gene therapy
BMP 52 -- Aventis Pasteur, Biopharm
BMP-2 -- Genetics Institute
BMS 182248 -- Bristol-Myers Squibb
BMS 202448 -- Bristol-Myers Squibb
bone growth factors -- IsoTis
BPC-15 -- Pfizer
brain natriuretic peptide –
Breast cancer -- Oxford GlycoSciences/Medarex
Breast cancer vaccine -- Therion Biologics, Oregon
BSSL -- PPL Therapeutics
BST-2001 – BioStratum
BST-3002 -- BioStratum
BTI 322 –
butyrylcholinesterase -- Shire
C 6822 -- COR Therapeutics
C1 esterase inhibitor -- Pharming
C3d adjuvant -- AdProTech
CAB-2.1 -- Millennium
calcitonin – Inhale Therapeutics Systems, Aventis, Genetronics, TranXenoGen, Unigene, Rhone Poulenc Rohrer
calcitonin -- oral – Nobex, Emisphere, Pharmaceutical Discovery
Calcitonin gene-related peptide -- Asahi Kasei -- Unigene
calcitonin, human -- Suntory
calcitonin, nasal – Novartis, Unigene
calcitonin, Panoderm -- Elan
calcitonin, Peptitrol -- Shire
calcitonin, salmon -- Therapicon
calin -- Biopharm
Calphobindin I
calphobindin I -- Kowa
calreticulin -- NYU
Campath-1

FIGURE 13H

Campath-1M
cancer therapy -- Cangene
cancer vaccine – Aixlie, Aventis Pasteur, Center of Molecular Immunology ,YM BioSciences, Cytos, Genzyme, Transgenics, GlobeImmune, Igeneon, ImClone, Virogenetics, InterCell, Iomai, Jenner Biotherapies, Memorial Sloan-Kettering Cancer Center, Sydney Kimmel Cancer Center, Novavax, Protein Sciences, Argonex, SIGA
Cancer vaccine ALVAC-CEA B7.1 -- Aventis Pasteur/Therion Biologics
Cancer vaccine CEA-TRICOM -- Aventis Pasteur/Therion Biologics
Cancer vaccine gene therapy -- Cantab Pharmaceuticals
Cancer vaccine HER-2/neu -- Corixa
Cancer vaccine THERATOPE -- Biomira
cancer vaccine, PolyMASC -- Valentis
Candida vaccine – Corixa, Inhibitex
Canstatin -- ILEX
CAP-18 -- Panorama
Cardiovascular gene therapy -- Collateral Therapeutics
carperitide -- Suntory
Casocidin-1 -- Pharis
CAT 152 -- Cambridge Antibody Tech.
CAT 192 -- Cambridge Antibody Tech.
CAT 213 -- Cambridge Antibody Tech.
Catalase-- Enzon
Cat-PAD -- Circassia
CB 0006 -- Celltech
CCK(27-32)-- Akzo Nobel
CCR2-64I -- NIH
CD, Procept -- Paligent
CD154 gene therapy
CD39 -- Immunex
CD39-L2 -- Hyseq
CD39-L4 -- Hyseq
CD4 fusion toxin -- Senetek
CD4 IgG -- Genentech
CD4 receptor antagonists -- Pharmacopeia/Progenics
CD4 soluble -- Progenics
CD4, soluble -- Genzyme Transgenics
CD40 ligand -- Immunex
CD4-ricin chain A -- Genentech
CD59 gene therapy -- Alexion
CD8 TIL cell therapy -- Aventis Pasteur
CD8, soluble -- Avidex
CD95 ligand -- Roche
CDP 571 -- Celltech
CDP 850 -- Celltech
CDP-860 (PEG-PDGF MAb) -- Celltech
CDP 870 -- Celltech
CDS-1 -- Ernest Orlando
Cedelizumab -- Ortho-McNeil
Cetermin -- Insmed
CETP vaccine -- Avant
Cetrorelix
Cetuximab
CGH 400 -- Novartis
CGP 42934 -- Novartis
CGP 51901 – Tanox
CGRP -- Unigene
CGS 27913 -- Novartis
CGS 32359 -- Novartis
Chagas disease vaccine -- Corixa
chemokines -- Immune Response
CHH 380 -- Novartis
chitinase – Genzyme, ICOS
Chlamydia pneumoniae vaccine – Antex Biologics
Chlamydia trachomatis vaccine -- Antex Biologics
Chlamydia vaccine -- GlaxoSmithKline
Cholera vaccine CVD 103-HgR -- Swiss Serum and Vaccine Institute Berne
Cholera vaccine CVD 112 -- Swiss Serum and Vaccine Institute Berne
Cholera vaccine inactivated oral -- SBL Vaccin
Chrysalin -- Chrysalis BioTech.

FIGURE 13I

CI-782 -- Hitachi Kase
Ciliary neurotrophic factor – Fidia, Roche
CIM project -- Active Biotech
CL 329753 -- Wyeth-Ayerst
CL22, Cobra -- ML Laboratories
Clenoliximab -- IDEC
Clostridium difficile antibodies -- Epicyte
clotting factors -- Octagene
CMB 401 -- Celltech
CNTF -- Sigma-Tau
Cocaine abuse vaccine – Cantab, ImmuLogic, Scripps
coccidiomycosis vaccine -- Arizo
collagen -- Type I -- Pharming
Collagen formation inhibitors -- FibroGen
Collagen/hydroxyapatite/bone growth factor -- Aventis Pasteur, Biopharm, Orquest
collagenase -- BioSpecifics
Colorectal cancer vaccine -- Wistar Institute
Component B, Recombinant -- Serono
Connective tissue growth factor inhibitors -- FibroGen/Taisho
Contortrostatin
contraceptive vaccine -- Zonagen
Contraceptive vaccine hCG
Contraceptive vaccine male reversible -- IMMUCON
Contraceptive vaccine zona pellucida -- Zonagen
Copper-64 labelled MAb TETA-1A3 -- NCI
Coralyne
Corsevin M
C-peptide analogues -- Schwarz
CPI-1500 -- Consensus
CRF -- Neurobiological Tech.
cRGDfV pentapeptide –
CRL 1095 -- CytRx
CRL 1336 -- CytRx
CRL 1605 -- CytRx
CS-560 -- Sankyo
CSF -- ZymoGenetics
CSF-G – Hangzhou, Dong-A, Hanmi
CSF-GM – Cangene, Hunan, LG Chem
CSF-M -- Zarix
CT 1579 – Merck Frosst
CT 1786 – Merck Frosst
CT-112^ -- BTG
CTB-134L -- Xenova
CTC-111 -- Kaketsuken
CTGF -- FibroGen
CTLA4-Ig -- Bristol-Myers Squibb
CTLA4-Ig gene therapy –
CTP-37 -- AVI BioPharma
C-type natriuretic peptide -- Suntory
CVS 995 – Corvas Intl.
CX 397 – Nikko Kyodo
CY 1747 -- Epimmune
CY 1748 -- Epimmune
Cyanovirin-N
Cystic fibrosis therapy -- CBR/IVAX
CYT 351
cytokine Traps -- Regeneron
cytokines – Enzon, Cytoclonal
Cytomegalovirus glycoprotein vaccine – Chiron, Aquila Biopharmaceuticals, Aventis Pasteur, Virogenetics
Cytomegalovirus vaccine live -- Aventis Pasteur
Cytosine deaminase gene therapy -- GlaxoSmithKline
DA-3003 -- Dong-A
DAB389interleukin-6 -- Senetek
DAB389interleukin-7
DAC:GLP-2 -- ConjuChem, Inc.
Daclizumab (anti-IL2R MAb) – Protein Design Labs
DAMP^ -- Incyte Genomics
Daniplestim -- Pharmacia
darbepoetin alfa -- Amgen
DBI-3019 -- Diabetogen
DCC -- Genzyme
DDI -- Hy[illegible]

FIGURE 13J

decorin – Integra, Telios
defensins -- Large Scale Biology
DEGR-VIIa
DeImmunised antibody 3B6/22 AGEN
Deimmunised anti-cancer antibodies -- Biovation/Viragen
Dendroamide A
Dengue vaccine -- Bavarian Nordic, Merck
denileukin diftitox -- Ligand
DES-1101 -- Desmos
desirudin -- Novartis
desmopressin -- Unigene
Desmoteplase – Merck, Schering AG
Destabilase
Diabetes gene therapy – DeveloGen, Pfizer
Diabetes therapy -- Crucell
Diabetes type 1 vaccine – Diamyd Therapeutics
DiaCIM -- YM BioSciences
dialytic oligopeptides -- Research Corp
Diamyd -- Diamyd Therapeutics
DiaPep227-- Pepgen
DiavaX -- Corixa
Digoxin MAb -- Glaxo
Diphtheria tetanus pertussis-hepatitis B vaccine -- GlaxoSmithKline
DIR therapy -- Solis Therapeutics –
DNase -- Genentech
Dornase alfa -- Genentech
Dornase alfa, inhalation -- Genentech
Doxorubicin-anti-CEA MAb conjugate – Immunomedics
DP-107 -- Trimeris
drotrecogin alfa -- Eli Lilly
DTctGMCSF
DTP-polio vaccine -- Aventis Pasteur
DU 257-KM231 antibody conjugate -- Kyowa
dural graft matrix -- Integra
Duteplase – Baxter Intl.
DWP-401 -- Daewoong
DWP-404 -- Daewoong
DWP-408 -- Daewoong
Dx 88 (Epi-KAL2) -- Dyax
Dx 890 (elastin inhibitors) -- Dyax
E coli O157 vaccine -- NIH
E21-R -- BresaGen
Eastern equine encephalitis virus vaccine –
Echicetin –
Echinhibin 1 –
Echistatin -- Merck
Echitamine –
Ecromeximab – Kyowa Hakko
EC-SOD -- PPL Therapeutics
Eculizumab (5G1.1) -- Alexion
EDF -- Ajinomoto
EDN derivative -- NIH
EDNA -- NIH
Edobacomab -- XOMA
Edrecolomab -- Centocor
EF 5077
Efalizumab -- Genentech
EGF fusion toxin – Seragen, Ligand
EGF-P64k vaccine -- Center of Molecular Immunology
EL 246 -- LigoCyte
elastase inhibitor -- Synergen
elcatonin -- Therapicon
EMD 72000 -- Merck KGaA
Emdogain -- BIORA
emfilermin -- AMRAD
Emoctakin -- Novartis
enamel matrix protein -- BIORA
Endo III -- NYU
endostatin – EntreMed, Pharis
Enhancins -- Micrologix
Enlimomab -- Isis Pharm.
Enoxaparin sodium -- Pharmuka
enzyme linked antibody nutrient depletion therapy -- KS Biomedix Holdings
Eosinophil-derived neutralizing agent –
[illegible]ca

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FIGURE 13K

EP-51389 -- Asta Medica
EPH family ligands -- Regeneron
Epidermal growth factor -- Hitachi Kasei, Johnson & Johnson
Epidermal growth factor fusion toxin -- Senetek
Epidermal growth factor-genistein –
EPI-HNE-4 -- Dyax
EPI-KAL2 -- Dyax
Epoetin-alfa – Amgen, Dragon Pharmaceuticals, Nanjing Huaxin
Epratuzumab – Immunomedics
Epstein-Barr virus vaccine -- Aviron/SmithKline Beecham, Bioresearch
Eptacog alfa -- Novo Nordisk
Eptifibatide -- COR Therapeutics
erb-38 –
Erlizumab -- Genentech
erythropoietin -- Alkermes, ProLease, Dong-A, Elanex, Genetics Institute, LG Chem, Protein Sciences, Serono, Snow Brand, SRC VB VECTOR, Transkaryotic Therapies
Erythropoietin Beta -- Hoffman La Roche
Erythropoietin/Epoetin alfa -- Chugai
Escherichia coli vaccine -- North American Vaccine, SBL Vaccin, Swiss Serum and Vaccine Institute Berne
etanercept -- Immunex
examorelin – Mediolanum
Exendin 4 -- Amylin
exonuclease VII
F 105 -- Centocor
F-992 -- Fornix
Factor IX -- Alpha Therapeutics, Welfide Corp., CSL, enetics Institute/AHP, Pharmacia, PPL Therapeutics
Factor IX gene therapy -- Cell Genesys
Factor VII -- Novo Nordisk, Bayer, Baxter Intl.
Factor VIIa -- PPL Therapeutics, ZymoGenetics
Factor VIII – Bayer Genentech, Beaufour-Ipsen, CLB, Inex, Octagen, Pharmacia, Pharming
Factor VIII -- PEGylated -- Bayer
Factor VIII fragments -- Pharmacia
Factor VIII gene therapy -- Targeted Genetics
Factor VIII sucrose formulation – Bayer, Genentech
Factor VIII-2 -- Bayer
Factor VIII-3 -- Bayer
Factor Xa inhibitors – Merck, Novo Nordisk, Mochida
Factor XIII -- ZymoGenetics
Factors VIII and IX gene therapy -- Genetics Institute/Targeted Genetics
Famoxin -- Genset
Fas (delta) TM protein – LXR BioTech.
Fas TR -- Human Genome Sciences
Felvizumab -- Scotgen
FFR-VIIa -- Novo Nordisk
FG-001 – F-Gene
FG-002 – F-Gene
FG-004 – F-Gene
FG-005 – F-Gene
FGF + fibrin -- Repair
Fibrimage -- Bio-Tech. General
fibrin-binding peptides – ISIS Innovation
fibrinogen -- PPL Therapeutics, Pharming
fibroblast growth factor – Chiron, NYU, Ramot, ZymoGenetics
fibrolase conjugate -- Schering AG
Filgrastim -- Amgen
filgrastim -- PDA modified -- Xencor
FLT-3 ligand -- Immunex
FN18 CRM9 –
follistatin -- Biotech Australia, Human Therapeutics
follitropin alfa – Alkermes, ProLease, PowderJect, Serono, Akzo Nobel

FIGURE 13L

Follitropin Beta – Bayer, Organon
FP 59
FSH – Ferring
FSH + LH -- Ferring
F-spondin -- CeNeS
fusion protein delivery system -- UAB Research Foundation
fusion toxins -- Boston Life Sciences
G 5598 -- Genentech
GA-II -- Transkaryotic Therapies
Gamma-interferon analogues -- SRC VB VECTOR
Ganirelix -- Roche
gastric lipase -- Meristem
Gavilimomab –
G-CSF – Amgen, SRC VB VECTOR
GDF-1 -- CeNeS
GDF-5 -- Biopharm
GDNF (glial derived neurotrophic factor) -- Amgen
gelsolin -- Biogen
Gemtuzumab ozogamicin -- Celltech
Gene-activated epoetin-alfa -- Aventis Pharma -- Transkaryotic Therapies
Glanzmann thrombasthenia gene therapy –
Glatiramer acetate -- Yeda
glial growth factor 2 -- CeNeS
GLP-1 – Amylin, Suntory, TheraTech, Watson
GLP-1 peptide analogues – Zealand Pharaceuticals
GLP-2 – Novo Nordisk, Ontario, Inc., Suntory Limited
glucagon -- Eli Lilly, ZymoGenetics
Glucagon-like peptide-1 7-36 amide -- Suntory
Glucogen-like peptide -- Amylin
Glucocerebrosidase -- Genzyme
glutamate decarboxylase -- Genzyme Transgenics
Glycoprotein S3 -- Kureha
GM-CSF -- Immunex
GM-CSF tumour vaccine -- PowderJect
GnRH immunotherapeutic -- Protherics
Goserelin (LhRH antagonist) -- AstraZeneca
gp75 antigen -- ImClone
gp96 -- Antigenics
GPI 0100 -- Galenica
GR 4991W93 -- GlaxoSmithKline
Granulocyte colony-stimulating factor -- Dong-A
Granulocyte colony-stimulating factor conjugate
grass allergy therapy -- Dynavax
GRF1-44 -- ICN
Growth Factor – Chiron, Atrigel, Atrix, Innogenetics, ZymoGenetics, Novo
growth factor peptides – Biotherapeutics
growth hormone -- LG Chem
growth hormone, Recombinant human -- Serono
GT 4086 -- Gliatech
GW 353430 -- GlaxoSmithKline
GW-278884 -- GlaxoSmithKline
H 11 -- Viventia Biotech
H5N1 influenza A virus vaccine -- Protein Sciences
haemoglobin -- Biopure
haemoglobin 3011, Recombinant -- Baxter Healthcare
haemoglobin crosfumaril – Baxter Intl.
haemoglobin stabilized -- Ajinomoto
haemoglobin, recombinant -- Apex
HAF -- Immune Response
Hantavirus vaccine
HB 19
HBNF -- Regeneron
HCC-1 -- Pharis
hCG -- Milkhaus
hCG vaccine -- Zonagen
HE-317 -- Hollis-Eden Pharmaceuticals

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FIGURE 13M

Heat shock protein cancer and influenza vaccines -- StressGen
Helicobacter pylori vaccine -- Acambis, AstraZeneca/CSL, Chiron, Provalis
Helistat-G -- GalaGen
Hemolink -- Hemosol
hepapoietin -- Snow Brand
heparanase -- InSight
heparinase I -- Ibex
heparinase III -- Ibex
Hepatitis A vaccine -- American Biogenetic Sciences
Hepatitis A vaccine inactivated
Hepatitis A vaccine Nothav -- Chiron
Hepatitis A-hepatitis B vaccine -- GlaxoSmithKline
hepatitis B therapy -- Tripep
Hepatitis B vaccine – Amgen, Chiron SpA, Meiji Milk, NIS, Prodeva, PowderJect, Rhein Biotech
Hepatitis B vaccine recombinant -- Evans Vaccines, Epitec Combiotech, Genentech, MedImmune, Merck Sharp & Dohme, Rhein Biotech, Shantha Biotechnics, Vector, Yeda
Hepatitis B vaccine recombinant TGP 943 -- Takeda
Hepatitis C vaccine -- Bavarian Nordic, Chiron, Innogenetics Acambis,
Hepatitis D vaccine -- Chiron Vaccines
Hepatitis E vaccine recombinant -- Genelabs/GlaxoSmithKline, Novavax
hepatocyte growth factor – Panorama, Sosei
hepatocyte growth factor kringle fragments -- EntreMed
Her-2/Neu peptides -- Corixa
Herpes simplex glycoprotein DNA vaccine – Merck, Wyeth-Lederle Vaccines-Malvern, Genentech, GlaxoSmithKline, Chiron, Takeda
Herpes simplex vaccine -- Cantab Pharmaceuticals, CEL-SCI, Henderson Morley
Herpes simplex vaccine live -- ImClone Systems/Wyeth-Lederle, Aventis Pasteur
HGF derivatives -- Dompe
hIAPP vaccine -- Crucell
Hib-hepatitis B vaccine -- Aventis Pasteur
HIC 1
HIP-- Altachem
Hirudins – Biopharma, Cangene, Dongkook, Japan Energy Corporation, Pharmacia Corporation, SIR International, Sanofi-Synthelabo, Sotragene, Rhein Biotech
HIV edible vaccine -- ProdiGene
HIV gp120 vaccine – Chiron, Ajinomoto, GlaxoSmithKline, ID Vaccine, Progenics, VaxGen
HIV gp120 vaccine gene therapy –
HIV gp160 DNA vaccine – PowderJect, Aventis Pasteur, Oncogen, Hyland Immuno, Protein Sciences
HIV gp41 vaccine -- Panacos
HIV HGP-30W vaccine -- CEL-SCI
HIV immune globulin – Abbott, Chiron
HIV peptides -- American Home Products
HIV vaccine -- Applied bioTech., Axis Genetics, Biogen, Bristol-Myers Squibb, Genentech, Korea Green Cross, NIS, Oncogen, Protein Sciences Corporation, Terumo, Tonen Corporation, Wyeth-Ayerst, Wyeth-Lederle Vaccines-Malvern, Advanced BioScience Laboratories, Bavarian Nordic, Bavarian Nordic/Statens Serum Institute, GeneCure, Immune Response, Progenics, Therion Biologics, United Biomedical, Chiron
HIV vaccine vCP1433 -- Aventis Pasteur
HIV vaccine vCP1452 -- Aventis Pasteur

FIGURE 13N

HIV vaccine vCP205 -- Aventis Pasteur
HL-9 -- American BioScience
HM-9239 -- Cytran
HML-103 -- Hemosol
HML-104 -- Hemosol
HML-105 -- Hemosol
HML-109 -- Hemosol
HML-110 -- Hemosol
HML-121 -- Hemosol
hNLP -- Pharis
Hookworm vaccine
host-vector vaccines -- Henogen
HPM 1 -- Chugai
HPV vaccine -- MediGene
HSA -- Meristem
HSF -- StressGen
HSP carriers –Weizmann, Yeda, Peptor
HSPPC-70 -- Antigenics
HSPPC-96, pathogen-derived -- Antigenics
HSV 863 -- Novartis
HTLV-I DNA vaccine
HTLV-I vaccine
HTLV-II vaccine -- Access
HU 901 -- Tanox
Hu23F2G -- ICOS
HuHMFG1
HumaLYM -- Intracell
Human krebs statika -- Yamanouchi
human monoclonal antibodies -- Abgenix/Biogen, Abgenix/ Corixa, Abgenix/Immunex, Abgenix/Lexicon, Abgenix/ Pfizer, Athersys/Medarex, Biogen/MorphoSys, CAT/Searle, Centocor/Medarex, Corixa/Kirin Brewery, Corixa/Medarex, Eos BioTech./Medarex, Eos/Xenerex, Exelixis/Protein Design Labs, ImmunoGen/ Raven, Medarex/ B.Twelve, MorphoSys/ImmunoGen, XTL Biopharmaceuticals/Dyax,
Human monoclonal antibodies -- Medarex/Northwest Biotherapeutics, Medarex/Seattle Genetics
human netrin-1 -- Exelixis
human papillomavirus antibodies -- Epicyte
Human papillomavirus vaccine -- Biotech Australia, IDEC, StressGen
Human papillomavirus vaccine MEDI 501 -- MedImmune/GlaxoSmithKline
Human papillomavirus vaccine MEDI 503/MEDI 504 -- MedImmune/GlaxoSmithKline
Human papillomavirus vaccine TA-CIN – Cantab Pharmaceuticals
Human papillomavirus vaccine TA-HPV -- Cantab Pharmaceuticals
Human papillomavirus vaccine TH-GW -- Cantab/GlaxoSmithKline
human polyclonal antibodies -- Biosite/Eos BioTech./ Medarex
human type II anti factor VIII monoclonal antibodies -- ThromboGenics
humanised anti glycoprotein Ib murine monoclonal antibodies -- ThromboGenics
HumaRAD -- Intracell
HuMax EGFR -- Genmab
HuMax-CD4 -- Medarex
HuMax-IL15 -- Genmab
HYB 190 -- Hybridon
HYB 676 -- Hybridon
I-125 MAb A33 -- Celltech
Ibritumomab tiuxetan -- IDEC
IBT-9401 -- Ibex
IBT-9402 -- Ibex
IC 14 -- ICOS
Idarubicin anti-Ly-2.1 –
IDEC 114 -- IDEC
IDEC 131 -- IDEC
IDEC 152 -- IDEC
IDM 1 -- IDM

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FIGURE 13O

IDPS -- Hollis-Eden Pharmaceuticals
iduronate-2-sulfatase -- Transkaryotic Therapies
IGF/IBP-2-13 -- Pharis
IGN-101 -- Igeneon
IK HIR02 -- Iketon
IL-11 -- Genetics Institute/AHP
IL-13-PE38 -- NeoPharm
IL-17 receptor -- Immunex
IL-18BP -- Yeda
IL-1Hy1 -- Hyseq
IL-1ß -- Celltech
IL-1ß adjuvant -- Celltech
IL-2 -- Chiron
IL-2 + IL-12 -- Hoffman La-Roche
IL-6/sIL-6R fusion -- Hadasit
IL-6R derivative -- Tosoh
IL-7-Dap 389 fusion toxin -- Ligand
IL-21 -- Novo Nordisk, ZymoGenetics
IM-862 -- Cytran
IMC-1C11 -- ImClone
imiglucerase -- Genzyme
Immune globulin intravenous (human) -- Hoffman La Roche
immune privilege factor -- Proneuron
Immunocal -- Immunotec
Immunogene therapy -- Briana Bio-Tech
Immunoliposomal 5-fluorodeoxyuridine-dipalmitate --
immunosuppressant vaccine -- Aixlie
immunotoxin -- Antisoma, NIH
ImmuRAIT-Re-188 -- Immunomedics
imreg-1 -- Imreg
infertility -- Johnson & Johnson, E-TRANS
Infliximab -- Centocor
Influenza virus vaccine -- Aventis Pasteur, Protein Sciences
inhibin -- Biotech Australia, Human Therapeutics
Inhibitory G protein gene therapy
INKP-2001 -- InKine
Inolimomab -- Diaclone
insulin -- AutoImmune, Altea, Biobras, BioSante, Bio-Tech. General, Chong Kun Dang, Emisphere, Flamel, Provalis, Rhein Biotech, TranXenoGen
insulin (bovine) -- Novartis
insulin analogue -- Eli Lilly
Insulin Aspart -- Novo Nordisk
insulin detemir -- Novo Nordisk
insulin glargine -- Aventis
insulin inhaled -- Inhale Therapeutics Systems, Alkermes
insulin oral -- Inovax
insulin, AeroDose -- AeroGen
insulin, AERx -- Aradigm
insulin, BEODAS -- Elan
insulin, Biphasix -- Helix
insulin, buccal -- Generex
insulin, I2R -- Flemington
insulin, intranasal -- Bentley
insulin, oral -- Nobex, Unigene
insulin, Orasome -- Endorex
insulin, ProMaxx -- Epic
insulin, Quadrant -- Elan
insulin, recombinant -- Aventis
insulin, Spiros -- Elan
insulin, Transfersome -- IDEA
insulin, Zymo, recombinant -- Novo Nordisk
insulinotropin -- Scios
Insulysin gene therapy --
integrin antagonists -- Merck
interferon (Alpha2) -- SRC VB VECTOR, Viragen, Dong-A, Hoffman La-Roche, Genentech
interferon -- BioMedicines, Human Genome Sciences
interferon (Alfa-n3)—Interferon Sciences Intl.
interferon (Alpha), Biphasix -- Helix

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FIGURE 13P

interferon (Alpha)—Amgen, BioNative, Novartis, Genzyme Transgenics, Hayashibara, Inhale Therapeutics Systems, Medusa, Flamel, Dong-A, GeneTrol, Nastech, Shantha, Wassermann, LG Chem, Sumitomo, Aventis, Behring EGIS, Pepgen, Servier, Rhein Biotech,
interferon (Alpha2A)
interferon (Alpha2B) – Enzon, Schering-Plough, Biogen, IDEA
interferon (Alpha-N1) -- GlaxoSmithKline
interferon (beta) – Rentschler, GeneTrol, Meristem, Rhein Biotech, Toray, Yeda, Daiichi, Mochida
interferon (Beta1A) – Serono, Biogen
interferon (beta1A),inhale -- Biogen
interferon (ß1b)-- Chiron
interferon (tau)-- Pepgen
Interferon alfacon-1 -- Amgen
Interferon alpha-2a vaccine
Interferon Beta 1b -- Schering/Chiron, InterMune
Interferon Gamma -- Boehringer Ingelheim, Sheffield, Rentschler, Hayashibara
interferon receptor , Type I -- Serono
interferon(Gamma1B) -- Genentech
Interferon-alpha-2b + ribavirin – Biogen, ICN
Interferon-alpha-2b gene therapy -- Schering-Plough
Interferon-con1 gene therapy –
interleukin-1 antagonists -- Dompe
Interleukin-1 receptor antagonist -- Abbott Bioresearch, Pharmacia
Interleukin-1 receptor type I -- Immunex
interleukin-1 receptor Type II -- Immunex
Interleukin-1 trap -- Regeneron
Interleukin-1-alpha -- Immunex/Roche
interleukin-2 -- SRC VB VECTOR, Ajinomoto, Biomira, Chiron
IL-2/ diphtheria toxin -- Ligand
Interleukin-3 -- Cangene
Interleukin-4 -- Immunology Ventures, Sanofi Winthrop, Schering-Plough, Immunex/ Sanofi Winthrop, Bayer, Ono
interleukin-4 + TNF-Alpha -- NIH
interleukin-4 agonist -- Bayer
interleukin-4 fusion toxin -- Ligand
Interleukin-4 receptor – Immunex, Immun
Interleukin-6 – Ajinomoto, Cangene, Yeda, Genetics Institute, Novartis
interleukin-6 fusion protein
interleukin-6 fusion toxin – Ligand, Serono
interleukin-7 -- IC Innovations
interleukin-7 receptor -- Immunex
interleukin-8 antagonists -- Kyowa Hakko/Millennium/Pfizer
interleukin-9 antagonists -- Genaera
Interleukin-10 – DNAX, Schering-Plough
Interleukin-10 gene therapy –
interleukin-12 -- Genetics Institute, Hoffman La-Roche
interleukin-13 -- Sanofi
interleukin-13 antagonists -- AMRAD
Interleukin-13-PE38QQR
interleukin-15 -- Immunex
interleukin-16 -- Research Corp
interleukin-18 -- GlaxoSmithKline
Interleukin-18 binding protein -- Serono
Ior-P3 -- Center of Molecular Immunology
IP-10 -- NIH
IPF -- Metabolex
IR-501 -- Immune Response
ISIS 9125 -- Isis Pharmaceuticals
ISURF No. 1554 -- Millennium
ISURF No. 1866 – Iowa State Univer.
ITF-1697 -- Italfarmaco
IxC 162 -- Ixion
J 695 -- Cambridge Antibody Tech., Genetics Inst., Knoll
Jagged + FGF -- Repair
JKC-362 -- Phoenix Pharmaceuticals

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FIGURE 13Q

JTP-2942 – Japan Tobacce
Juman monoclonal antibodies -- Medarex/Raven
K02 -- Axys Pharmaceuticals
Keliximab -- IDEC
Keyhole limpet haemocyanin
KGF -- Amgen
KM 871 -- Kyowa
KPI 135 -- Scios
KPI-022 -- Scios
Kringle 5
KSB 304
KSB-201 -- KS Biomedix
L 696418 -- Merck
L 703801 -- Merck
L1 -- Acorda
L-761191 -- Merck
lactoferrin – Meristem, Pharming, Agennix
lactoferrin cardio -- Pharming
LAG-3 -- Serono
LAIT -- GEMMA
LAK cell cytotoxin -- Arizona
lamellarins -- PharmaMar/University of Malaga
laminin A peptides -- NIH
lanoteplase -- Genetics Institute
laronidase -- BioMarin
Lassa fever vaccine
LCAT -- NIH
LDP 01 -- Millennium
LDP 02 -- Millennium
Lecithinized superoxide dismutase -- Seikagaku
LeIF adjuvant -- Corixa
leishmaniasis vaccine -- Corixa
lenercept -- Hoffman La-Roche
Lenograstim – Aventis, Chugai
lepirudin -- Aventis
leptin – Amgen, IC Innovations
Leptin gene therapy -- Chiron Corporation
leptin, 2nd-generation -- Amgen
leridistim -- Pharmacia
leuprolide, ProMaxx -- Epic
leuprorelin, oral -- Unigene
LeuTech -- Papatin
LEX 032 -- SuperGen
LiDEPT -- Novartis
Lintuzumab (anti-CD33 MAb) -- Protein Design Labs
lipase -- Altus Biologics
lipid A vaccine -- EntreMed
lipid-linked anchor Tech. – ICRT, ID Biomedical
liposome-CD4 Tech. -- Sheffield
Listeria monocytogenes vaccine
LMB 1
LMB 7
LMB 9 -- Battelle Memorial Institute, NIH
LM-CD45 -- Cantab Pharmaceuticals
lovastatin -- Merck
LSA-3
LT-ß receptor – Biogen
lung cancer vaccine -- Corixa
lusupultide -- Scios
L-Vax -- AVAX
LY 355455 -- Eli Lilly
LY 366405 -- Eli Lilly
LY-355101 -- Eli Lilly
Lyme disease DNA vaccine -- Vical/Aventis Pasteur
Lyme disease vaccine -- Aquila Biopharmaceuticals, Aventis, Pasteur, Symbicom, GlaxoSmithKline, Hyland Immuno, MedImmune
Lymphocytic choriomeningitis virus vaccine
lymphoma vaccine – Biomira, Genitope
LYP18
lys plasminogen, recombinant
Lysosomal storage disease gene therapy -- Avigen
lysostaphin -- Nutrition 21
M 23 -- Gruenenthal

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FIGURE 13R

M1 monoclonal antibodies -- Acorda Therapeutics
MA 16N7C2 – Corvas Intl.
malaria vaccine -- GlaxoSmithKline, AdProTech, Antigenics, Apovia, Aventis Pasteur, Axis Genetics, Behringwerke, CDCP, Chiron Vaccines, Genzyme Transgenics, Hawaii, MedImmune, NIH, NYU, Oxxon, Roche/Saramane, Biotech Australia, Rx Tech
Malaria vaccine CDC/NIIMALVAC-1
malaria vaccine,multicomponent
mammaglobin -- Corixa
mammastatin -- Biotherapeutics
mannan-binding lectin -- NatImmu
mannan-MUC1 -- Psiron
MAP 30
Marinovir -- Phytera
MARstem -- Maret
MB-015 -- Mochida
MBP -- ImmuLogic
MCI-028 -- Mitsubishi-Tokyo
MCIF -- Human Genome Sciences
MDC -- Advanced BioScience -- Akzo Nobel, ICOS
MDX 11 -- Medarex
MDX 210 -- Medarex
MDX 22 -- Medarex
MDX 22
MDX 240 -- Medarex
MDX 33
MDX 44 -- Medarex
MDX 447 -- Medarex
MDX H210 -- Medarex
MDX RA -- Houston BioTech., Medarex
ME-104 -- Pharmexa
Measles vaccine
Mecasermin -- Cephalon/Chiron, Chiron
MEDI 488 -- MedImmune
MEDI 500
MEDI 507 -- BioTransplant
melanin concentrating hormone -- Neurocrine Biosciences
melanocortins -- OMRF
Melanoma monoclonal antibodies -- Viragen
melanoma vaccine -- GlaxoSmithKline, Akzo Nobel, Avant, Aventis Pasteur, Bavarian Nordic, Biovector, CancerVax, Genzyme Molecular Oncology, Humbolt, ImClone Systems, Memorial, NYU, Oxxon
Melanoma vaccine Magevac -- Therion
memory enhancers -- Scios
meningococcal B vaccine -- Chiron
meningococcal vaccine -- CAMR
Meningococcal vaccine group B conjugate -- North American Vaccine
Meningococcal vaccine group B recombinant -- BioChem Vaccines, Microscience
Meningococcal vaccine group Y conjugate -- North American Vaccine
Meningococcal vaccine groups A B and C conjugate -- North American Vaccine
Mepolizumab -- GlaxoSmithKline
Metastatin – EntreMed, Takeda
Met-CkB7 -- Human Genome Sciences
met-enkephalin -- TNI
METH-1 -- Human Genome Sciences
methioninase -- AntiCancer
Methionine lyase gene therapy -- AntiCancer
Met-RANTES – Genexa Biomedical, Serono
Metreleptin
Microtubule inhibitor MAb Immunogen/Abgenix
MGDF -- Kirin
MGV -- Progenics
micrin -- Endocrine
microplasmin -- ThromboGenics
MIF -- Genetics Institute

FIGURE 13S

migration inhibitory factor -- NIH
Mim CD4.1 – Xycte Therapies
mirostipen -- Human Genome Sciences
Mitumomab (BEC-2) – ImClone Systems, Merck KGaA
MK 852 -- Merck
MLN 1202 (Anti-CCR2 monoclonal antibody) – Millenium Pharmaceuticals
Mobenakin -- NIS
molgramostim -- Genetics Institute, Novartis
monoclonal antibodies -- Abgenix/Celltech, Immusol/ Medarex, Viragen/ Roslin Institute, Cambridge Antibody Tech./Elan
MAb 108 –
MAb 10D5 --
MAb 14.18-interleukin-2 immunocytokine -- Lexigen
MAb 14G2a –
MAb 15A10 –
MAb 170 -- Biomira
MAb 177Lu CC49 --
MAb 17F9
MAb 1D7
MAb 1F7 – Immune Network
MAb 1H10-doxorubicin conjugate
MAb 26-2F
MAb 2A11
MAb 2E1 -- RW Johnson
MAb 2F5
MAb 31.1 -- International BioImmune Systems
MAb 32 -- Cambridge Antibody Tech., Peptech
MAb 323A3 -- Centocor
MAb 3C5
MAb 3F12
MAb 3F8
MAb 42/6
MAb 425 -- Merck KGaA
MAb 447-52D -- Merck Sharp & Dohme
MAb 45-2D9- – haematoporphyrin conjugate
MAb 4B4
MAb 4E3-CPA conjugate -- BCM Oncologia
MAb 4E3-daunorubicin conjugate
MAb 50-6
MAb 50-61A – Institut Pasteur
MAb 5A8 -- Biogen
MAb 791T/36-methotrexate conjugate
MAb 7c11.e8
MAb 7E11 C5-selenocystamine conjugate
MAb 93KA9 -- Novartis
MAb A5B7-cisplatin conjugate -- Biodynamics Research, Pharmacia
MAb A5B7-I-131
MAb A7
MAb A717 -- Exocell
MAb A7-zinostatin conjugate
MAb ABX-RB2 -- Abgenix
MAb ACA 11
MAb AFP-I-131 – Immunomedics
MAb AP1
MAb AZ1
MAb B3-LysPE40 conjugate
MAb B4 – United Biomedical
MAb B43 Genistein-conjugate
MAb B43.13-Tc-99m -- Biomira
MAb B43-PAP conjugate
MAb B4G7-gelonin conjugate
MAb BCM 43-daunorubicin conjugate -- BCM Oncologia
MAb BIS-1
MAb BMS 181170 -- Bristol-Myers Squibb
MAb BR55-2
MAb BW494
MAb C 242-DM1 conjugate -- ImmunoGen
MAb C242-PE conjugate
MAb c30-6
MAb CA208-cytorhodin-S conjugate -- Hoechst Japan
MAb CC49 -- Enzon

FIGURE 13T

MAb ch14.18 –
MAb CH14.18-GM-CSF fusion protein -- Lexigen
MAb chCE7
MAb CI-137 -- AMRAD
MAb cisplatin conjugate
MAb CLB-CD19
MAb CLB-CD19v
MAb CLL-1 -- Peregrine
MAb CLL-1-GM-CSF conjugate
MAb CLL-1-IL-2 conjugate -- Peregrine
MAb CLN IgG -- doxorubicin conjugates
MAb conjugates – Tanox
MAb D612
MAb Dal B02
MAb DC101 -- ImClone
MAb EA 1 –
MAb EC708 -- Biovation
MAb EP-5C7 -- Protein Design Labs
MAb ERIC-1 -- ICRT
MAb F105 gene therapy
MAb FC 2.15
MAb G250 -- Centocor
MAb GA6
MAb GA733
MAb Gliomab-H -- Viventia Biotech
MAb HB2-saporin conjugate
MAb HD 37 –
MAb HD37-ricin chain-A conjugate
MAb HNK20 -- Acambis
MAb huN901-DM1 conjugate -- ImmunoGen
MAb I-131 CC49 -- Corixa
MAb ICO25
MAb ICR12-CPG2 conjugate
MAb ICR-62
MAb IRac-ricin A conjugate
MAb K1
MAb KS1-4-methotrexate conjugate
MAb L6 -- Bristol-Myers Squibb, Oncogen
MAb LiCO 16-88
MAb LL2-I-131 – Immunomedics
MAb LL2-Y-90
MAb LS2D617 -- Hybritech
MAb LYM-1-gelonin conjugate
MAb LYM-1-I-131
MAb LYM-1-Y-90
MAb LYM-2 -- Peregrine
MAb M195
MAb M195-bismuth 213 conjugate -- Protein Design Labs
MAb M195-gelonin conjugate
MAb M195-I-131
MAb M195-Y-90
MAb MA 33H1 -- Sanofi
MAb MAD11
MAb MGb2
MAb MINT5
MAb MK2-23
MAb MOC31 ETA(252-613) conjugate
MAb MOC-31-In-111
MAb MOC-31-PE conjugate
MAb MR6 –
MAb MRK-16 – Aventis Pasteur
MAb MS11G6
MAb MX-DTPA BrE-3
MAb MY9
MAb Nd2 -- Tosoh
MAb NG-1 -- Hygeia
MAb NM01 – Nissin Food
MAb OC 125
MAb OC 125-CMA conjugate
MAb OKI-1 -- Ortho-McNeil
MAb OX52 -- Bioproducts for Science
MAb PMA5
MAb PR1
MAb prost 30
MAb R-24
MAb R-24 α Human GD3 -- Celltech
MAb RFB4-ricin chain A conjugate
MAb RFT5-ricin chain A conjugate
MAb SC 1
MAb SM-3 -- ICRT
MAb SMART 1D10 -- Protein Design Labs

FIGURE 13U

MAb SMART ABL 364 -- Novartis
MAb SN6f
MAb SN6f-deglycosylated ricin A chain conjugate –
MAb SN6j
MAb SN7-ricin chain A conjugate
MAb T101-Y-90 conjugate -- Hybritech
MAb T-88 -- Chiron
MAb TB94 -- Cancer ImmunoBiology
MAb TEC 11
MAb TES-23 -- Chugai
MAb TM31 -- Avant
MAb TNT-1 -- Cambridge Antibody Tech., Peregrine
MAb TNT-3
MAb TNT-3 -- IL2 fusion protein –
MAb TP3-At-211
MAb TP3-PAP conjugate –
MAb UJ13A -- ICRT
MAb UN3
MAb ZME-018-gelonin conjugate
MAb-BC2 -- GlaxoSmithKline
MAb-DM1 conjugate -- ImmunoGen
MAb-ricin-chain-A conjugate -- XOMA
MAb-temoporfin conjugates
Monopharm C -- Viventia Biotech
monteplase -- Eisai
montirelin hydrate -- Gruenenthal
moroctocog alfa -- Genetics Institute
Moroctocog-alfa -- Pharmacia
MP 4
MP-121 -- Biopharm
MP-52 -- Biopharm
MRA -- Chugai
MS 28168 -- Mitsui Chemicals, Nihon Schering
MSH fusion toxin -- Ligand
MSI-99 -- Genaera
MT 201 -- Micromet
Muc-1 vaccine -- Corixa
mucosal tolerance -- Aberdeen
mullerian inhibiting subst
muplestim -- Genetics Institute, Novartis, DSM Anti-Infectives
murine MAb -- KS Biomedix
Mutant somatropin -- JCR Pharmaceutical
MV 833 -- Toagosei
Mycoplasma pulmonis vaccine
Mycoprex -- XOMA
myeloperoxidase -- Henogen
myostatin -- Genetics Institute
Nacolomab tafenatox -- Pharmacia
Nagrecor -- Scios
nagrestipen -- British Biotech
NAP-5 – Corvas Intl.
NAPc2 – Corvas Intl.
nartograstim -- Kyowa
Natalizumab -- Protein Design Labs
Nateplase – NIH, Nihon Schering
nateplase -- Schering AG
NBI-3001 -- Neurocrine Biosci.
NBI-5788 -- Neurocrine Biosci.
NBI-6024 -- Neurocrine Biosci.
Nef inhibitors -- BRI
Neisseria gonorrhoea vaccine -- Antex Biologics
Neomycin B-arginine conjugate
Nerelimomab -- Chiron
Nerve growth factor – Amgen – Chiron, Genentech
Nerve growth factor gene therapy
nesiritide citrate -- Scios
neuregulin-2 -- CeNeS
neurocan -- NYU
neuronal delivery system -- CAMR
Neurophil inhibitory Factor -- Corvas
Neuroprotective vaccine -- University of Auckland
neurotrophic chimaeras -- Regeneron
neurotrophic factor – NsGene, CereMedix
NeuroVax -- Immune Response
neurturin -- Genentech
neutral endopeptidase -- Genentech
NGF enhancers -- NeuroSearch

FIGURE 13V

NHL vaccine -- Large Scale Biology
NIP45 -- Boston Life Sciences
NKI-B20
NM 01 – Nissin Food
NMI-139 -- NitroMed
NMMP -- Genetics Institute
NN-2211 -- Novo Nordisk
Noggin -- Regeneron
Nonacog alfa
Norelin -- Biostar
Norwalk virus vaccine
NRLU 10 -- NeoRx
NRLU 10 PE -- NeoRx
NT-3 -- Regeneron
NT-4/5 -- Genentech
NU 3056
NU 3076
NX 1838 -- Gilead Sciences
NY ESO-1/CAG-3 antigen -- NIH
NYVAC-7 -- Aventis Pasteur
NZ-1002 -- Novazyme
obesity therapy -- Nobex
OC 10426 -- Ontogen
OC 144093 -- Ontogen
OCIF -- Sankyo
Oct-43 -- Otsuka
Odulimomab -- Immunotech
OK PSA - liposomal
OKT3-gamma-1-ala-ala
OM 991
OM 992
Omalizumab -- Genentech
oncoimmunin-L -- NIH
Oncolysin B -- ImmunoGen
Oncolysin CD6 -- ImmunoGen
Oncolysin M -- ImmunoGen
Oncolysin S -- ImmunoGen
Oncophage -- Antigenics
Oncostatin M -- Bristol-Myers Squibb
OncoVax-CL -- Jenner Biotherapies
OncoVax-P -- Jenner Biotherapies
onercept -- Yeda
onychomycosis vaccine -- Boehringer Ingelheim
opebecan -- XOMA
opioids -- Arizona
Oprelvekin -- Genetics Institute
Oregovomab -- AltaRex
Org-33408 b-- Akzo Nobel
Orolip DP -- EpiCept
oryzacystatin
OSA peptides – GenSci Regeneration
osteoblast-cadherin GF -- Pharis
Osteocalcin-thymidine kinase gene therapy
osteogenic protein -- Curis
osteopontin -- OraPharma
osteoporosis peptides – Integra, Telios
osteoprotegerin – Amgen, SnowBrand
otitis media vaccines -- Antex Biologics
ovarian cancer -- University of Alabama
OX40-IgG fusion protein -- Cantab, Xenova
P 246 -- Diatide
P 30 -- Alfacell
p1025 -- Active Biotech
P-113^ -- Demegen
P-16 peptide -- Transition Therapeutics
p43 -- Ramot
P-50 peptide -- Transition Therapeutics
p53 + RAS vaccine -- NIH, NCI
PACAP(1-27) analogue
paediatric vaccines -- Chiron
Pafase -- ICOS
PAGE-4 plasmid DNA -- IDEC
PAI-2 -- Biotech Australia, Human Therapeutics
Palifermin (keratinocyte growth factor) -- Amgen
Palivizumab -- MedImmune
PAM 4 -- Merck
pamiteplase -- Yamanouchi
pancreatin, Minitabs -- Eurand
Pangen -- Fournier

FIGURE 13W

Pantarin – Selective Genetics
Parainfluenza virus vaccine – Pharmacia, Pierre Fabre
paraoxanase -- Esperion
parathyroid hormone – Abiogen, Korea Green Cross
Parathyroid hormone (1-34) -- Chugai/Suntory
Parkinson's disease gene therapy -- Cell Genesys/ Ceregene
Parvovirus vaccine -- MedImmune
PCP-Scan – Immunomedics
PDGF -- Chiron
PDGF cocktail -- Theratechnologies
peanut allergy therapy -- Dynavax
PEG anti-ICAM MAb -- Boehringer Ingelheim
PEG asparaginase -- Enzon
PEG glucocerebrosidase
PEG hirudin – Knoll
PEG interferon-alpha-2a -- Roche
PEG interferon-alpha-2b + ribavirin – Biogen, Enzon, ICN Pharmaceuticals, Schering-Plough
PEG MAb A5B7 –
Pegacaristim – Amgen -- Kirin Brewery -- ZymoGenetics
Pegaldesleukin -- Research Corp
pegaspargase -- Enzon
pegfilgrastim -- Amgen
PEG-interferon Alpha -- Viragen
PEG-interferon Alpha 2A -- Hoffman La-Roche
PEG-interferon Alpha 2B -- Schering-Plough
PEG-r-hirudin -- Abbott
PEG-rHuMGDF -- Amgen
PEG-uricase -- Mountain View
Pegvisomant – Genentech
PEGylated proteins, PolyMASC -- Valentis
PEGylated recombinant native human leptin -- Roche
Pemtumomab
Penetratin -- Cyclacel
Pepscan – Antisoma
peptide G – Peptech, ICRT
peptide vaccine -- NIH ,NCI
Pexelizumab
pexiganan acetate -- Genaera
Pharmaprojects No. 3179 -- NYU
Pharmaprojects No. 3390 -- Ernest Orlando
Pharmaprojects No. 3417 -- Sumitomo
Pharmaprojects No. 3777 -- Acambis
Pharmaprojects No. 4209 -- XOMA
Pharmaprojects No. 4349 – Baxter Intl.
Pharmaprojects No. 4651
Pharmaprojects No. 4915 -- Avanir
Pharmaprojects No. 5156 -- Rhizogenics
Pharmaprojects No. 5200 -- Pfizer
Pharmaprojects No. 5215 -- Origene
Pharmaprojects No. 5216 -- Origene
Pharmaprojects No. 5218 -- Origene
Pharmaprojects No. 5267 -- ML Laboratories
Pharmaprojects No. 5373 -- MorphoSys
Pharmaprojects No. 5493 -- Metabolex
Pharmaprojects No. 5707 -- Genentech
Pharmaprojects No. 5728 -- Autogen
Pharmaprojects No. 5733 -- BioMarin
Pharmaprojects No. 5757 -- NIH
Pharmaprojects No. 5765 -- Gryphon
Pharmaprojects No. 5830 -- AntiCancer
Pharmaprojects No. 5839 -- Dyax
Pharmaprojects No. 5849 -- Johnson & Johnson
Pharmaprojects No. 5860 -- Mitsubishi-Tokyo
Pharmaprojects No. 5869 – Oxford GlycoSciences
Pharmaprojects No. 5883 -- Asahi Brewery
Pharmaprojects No. 5947 -- StressGen

FIGURE 13X

Pharmaprojects No. 5961 -- Theratechnologies
Pharmaprojects No. 5962 -- NIH
Pharmaprojects No. 5966 -- NIH
Pharmaprojects No. 5994 -- Pharming
Pharmaprojects No. 5995 -- Pharming
Pharmaprojects No. 6023 -- IMMUCON
Pharmaprojects No. 6063 -- Cytoclonal
Pharmaprojects No. 6073 -- SIDDCO
Pharmaprojects No. 6115 -- Genzyme
Pharmaprojects No. 6227 -- NIH
Pharmaprojects No. 6230 -- NIH
Pharmaprojects No. 6236 -- NIH
Pharmaprojects No. 6243 -- NIH
Pharmaprojects No. 6244 -- NIH
Pharmaprojects No. 6281 -- Senetek
Pharmaprojects No. 6365 -- NIH
Pharmaprojects No. 6368 -- NIH
Pharmaprojects No. 6373 -- NIH
Pharmaprojects No. 6408 – Pan Pacific
Pharmaprojects No. 6410 -- Athersys
Pharmaprojects No. 6421 – Oxford GlycoSciences
Pharmaprojects No. 6522 -- Maxygen
Pharmaprojects No. 6523 -- Pharis
Pharmaprojects No. 6538 -- Maxygen
Pharmaprojects No. 6554 -- APALEXO
Pharmaprojects No. 6560 -- Ardana
Pharmaprojects No. 6562 -- Bayer
Pharmaprojects No. 6569 -- Eos
Phenoxazine
Phenylase -- Ibex
Pigment epithelium derived factor –
plasminogen activator inhibitor-1, recombinant -- DuPont Pharmaceuticals
Plasminogen activators -- Abbott Laboratories, American Home Products, Boehringer Mannheim, Chiron Corporation, DuPont Pharmaceuticals, Eli Lilly, Shionogi, Genentech, Genetics Institute, GlaxoSmithKline, Hemispherx Biopharma, Merck & Co, Novartis, Pharmacia Corporation, Wakamoto, Yeda
plasminogen-related peptides -- Bio-Tech. General/MGH
platelet factor 4 -- RepliGen
Platelet-derived growth factor – Amgen -- ZymoGenetics
plusonermin-- Hayashibara
PMD-2850 -- Protherics
Pneumococcal vaccine -- Antex Biologics, Aventis Pasteur
Pneumococcal vaccine intranasal – BioChem Vaccines/Biovector
PR1A3
PR-39
pralmorelin -- Kaken
Pretarget-Lymphoma -- NeoRx
Priliximab -- Centocor
PRO 140 -- Progenics
PRO 2000 -- Procept
PRO 367 -- Progenics
PRO 542 -- Progenics
pro-Apo A-I -- Esperion
prolactin -- Genzyme
Prosaptide TX14(A) -- Bio-Tech. General
prostate cancer antbodies – Immunex, UroCor
prostate cancer antibody therapy -- Genentech/UroGenesys, Genotherapeutics
prostate cancer immunotherapeutics -- The PSMA Development Company
prostate cancer vaccine -- Aventis Pasteur, Zonagen, Corixa, Dendreon, Jenner Biotherapies, Therion Biologics

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FIGURE 13Y

prostate-specific antigen -- EntreMed
protein A -- RepliGen
protein adhesives -- Enzon
protein C – Baxter Intl., PPL Therapeutics, ZymoGenetics
protein C activator – Gilead Sciences
protein kinase R antags -- NIH
protirelin -- Takeda
protocadherin 2 -- Caprion
Pro-urokinase – Abbott, Bristol-Myers Squibb, Dainippon, Tosoh -- Welfide
P-selectin glycoprotein ligand-1 -- Genetics Institute
pseudomonal infections -- InterMune
Pseudomonas vaccine -- Cytovax
PSGL-Ig -- American Home Products
PSP-94 -- Procyon
PTH 1-34 -- Nobex
Quilimmune-M -- Antigenics
R 744 -- Roche
R 101933
R 125224 -- Sankyo
RA therapy -- Cardion
Rabies vaccine recombinant -- Aventis Pasteur, BioChem Vaccines, Kaketsuken Pharmaceuticals
RadioTheraCIM -- YM BioSciences
Ramot project No. 1315 -- Ramot
Ramot project No. K-734A -- Ramot
Ramot project No. K-734B -- Ramot
Ranibizumab (Anti-VEGF fragment) -- Genentech
RANK -- Immunex
ranpirnase -- Alfacell
ranpirnase-anti-CD22 MAb -- Alfacell
RANTES inhibitor -- Milan
RAPID drug delivery systems -- ARIAD
rasburicase -- Sanofi
rBPI-21, topical -- XOMA
RC 529 -- Corixa
rCFTR -- Genzyme Transgenics
RD 62198
rDnase -- Genentech
RDP-58 -- SangStat
RecepTox-Fce -- Keryx
RecepTox-GnRH – Keryx, MTR Technologies
RecepTox-MBP – Keryx, MTR Technologies
recFSH -- Akzo Nobel, Organon
REGA 3G12
Regavirumab -- Teijin
relaxin -- Connetics Corp
Renal cancer vaccine -- Macropharm
repifermin -- Human Genome Sciences
Respiratory syncytial virus PFP-2 vaccine -- Wyeth-Lederle
Respiratory syncytial virus vaccine – GlaxoSmithKline, Pharmacia, Pierre Fabre
Respiratory syncytial virus vaccine inactivated
Respiratory syncytial virus-parainfluenza virus vaccine -- Aventis Pasteur, Pharmacia
Reteplase -- Boehringer Mannheim, Hoffman La-Roche
Retropep -- Retroscreen
RFB4 (dsFv) PE38
RFI 641 -- American Home Products
RFTS -- UAB Research Foundation
RG 12986 -- Aventis Pasteur
RG 83852 -- Aventis Pasteur
RG-1059 -- RepliGen
rGCR -- NIH
rGLP-1 -- Restoragen
rGRF -- Restoragen
rh Insulin – Eli Lilly
RHAMM targeting peptides -- Cangene
rHb1.1 – Baxter Intl.
rhCC10 -- Claragen
rhCG -- Serono
Rheumatoid arthritis gene therapy

FIGURE 13Z

Rheumatoid arthritis vaccine -- Veterans Affairs Medical Center
rhLH -- Serono
Ribozyme gene therapy -- Genset
Rickettsial vaccine recombinant
RIGScan CR -- Neoprobe
RIP-3 -- Rigel
Rituximab -- Genentech
RK-0202 -- RxKinetix
RLT peptide -- Esperion
rM/NEI -- IVAX
rmCRP -- Immtech
RN-1001 -- Renovo
RN-3 -- Renovo
RNAse conjugate -- Immunomedics
RO 631908 -- Roche
Rotavirus vaccine -- Merck
RP 431 -- DuPont Pharmaceuticals
RP-128 -- Resolution
RPE65 gene therapy –
RPR 110173 -- Aventis Pasteur
RPR 115135 -- Aventis Pasteur
RPR 116258A -- Aventis Pasteur
rPSGL-Ig -- American Home Products
r-SPC surfactant -- Byk Gulden
RSV antibody -- Medimmune
Ruplizumab -- Biogen
rV-HER-2/neu -- Therion Biologics
SA 1042 -- Sankyo
sacrosidase – Orphan Medical
Sant 7
Sargramostim -- Immunex
saruplase -- Gruenenthal
Satumomab -- Cytogen
SB 1 -- COR Therapeutics
SB 207448 -- GlaxoSmithKline
SB 208651 -- GlaxoSmithKline
SB 240683 -- GlaxoSmithKline
SB 249415 -- GlaxoSmithKline
SB 249417 -- GlaxoSmithKline
SB 6 -- COR Therapeutics
SB RA 31012 –
SC 56929 -- Pharmacia
SCA binding proteins – Curis, Enzon
scFv(14E1)-ETA Berlex Laboratories, Schering AG
ScFv(FRP5)-ETA –
ScFv6C6-PE40 –
SCH 55700 -- Celltech
Schistosomiasis vaccine -- Glaxo Wellcome/Medeva, Brazil
SCPF -- Advanced Tissue Sciences
scuPA-suPAR complex -- Hadasit
SD-9427 -- Pharmacia
SDF-1 -- Ono
SDZ 215918 -- Novartis
SDZ 280125 -- Novartis
SDZ 89104 -- Novartis
SDZ ABL 364 -- Novartis
SDZ MMA 383 -- Novartis
Secretin – Ferring, Repligen
serine protease inhibs -- Pharis
sermorelin acetate -- Serono
SERP-1 -- Viron
sertenef -- Dainippon
serum albumin, Recombinant human -- Aventis Behring
serum-derived factor -- Hadasit
Sevirumab -- Novartis
SGN 14 -- Seatle Genetics
SGN 15 -- Seatle Genetics
SGN 17/19 – Seatle Genetics
SGN 30 -- Seatle Genetics
SGN-10 -- Seatle Genetics
SGN-11 -- Seatle Genetics
SH 306 -- DuPont Pharmaceuticals
Shanvac-B -- Shantha
Shigella flexneri vaccine – Avant, Acambis, Novavax
Shigella sonnei vaccine –
sICAM-1 -- Boehringer Ingelheim
Silteplase -- Genzyme
SIV vaccine – Endocon, Institut Pasteur
SK 896 -- Sanwa Kagaku Kenkyusho

FIGURE 13AA

SK-827 -- Sanwa Kagaku Kenkyusho
Skeletex -- CellFactors
SKF 106160 -- GlaxoSmithKline
S-nitroso-AR545C –
SNTP -- Active Biotech
somatomedin-1 – GroPep, Mitsubishi-Tokyo, NIH
somatomedin-1 carrier protein -- Insmed
somatostatin -- Ferring
Somatotropin/
Human Growth Hormone -- Bio-Tech. General, Eli Lilly
somatropin -- Bio-Tech. General, Alkermes, ProLease, Aventis Behring, Biovector, Cangene, Dong-A, Eli Lilly, Emisphere, Enact, Genentech, Genzyme Transgenics, Grandis/InfiMed, CSL, InfiMed, MacroMed, Novartis, Novo Nordisk, Pharmacia Serono, TranXenoGen
somatropin derivative -- Schering AG
somatropin, AIR -- Eli Lilly
Somatropin, inhaled -- Eli Lilly/Alkermes
somatropin, Kabi -- Pharmacia
somatropin, Orasome -- Novo Nordisk
Sonermin -- Dainippon Pharmaceutical
SP(V5.2)C -- Supertek
SPf66
sphingomyelinase -- Genzyme
SR 29001 -- Sanofi
SR 41476 -- Sanofi
SR-29001 -- Sanofi
SS1(dsFV)-PE38 -- NeoPharm
ß2 microglobulin -- Avidex
ß2-microglobulin fusion proteins -- NIH
ß-amyloid peptides -- CeNeS
ß-defensin -- Pharis
Staphylococcus aureus infections -- Inhibitex/ZLB
Staphylococcus aureus vaccine conjugate -- Nabi
Staphylococcus therapy -- Tripep
Staphylokinase – Biovation, Prothera, Thrombogenetics
Streptococcal A vaccine -- M6 Pharmaceuticals, North American Vaccine
Streptococcal B vaccine -- Microscience
Streptococcal B vaccine recombinant -- Biochem Vaccines
Streptococcus pyogenes vaccine
STRL-33 -- NIH
Subalin -- SRC VB VECTOR
SUIS -- United Biomedical
SUIS-LHRH -- United Biomedical
SUN-E3001 -- Suntory
super high affinity monoclonal antibodies -- YM BioSciences
Superoxide dismutase – Chiron, Enzon, Ube Industries, Bio-Tech, Yeda
superoxide dismutase-2 -- OXIS
suppressin -- UAB Research Foundation
SY-161-P5 -- ThromboGenics
SY-162 -- ThromboGenics
Systemic lupus erythematosus vaccine -- MedClone/VivoRx
T cell receptor peptides -- Xoma
T cell receptor peptide vaccine
T4N5 liposomes -- AGI Dermatics
TACI, soluble -- ZymoGenetics
targeted apoptosis -- Antisoma
tasonermin -- Boehringer Ingelheim
TASP
TASP-V
Tat peptide analogues -- NIH
TBP I -- Yeda
TBP II
TBV25H -- NIH
Tc 99m ior cea1 -- Center of Molecular Immunology
Tc 99m P 748 -- Diatide
Tc 99m votumumab -- Intracell
Tc 99m rh-Annexin V – Theseus Imaging
teceleukin -- Biogen

FIGURE 13BB

tenecteplase -- Genentech
Teriparatide -- Armour Pharmaceuticals, Asahi Kasei, Eli Lilly
terlipressin -- Ferring
testisin -- AMRAD
Tetrafibricin -- Roche
TFPI -- EntreMed
tgD-IL-2 -- Takeda
TGF-Alpha -- ZymoGenetics
TGF-ß -- Kolon
TGF-ß2 -- Insmed
TGF-ß3 -- OSI
Thalassaemia gene therapy -- Crucell
TheraCIM-h-R3 -- Center of Molecular Immunology, YM BioSciences
Theradigm-HBV -- Epimmune
Theradigm-HPV -- Epimmune
Theradigm-malaria -- Epimmune
Theradigm-melanoma -- Epimmune
TheraFab – Antisoma
ThGRF 1-29 -- Theratechnologies
ThGRF 1-44 -- Theratechnologies
Thrombin receptor activating peptide -- Abbott
thrombomodulin – Iowa, Novocastra
Thrombopoietin -- Dragon Pharmaceuticals, Genentech
thrombopoietin, Pliva -- Receptron
Thrombospondin 2 –
thrombostatin -- Thromgen
thymalfasin -- SciClone
thymocartin – Gedeon Richter
thymosin Alpha1 -- NIH
thyroid stimulating hormone -- Genzyme
tICAM-1 -- Bayer
Tick anticoagulant peptide -- Merck
TIF -- Xoma
Tifacogin – Chiron, NIS, Pharmacia
Tissue factor -- Genentech
Tissue factor pathway inhibitor
TJN-135 -- Tsumura
TM 27 -- Avant
TM 29 -- Avant
TMC-151 – Tanabe Seiyaku
TNF tumour necrosis factor -- Asahi Kasei
TNF Alpha -- CytImmune
TNF antibody -- Johnson & Johnson
TNF binding protein -- Amgen
TNF degradation product -- Oncotech
TNF receptor -- Immunex
TNF receptor 1, soluble -- Amgen
TNF Tumour necrosis factor-alpha -- Asahi Kasei, Genetech, Mochida
TNF-Alpha inhibitor -- Tripep
TNFR:Fc gene therapy – Targeted Genetics
TNF-SAM2
ToleriMab -- Innogenetics
Toxoplasma gondii vaccine -- GlaxoSmithKline
TP 9201 -- Telios
TP10 -- Avant
TP20 -- Avant
tPA -- Centocor
trafermin -- Scios
TRAIL/Apo2L -- Immunex
TRAIL-R1 MAb – Cambridge Antibody Technologies
transferrin-binding proteins -- CAMR
Transforming growth factor-beta-1 -- Genentech
transport protein -- Genesis
Trastuzumab -- Genetech
TRH -- Ferring
Triabin -- Schering AG
Triconal
Triflavin
troponin I -- Boston Life Sciences
TRP-2^ -- NIH
trypsin inhibitor -- Mochida
TSP-1 gene therapy –
TT-232
TTS-CD2 -- Active Biotech

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FIGURE 13CC

Tuberculosis vaccine -- Aventis Pasteur, Genesis
Tumor Targeted Superantigens -- Active Biotech -- Pharmacia
tumour vaccines -- PhotoCure
tumour-activated prodrug antibody conjugates -- Millennium/ImmunoGen
tumstatin -- ILEX
Tuvirumab -- Novartis
TV-4710 – Teva
TWEAK receptor -- Immunex
TXU-PAP
TY-10721 – TOA Eiyo
Type I diabetes vaccine -- Research Corp
Typhoid vaccine CVD 908
U 143677 -- Pharmacia
U 81749 -- Pharmacia
UA 1248 -- Arizona
UGIF -- Sheffield
UIC 2
UK 101
UK-279276 – Corvas Intl.
urodilatin -- Pharis
urofollitrophin -- Serono
Urokinase -- Abbott
uteroferrin-- Pepgen
V 20 -- GLYCODesign
V2 vasopressin receptor gene therapy
vaccines -- Active Biotech
Varicella zoster glycoprotein vaccine -- Research Corporation Technologies
Varicella zoster virus vaccine live -- Cantab Pharmaceuticals
Vascular endothelial growth factor – Genentech, University of California
Vascular endothelial growth factors – R&D Systems
vascular targeting agents -- Peregrine
vasopermeation enhancement agents -- Peregrine
vasostatin -- NIH
VCL -- Bio-Tech. General
VEGF – Genentech, Scios
VEGF inhibitor -- Chugai
VEGF-2 -- Human Genome Sciences
VEGF-Trap -- Regeneron
viscumin, recombinant -- Madaus
Vitaxin
Vitrase -- ISTA Pharmaceuticals
West Nile virus vaccine -- Bavarian Nordic
WP 652
WT1 vaccine -- Corixa
WX-293 -- Wilex BioTech.
WX-360 -- Wilex BioTech.
WX-UK1 -- Wilex BioTech.
XMP-500 -- XOMA
XomaZyme-791 -- XOMA
XTL 001 -- XTL Biopharmaceuticals
XTL 002 -- XTL Biopharmaceuticals
yeast delivery system -- GlobeImmune
Yersinia pestis vaccine
YIGSR-Stealth -- Johnson & Johnson
Yissum Project No. D-0460 -- Yissum
YM 207 -- Yamanouchi
YM 337 -- Protein Design Labs
Yttrium-90 labelled biotin
Yttrium-90-labeled anti-CEA MAb T84.66 –
ZD 0490 – AstraZeneca
ziconotide -- Elan
ZK 157138 -- Berlex Laboratories
Zolimomab aritox
Zorcell -- Immune Response
ZRXL peptides -- Novartis

FIGURE 14A

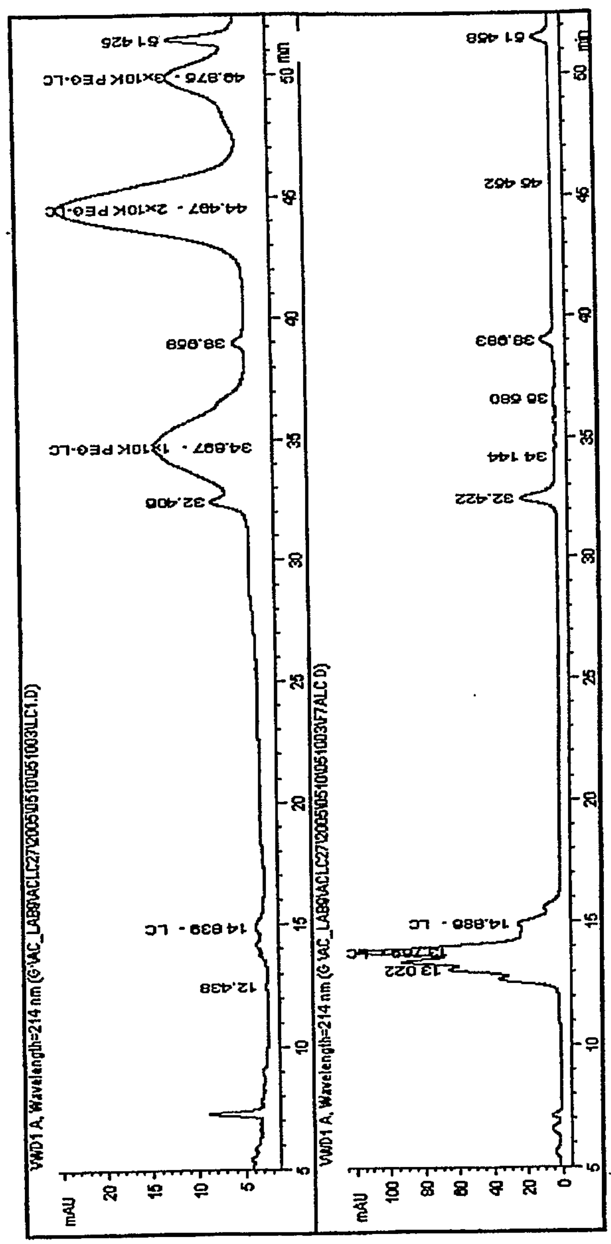

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FIGURE 14B

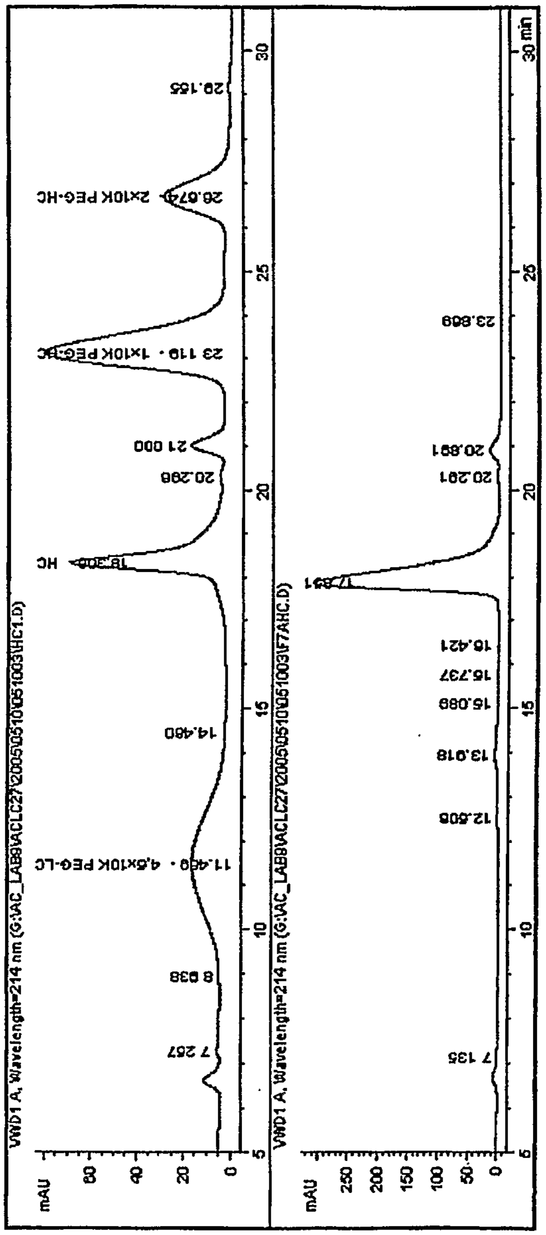
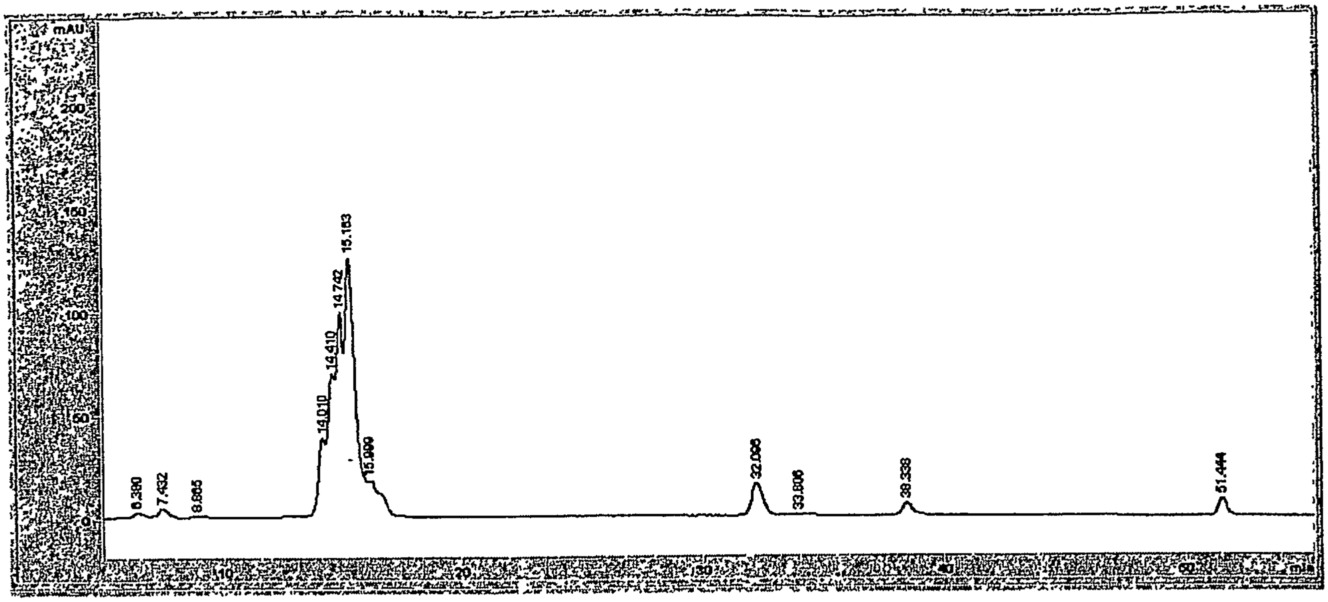


FIGURE 15A



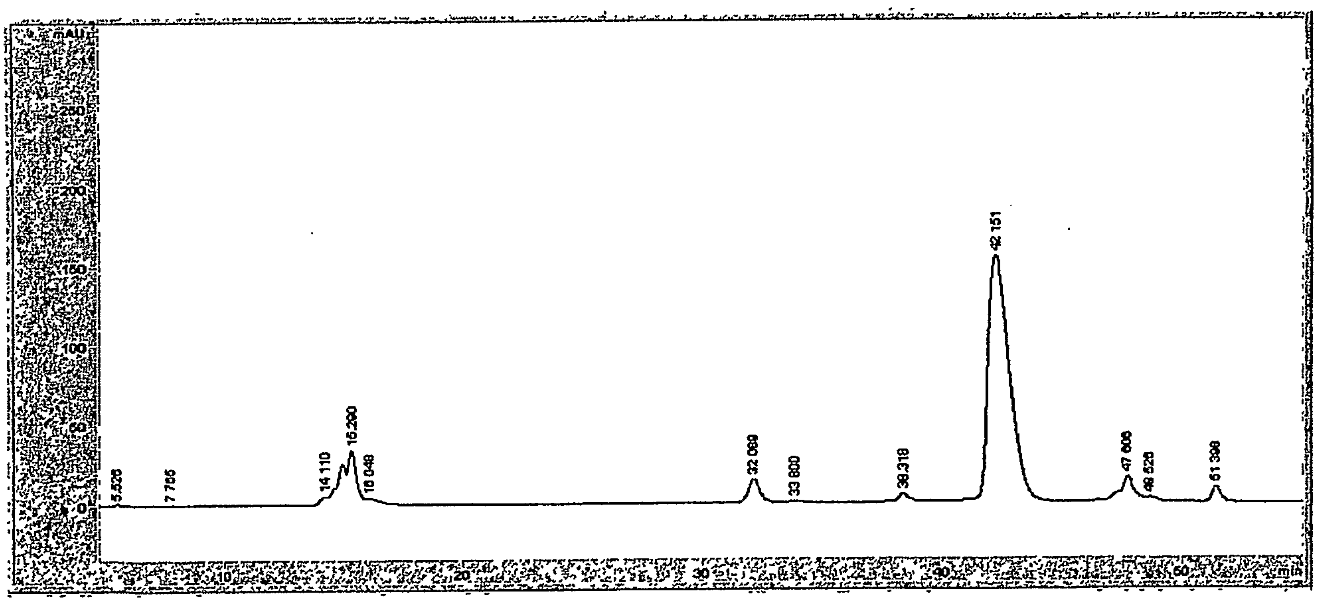


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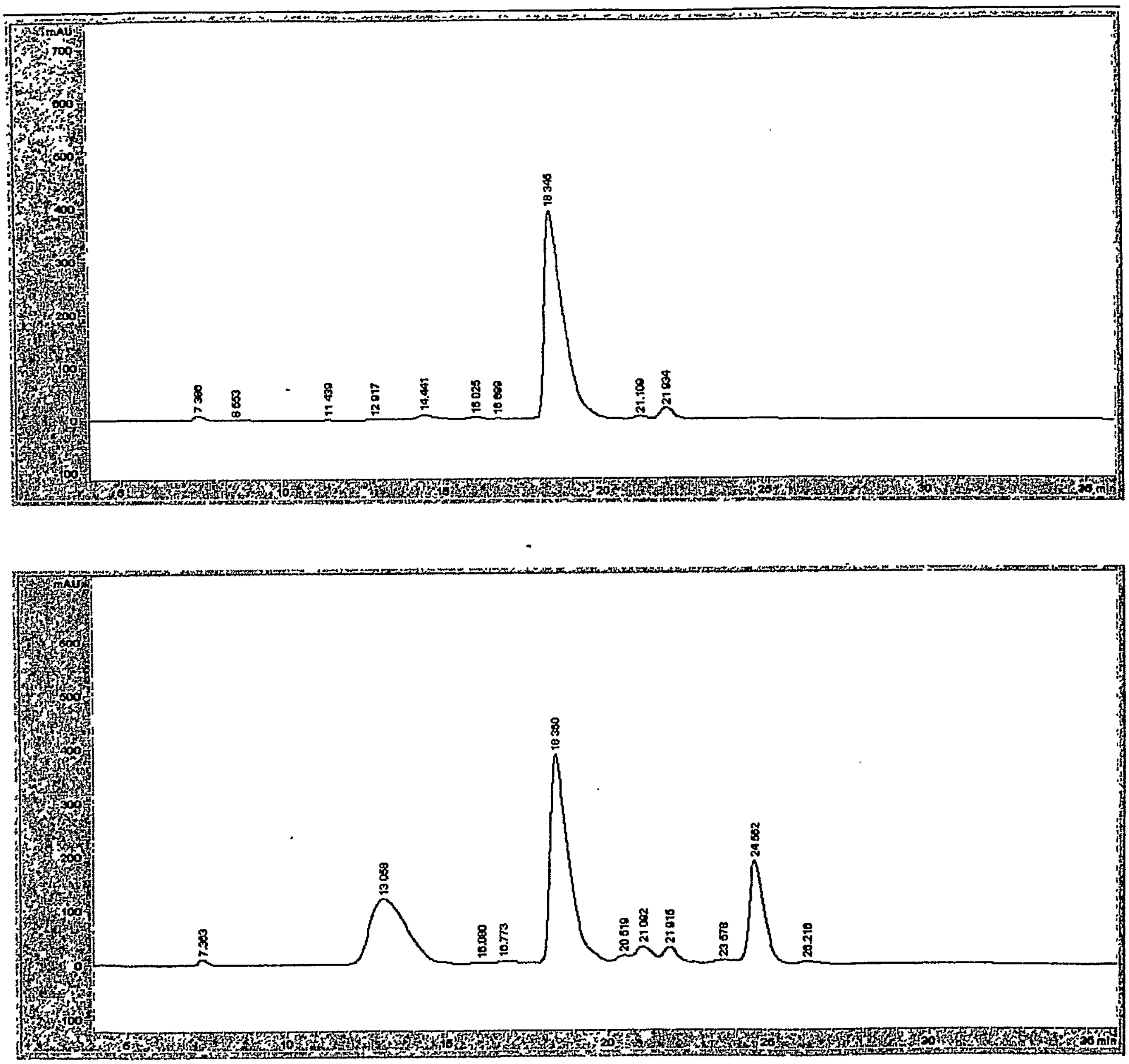
FIGURE 15B



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